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A HALF-CENTURY OF RESEARCH ON MICROALGAE-BACTERIA FOR WASTEWATER TREATMENT

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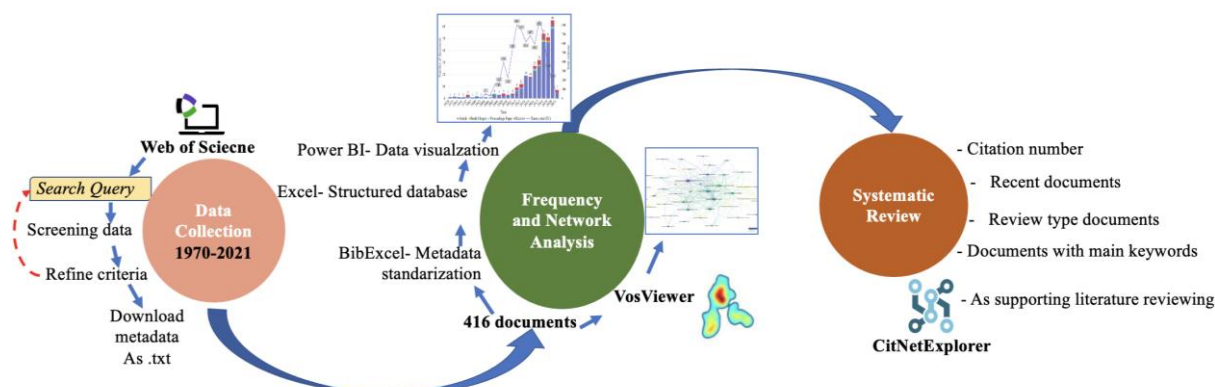
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Abstract:

The use of microalgae-bacteria consortia (MBC) for wastewater treatment have received increasing attention because of its high capacity for nutrients and organic matter removal, using sunlight as an energy source, through self-sufficient oxygen production provided by photosynthesis. This review addresses the research hotspots and main challenges of this emerging technology, using bibliometric analysis. The evolution of MBC-related literature from 1970 to 2021 was analyzed, identifying of the main research topics explored, based on the frequency of the keywords. This document develops the following topics identified as research hotspots: **1)** The factors that influence the complex interactions between microalgae and bacteria (e.g., pH, incident light, dissolved oxygen), and the removal mechanisms of nutrients and organic matter, which are still relevant issues still requiring significant research efforts. **2)** The new enhanced pathways of total nitrogen and phosphorus removal, such as short cut nitrogen removal, alammox, and phosphorus luxury uptake, which have been explored during the last decade. **3)** The application of mathematical models to represent the behavior of MBC, as a tool for prediction and optimization of MBC systems. And **4)** The biomass retention technologies, such as biofilms, membrane bioreactors, and granular sludge; that allows the uncoupling of HRT from SRT, a condition required for high-rate wastewater treatment.

Graphical Abstract:



Keywords: Bibliometric analysis, decentralized system, microalgae-bacteria consortia, nutrients removal, wastewater treatment

1. INTRODUCTION

Nowadays, about 80% of the produced wastewater (WW) worldwide is discharged into the environment without prior treatment. In developing countries, this number reached up to 90% in 2010 [1,2]. This situation promotes social, environmental, and public health problems, mainly in rural or small communities [3,4]. In this context, the most widely used wastewater treatment process worldwide is the conventional activated sludge (CAS) [5,6]. However, the application of this technology in small communities could be limited due to the high energy demand associated with mechanical aeration [7,8]. In fact, mechanical aeration in CAS represents between 50 and 90% of total energy demand [9–11]. In addition, CAS entails a high CO₂ footprint and a waste of nutrients (i.e., nitrogen and phosphorous).

Microalgae-bacteria consortia (MBC) based treatment systems are considered a promising bioremediation platform due to the self-sufficient production of oxygen, provided by microalgae in the presence of light (photooxygenation), which reduces the operational costs associated with mechanical aeration [12,13]. This O₂ production supplies the requirements of heterotrophic bacteria (HB) for the oxidation of chemical oxygen demand (COD) and of nitrifying bacteria (NB) for the oxidation of ammonium into NO₃⁻ during the nitrification process. Meanwhile, CO₂ is produced by HB, which is subsequently used as a carbon source by microalgae and NB [14–16]. Moreover, MBC systems have shown a high capacity for nutrients removal. This is the result of the ability of microalgae to assimilate nutrients, and the mechanisms associated with nitrifying bacteria [17–20].

Use of MBC has the potential to transform the way wastewater treatment is conventionally conceived, by using sunlight as an energy source driving the biological process. Thus, operational costs are reduced, and potentially valuable products could be obtained from wastewater through the further biomass transformation in biofertilizer, biofuels, and/or bioproducts [21]. In addition, when compared to CAS, a reduction of CO₂ emissions is expected, mediated by the capture of CO₂ by microalgae [22–24]. Additionally, the treated effluent from MBC is expected to meet the wastewater discharge and reuse standards [25,26].

However, despite the advances and the knowledge acquired in the past 70 years in the application of MBC, there is still a lack of fundamental knowledge regarding the complex interactions between microorganisms, and substantial challenges associated with biomass separation. In fact, the poor settling properties of biomass is one of the main limitations of the application of MBC [27]. Although some researchers have addressed this limitation by applying different biomass retention technologies (e.g. biomass immobilization, membrane bioreactors, etc.), more research is still required [28].

On the other hand, several works published in the last decade dealing with MBC are devoted to treating different types of wastewaters, and literature on MBC keeps on growing steadily. In this context, a comprehensive analysis of the existing literature seems necessary to understand the current state of this research field, its advances, challenges, and opportunities.

To go beyond a standard state of the art review, the objective of this paper is to quantify and analyze reported research using bibliometric analysis. This tool has proven useful to analyze a scalable body of research literature, providing means for assessing and evaluating output of scientists, collaboration between groups and identification of key or emerging topics [29,30]. Then, this work is intended to provide a deep analysis of available literature dealing with the application of MBC to wastewater treatment, offering an in-depth synthesis of the most challenging topics.

2. MATERIAL AND METHODS

Bibliometric analysis enables the identification of the structure, characteristics, and trends for a given topic by exploring different bibliographic characteristics such as the number and type of documents, countries, institutions, keywords, citation analysis and using descriptive, impact, or/and relational indicators [31,32]. The input data to perform bibliometric analysis in scientific fields is typically provided by referential databases such as Web of Science (WoS). Based on our experience [33,34], a bibliometric analysis procedure was established, which is described in this section, along with the software tools used to generate diverse bibliometric results and trends.

2.1 Data collection

Data were collected up to December 2021, using the WoS Core Collection database, including the SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH, BKCI-S, BKCI-SSH, ESCI, CCR-EXPANDED, IC. WoS database was selected since it is considered as a source of high-quality scientific data and frequently used for bibliometric analysis [35–37].

2.2 Search strategy

The bibliometric analysis involves a search query (SQ) that includes the use of keywords, boolean operators, and wildcards. In this work, the SQ used was “**TOPIC: (((*alga*\$(bacteri* OR activated sludge) AND (consort* OR symbio*) and (*wastewater OR sewage)))) NOT TITLE: (((*hydrogen OR “fuel cell” OR *ferment* OR *ethan*))**”. Keywords involved in the SQ were searched in the fields title, abstract and keywords of the documents, using WoS database. In addition, the following document types were excluded from the analysis: early access, meeting abstract and editorial material, since they are not expected to provide a significant contribution to our research. The SQ used in this work was developed in order to provide documents related to MBC for wastewater treatment, excluding those that use the algal-based consortia for other purposes, such as the production of biofuels or biochemicals.

2.3 Frequency and network analysis and systematic review

After performing the SQ in WoS, metadata of all documents were obtained in the so-called local database. The following indicators and software were used to perform the bibliometric analysis.

- Bibexcel [38] was used for metadata standardization and the estimation of the local h-index impact indicator for countries [39,40]. The h-index was proposed by Hirsch [41] to characterize the scientific output of a researcher or country. It is defined as follows: “a scientist has index h if h of his/her N papers have at least h citations each and the other (Np - h) papers have $\leq h$ citations each”. Thereby, the importance and broad impact of scientists’ or country’s cumulative research contribution could be addressed by using both the quantity (total documents) and quality (citation number) of publications. Moreover, the h-index was normalized (NH) as the ratio between the h-index and total documents. The NH ranges from 0 to 1 to describe the influence and impact of publications of each country. A value of 0 means that the country has an h-index equal to 0, i.e., none of its documents were cited. A value of 1 entail that the country has reached the maximum h-index.
- Microsoft Excel v.16.40 and PowerBI® v.2.87.762.0 were used to create a structured database for visualization purposes [42,43]. Descriptive indicators used in this work were the annual distribution of documents and citations, the performance of most productive countries, and normalized h-index (SH).

- VOSviewer v.1.6.16. was used to study the collaborative network of countries, and to elaborate the co-occurrence map of all keywords (author-defined keywords and keyword plus), using a frequency of 3 co-occurrences and a thesaurus file. The co-occurrence keyword map was performed using overlay visualization, with a weight or size of the circle representing the co-occurrence value and the color representing the average publication year. The clusterization in VOSviewer was performed by similarity [44].
- CitNetExplorer was used to analyse cited documents in the database and to facilitate systematic literature by identifying publications that are closely connected in terms of citation [45].

Based on the results provided by bibliometric data, publication trends, keywords co-occurrence evolution, and collaboration network were determined. Then, technical aspects related with the application of MBC for wastewater treatment were discussed.

3. BIBLIOMETRIC ANALYSIS

3.1 Annual distribution of documents and citations

In total, 416 documents were found in the local database, including reports from 1970 to 2021. The documents considered were articles (350 documents, corresponding to 84%), review articles (48 documents, corresponding to 12%), and proceeding papers or book chapters (18 documents, corresponding to 4 %). The distribution of documents by year is shown in Figure 1, along with their number of citations.

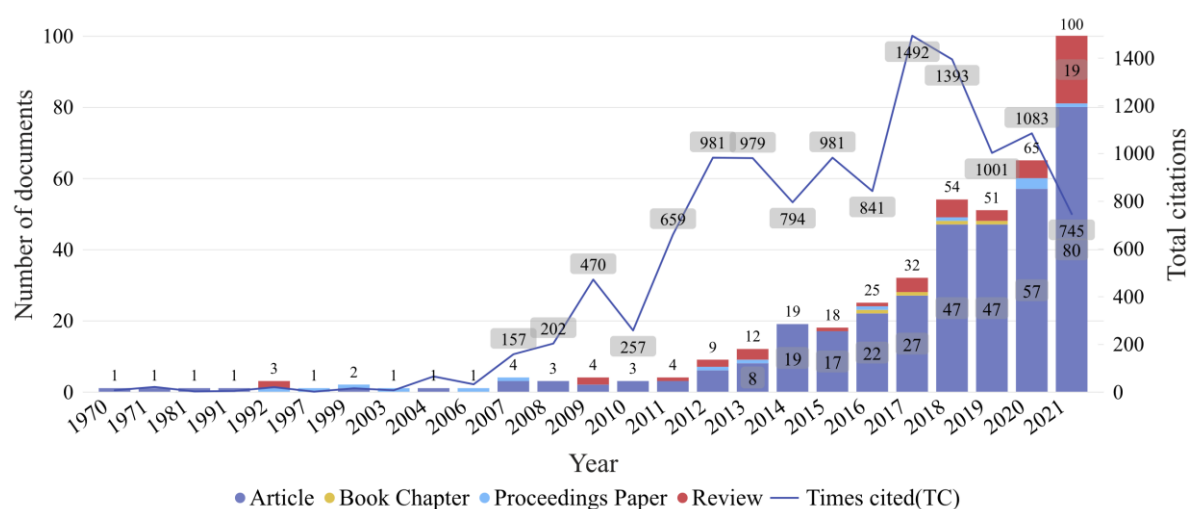


Figure 1. Yearly distribution of type of documents and number of citations dealing with the application of MBC for wastewater treatment from 1970 to 2021

The data presented in Figure 1 shows a growing interest in the study of MBC during the last decade. Three stages can be identified for bibliographic production and knowledge development. The first stage ranges from 1970 to 2012 and involves 40 documents (10 %) with an average annual frequency below 5 documents. Interestingly, the bibliographic production during this period is not continuous, as can be observed in Figure 1, which may be attributed to the use of microalgae in other research contexts (e.g., bioproducts or biofuels). The second stage ranges from 2013 to 2017 and involves 106 documents (25 %). Despite the shorter period

involved, the second stage includes 2.7 times more documents than stage 1. The cumulative average growth rate is about 5 documents per year, only including a slight drop in 2015. The third stage ranges from 2018 to 2021. It involves 270 documents, corresponding to 65% of all the documents, showing that the use of MBC for wastewater treatment is becoming an attractive technology, attracting growing research efforts. The average frequency of review production increased from an average of 0.16 documents per year in the first stage to 1.8 in the second stage, and to 8 in the third stage. This increase may be interpreted as the result of a rapid build-up of knowledge on the subject during recent years. Similar trends were obtained by Qi et al. [46], highlighting the growing interest in applying MBC for wastewater treatment.

Several relevant reports that have contributed to the development of MBC-based wastewater treatment were identified by CitNetExplorer, based on their high citation score. The document with the highest citation score was published in 2006 by Muñoz and Guieysse [16], with 875 total citations, with 121 of them from documents included in the local database used in this bibliometric analysis. Review article from Muñoz and Guieysse [16] has contributed to conceptualize the potential use of MBC and photooxygenation for the removal of different pollutants during wastewater treatment. In addition, the application of different operational conditions and bioreactor configurations were discussed, as well as the promising use of microalgae as a feedstock to produce high-value chemicals or biogas. The second most cited article, with 240 citations (75 of them by documents included in the local database), was published in 2012 by Su et al. [47], who investigated the synergistic cooperation between algae (wastewater-born) and activated sludge, at different algae/sludge inoculation ratios. This paper revealed that an algae/sludge ratio of 5:1 showed the highest ammonium and phosphorus removal, assimilation being the main removal mechanisms of NH_4^+ in the consortia.

Several documents published during the third stage also deserve to be highlighted, based on the relevant number of citations received in such a short period of time. The most cited article is the one from Sepehri et al. [48] published in 2020, with 221 citations, reporting that aeration in the conventional nitrification process could be replaced by photo-oxygenation, achieving 100% of NH_4^+ removal at the ratio of 10% of microalgae (*Chlorella Vulgaris*) and 90% of nitrifying activated sludge (w/w) at a low C/N ratio. On the other hand, Su et al. (2012), reported that the removal mechanism of NH_4^+ was mainly governed by biomass assimilation. Therefore, several factors, such as wastewater characteristics and dominant microbial species, could change the mechanisms of removal as well as the optimal ratio of algae and bacteria.

The second most cited article during third stage is the one published by Mohsenpour et al. [49], with 117 citations. The authors reviewed the incorporation and importance of microalgae in wastewater treatment, as well as the cellular and metabolic mechanisms involved in nitrogen, phosphorus and organic matter removal. The third most cited article in the third stage was the one from Renuka et al. [50], with 115 citations. It reviews the potential of microalgae as biofertilizers in agriculture and how cultivation using waste-substrates is emerging as an economically viable strategy. The article also highlighted that future efforts should be directed to the validation and commercialization of the biofertilizer. The fourth most cited article was the one from Ji et al. [51], with 88 citations. It studies the capacity of coexisting *Chlorella Vulgaris* and *Bacillus Licheniformis* for the removal of nitrogen, phosphorus and COD. Finally, the fifth most cited article corresponds to the published by Zhang et al. [52], with 87 citations, who explored the advances in granulation and how this process is enhanced by MBC (i.e., providing granular stability, improving nutrient removal efficiency, and biomass settleability). These reports and their high citation level provide an insight into the relevance and importance of topics involving MBC systems behavior, such as microalgae/bacteria ratio, granulation, biomass recovery, and valorization.

A comprehensive analysis of the citations showed low values until 2007, when a gradual rise

was observed. The highest number of citations occurred in 2017 and 2012, with 1492 and 981, respectively. Unusually, a topic that has been researched for more than 50 years presents the largest number of citations in the last 10 years. The observed citations evolution could be considered as an indicator of two phenomena: First, the popularization of MBC for wastewater treatment by the end of the first stage, resulting in fast consolidation of knowledge [27,53], and second, the application, improvement, and valorization of biomass from MBC, especially during the third stage [50,54,55].

3.2 Most productive and collaborative countries

A total of 60 countries have contributed to the available literature in MBC for wastewater treatment, between 1970 and 2021. Figure 2 (A) shows the performance of the 10 countries showing the highest productivities from 1970 to 2021. Information on total documents (TD), total citations (TC), international collaboration (Col), h-index and normalized h-index (NH) is provided. The countries that have the largest contribution are China, USA, and Spain, with 118 (28,3%), 51 (12,3%), and 48 (11,5%) documents, respectively.

However, when considering the most productive countries during 2020 and 2021, few emerging countries appear: Italy with 10 documents during 2020, India with 14 documents during 2021 and Mexico with 8 documents during 2021. Despite China ranking 1st place in bibliographic production from 1970 to 2021, it has the lowest NH (0.26), suggesting a modest impact. On the other hand, HS values for Spain, USA, Italy, and India are 0.45, 0.46, 0.54, and 0.49 respectively. Meanwhile, countries with lower productivity, such as Malaysia, Mexico, Netherlands, and South Korea, have the most significant impact (high NH), probably due to their collaboration with the most productive countries.

The topic of MBC for wastewater treatment is being developed in a collaborative network, as it is shown in **Figure 2 (B)**, where the 25 most productive countries were grouped into 8 clusters, using VOSviewer. All clusters show interconnection, except for Germany, which appears isolated. The closer each cluster and country are located to each other, the stronger their collaboration relatedness. China shows the highest international cooperation, showing interaction with 14 of the 25 countries included in the collaboration network (**Figure 2 A, col**). USA and China have the highest joint collaboration, with 12 documents, followed by Spain-Italy (6) China-Singapore (6), Canada-Sweden (5), and Japan-Mexico (5). A strong collaboration between USA and countries with low productivity but high impact (high NS) is observed (for example with The Netherlands and South Korea) (**Figure 2 B**). Although an interconnected network can be identified, collaboration could be considered low, if the total number of documents and countries contributing to MBC research is considered. Improving the collaboration between countries with different levels of experience and productivity would undoubtedly promote the transfer of knowledge and technology, positively impacting and enhancing research.

The analysis also reveals another interesting aspect of the processes for wastewater treatment involving microalgae: the topics on which each country has focused. China has studied lipid and biomass production using aerobic granular biomass [56–58], biofilm bioreactors [59,60], and membrane bioreactors [61–63]. China was the first country to conduct studies combining membranes and MBC for wastewater treatment, addressing membrane performance and fouling [63,64]. USA has researched the effect of parameters such as biomass inoculation, carbon-nitrogen ratio (C/N), and incident light on MBC-based wastewater treatment [48,65,66]. Italy has focused on photosynthesis and respiration models [67,68], respirometry tests to evaluate the activity of consortia [69,70], mixotrophic metabolism of microalgae [71], and studies involving sewage and outdoor pilot raceways [72,73].

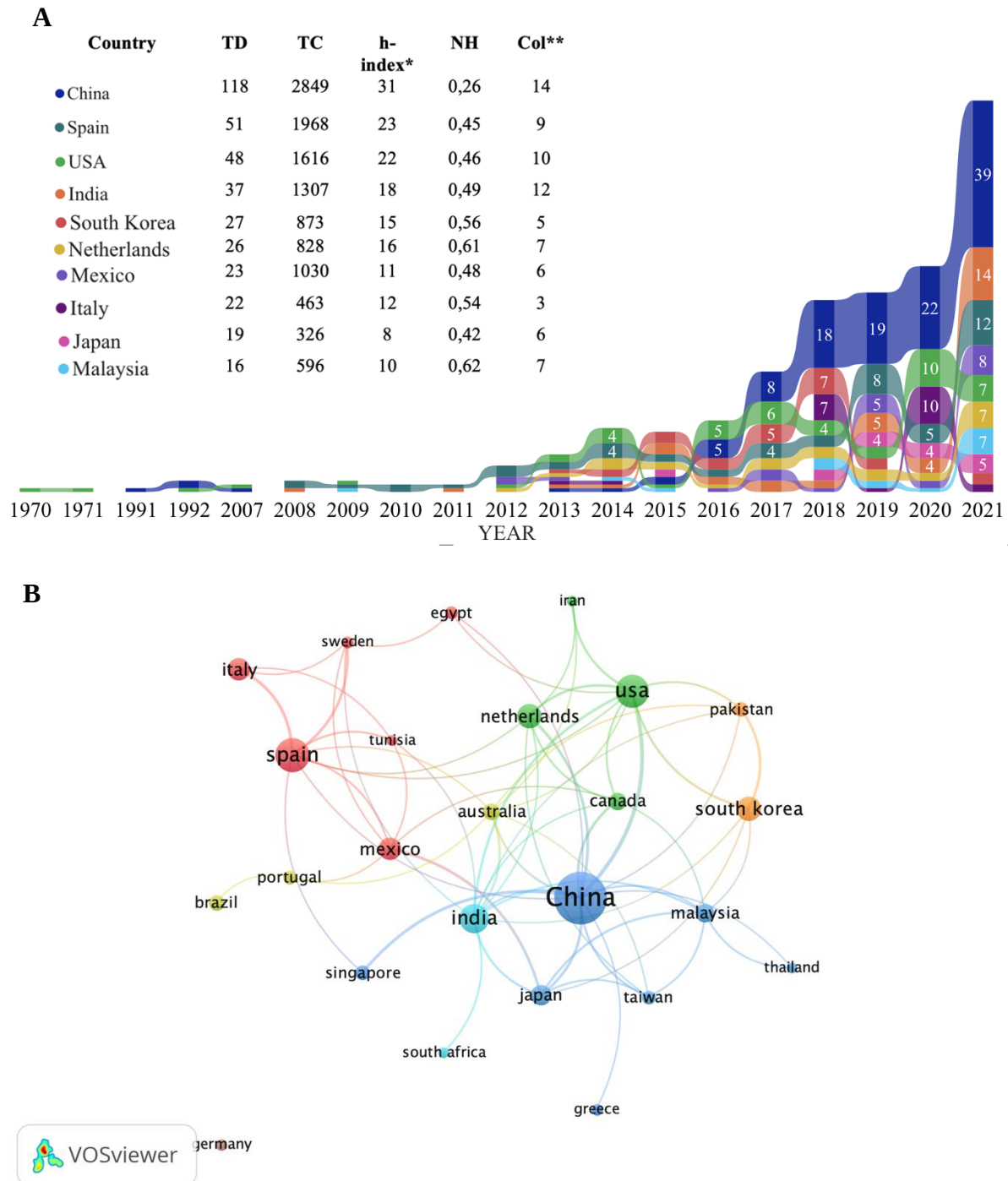


Figure 2. Contribution of countries on MBC field from 1970 to 2021 (A) Information of the 10 countries showing the highest bibliographic productivity: Total documents (TD), total citations (TC), h-index, NH and international collaboration (Col). * Local h-index calculated using BibExcel. **International collaboration determined using VOSviewer (B) Collaboration network between countries presented using VOSviewer. Each circle represents one country, and its size is proportional to the number of published documents

(TD). Clusters are identified by different colors.

Data shows a low presence of Latin American countries on the revised scientific literature dealing with MBC for wastewater treatment. This region contributed with only 12% of the bibliographic production (50 documents) and included countries such as Brazil, Nicaragua, Bolivia, Chile, Honduras, Argentina, and Colombia. Interestingly, Latin America is a region where the application of MBC for wastewater treatment may represent an alternative to cope with the lack of access to proper sanitation in medium and small communities. One could expect the rapid development of research in this geographic area in the future, which may be enhanced by the opportunities provided by collaborative work.

3.3 Evolution of MBC research for wastewater treatment

The evolution of the research on MBC for wastewater treatment was addressed using all keywords co-occurrence network map over time. CitNetExplorer was used to analyze the cited documents. A total of 1932 keywords were found, which were reduced to 49, using a thesaurus and a minimum of co-occurrence of 5 times, as is shown in **Figure 3**. CitNetExplorer showed how studies on MBC for wastewater treatment dated back more than 60 years. The keywords “*activated sludge*”, “*carbon and nutrient removal*”, “*high-rate algal ponds (HRAPs)*”, and “*photo-oxygenation*” are strongly related to the oldest author cited. Oswald and his research group proposed the use of HRAPs already in 1957 [74]. They proposed for the first time the symbiosis of MBC as a platform for wastewater treatment, reporting COD, nitrogen, and phosphorus removals in the range of 70-90%, 50-90%, and 60-99%, respectively [75–81].

Between 2014 and 2017, the use of MBC was intensified. The emergence of keywords such as “*piggery wastewater*”, “*municipal wastewater*”, and “*domestic wastewater*” shows a speciation of the scientific community towards different technologies depending on the wastewater [82–84]. In the last decade, some strategies have been adopted to address the low settleability of microalgae. This has been identified as one of the most significant limitations of MBC, resulting in unclarified effluents and the need for high HRT and footprints [14,85]. The keywords “*bioflocculation*”, “*biofilm photobioreactor*” [86,87], “*aerobic granular sludge*” [58,88]), “*immobilization*” [89,90], “*membrane photobioreactor*” and “*EPS*” [62,63] refer to reactor configuration, biomass recovery and biomass structure to improve the “*settleability*” of biomass.

synthesise the state of the art along these four cross-cutting research streams.

4. INTERACTIONS BETWEEN MICROALGAE AND BACTERIA

Interactions between microalgae and bacteria have been explored for over 70 years [12,81] when terms such as "*photo-oxygenation*" were introduced. As shown in **Figure 4**, the microbial metabolisms coexisting in MBC are photoautotrophic, heterotrophic, and autotrophic [104,105]. The most characteristic interaction between algae and bacteria is the O_2/CO_2 transfer. Microalgae produce O_2 (photo-oxygenation) as a consequence of the photosynthetic process, which is then used by HB and NB to oxidize organic matter and ammoniacal nitrogen, respectively [13,16]. The photo-oxygenation drives, therefore, the availability in the culture broth of O_2 , a key electron acceptor during nitrogen and organic matter oxidation [12,106]. Moreover, HB releases CO_2 that is later used by microalgae and AB as a carbon source [14].

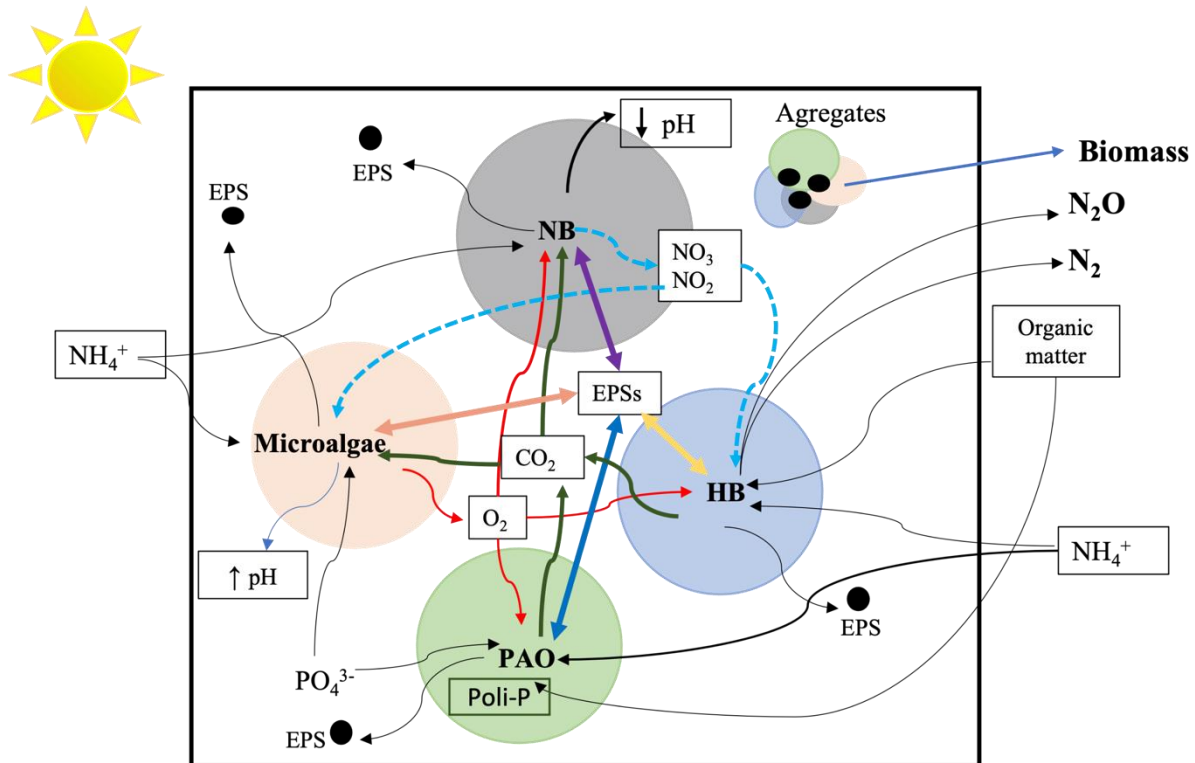


Figure 4. Interactions within MBC. Adapted from Yang et al. [28]

In addition to the above-mentioned O_2/CO_2 exchange, soluble metabolite exchange also plays an essential role in the removal of pollutants. Nitrogen in domestic wastewater is typically found as urea, and ammonia (NH_4^+ and NH_3) and is further oxidized to NO_2^- and NO_3^- by NB. The nitrogen forms that microalgae can assimilate are NO_3^- , NO_2^- and NH_4^+ . They are catalysed by different enzymatic transformations from NO_3^- to NH_4^+ and finally to the organic nitrogen present in amino acids. The enzymes that act in the nitrogen assimilation process in microalgae are nitrate and nitrite reductase to transform NO_3^- into NO_2^- and into NH_4^+ , respectively, and glutamine synthetase to convert NH_4^+ into organic nitrogen [20,107,108]. Nitrate and nitrite reductase enzymes are inhibited at high ammonium concentrations since microalgae prefer NH_4^+ as a nitrogen source due to the lower energy required for its assimilation [53,109]. On the other hand, the activity of denitrifying heterotrophic bacteria has been evidenced under conditions of low O_2 concentration (typically during the dark phase and for low oxygen transfer

rate) [60,93,99,100,110]. Phosphorus (PO_4^{3-}) can also be removed from wastewater by both microalgae and bacteria through assimilation in the biomass [111,112].

Cell to cell communication has also been reported as a relevant mechanism for microbial interaction. It involves the intracellular exchange of small signal compounds between bacteria, and the response of microalgae to these signals have been recently also reported in MBC cultures [113]. This cell to cell communication mechanism is normally referred to as quorum sensing, and it is density-cell dependent [114,115]. In MBC, the quorum sensing signals play a crucial role since it can determine the stimulation or inhibition of microorganisms present in the consortium [116–119], the formation of biofilm or aggregates [120–122], the processing and transformation of nutrients [123,124], and lipid synthesis [125,126]. The secretion by microalgae or bacteria of toxic substances, such as algicidal or bactericidal compounds, could be non-beneficial for the culture [21,91,127,128]. For instance, it has been reported that a concentration of $19,45 \mu\text{g L}^{-1}$ of urocanic acid, excreted by bacteria, negatively affects the activity of microalgae *P. Globosa*, *P. Donghaiense*, *H. Akashiwo*, and *S. Costatum* [59]. The secretion of algicidal substances by bacteria are triggered by bacterial cell concentration rather than the detection of microalgae by bacteria [114]. Some reported algicidal compounds are amylase, 2-undecen-1'-yl-4-quinolone, 2-undecyl-4-quinolone indole-3-acetic acid and octadecenoic acid [128]. Moreover, bactericidal substances such as chlorellin, secreted by microalgae, exhibit an inhibitory effect on both Gram-positive and Gram-negative bacteria [21,116,129]. Others compounds reported as relevant for cell to cell communication are indole-3-acetic acid (phytohormone) that stimulate the metabolism of microalgae, enhancing the interaction between microalgae and bacteria [118,130,131]. Likewise, vitamins produced by bacteria such as thiamine pyrophosphate and 4-amino-5-hydroxymethyl-2-methylpyrimidine were found to promote the growth, lipid content, and glucose uptake of *A. Protothecoides* [132]. Moreover, vitamins secreted by bacteria such as B12, B1 and B7 are required for supported the growth of microalgae in the MBC culture [133,134].

Excretion of extracellular polymeric substances (EPS) has been reported as a relevant factor triggering floc formation in MBC, acting as a biological glue, promoting the agglomeration of microalgae and bacteria, and thus improving its settleability [92,135]. The secretion of EPS have also been correlated with quorum sensing acyl-homoserine lactones secreted by bacteria [120,136]. For example, Zhou et al. 2017 [121] reported self-aggregation of *Chlorophyta sp* in $200 \mu\text{m}$ bioflocs by secreting 460–1000 kDa of aromatic proteins, upon interacting with N-acyl-homoserine lactones.

Gene transfer is another mechanism of microbial that may play a role in the development of MBC. It implies the change on gene expression of individual organisms during the interaction in the consortia. These changes may play a role in microalgae adaptation to changes on environmental conditions [133,137].

Thus, studying the interactions between microalgae and bacteria is of utmost importance to explore and investigate how to better manage their dynamics, and enhance the pollutant removal mechanisms efficiency as well as the quality and quantity of sludge generated during wastewater treatment [118,138].

5. NUTRIENT REMOVAL MECHANISMS

The keywords “*Phosphorus removal*” and “*nitrogen removal*” were identified as relevant terms when analyzing available literature on the application of MBC municipal wastewater treatment (**Figure 4**). MBC can provide a simultaneous removal of carbon, phosphorus, and nitrogen in a single-step process. Many authors agree that the removal of nutrients provided by

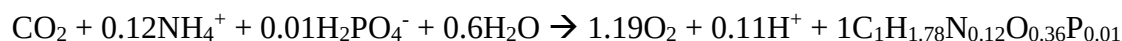
MBC are higher than those provided by algae and bacteria alone [139,140]. Nutrient removal has been approached in different ways. Several mechanisms for the removal of phosphorus and nitrogen in MBC have been identified. Such variety can be considered as an advantage since it provides options or alternatives. However, it also represents a challenge since it involves complexity for understanding, controlling, or simulating the process.

5.1 Phosphorus removal

The mechanisms reported for phosphorus removal in MBC are precipitation and assimilation by microalgal and/or bacterial biomass. Phosphorus precipitation is an indirect removal mechanism induced at high pH values (pH>8), where the pH increase is mainly attributed to the microalgal activity. In this context, some authors have highlighted the importance of Ca^{++} availability in phosphorus precipitation, Mg^{++} playing a minor role in the precipitation process [75,141].

Phosphorus removal through assimilation in microalgae could be limited by unbalanced nitrogen/phosphorus (N/P) ratio in wastewaters. **Equation 1** shows the stoichiometric growth of microalgae on NH_4^+ , PO_4^- and CO_2 , assuming a biomass composition of $\text{C}_{1.78}\text{H}_{1.78}\text{N}_{0.12}\text{O}_{0.36}\text{P}_{0.01}$ [142]. Then, it could be expected that wastewater with an N/P ratio close to 12 would be required for N and P removal without nutrient limitation.

Nonetheless, some authors have observed an optimal N/P ratio between 5 and 12 for nutrient removal, using municipal wastewater [76,142]. The range of N/P can vary since the optimal N/P ratio in microalgae biomass is flexible, according to the cell stoichiometry and strongly depends both on the species of microalgae and their capacity of performing luxury phosphorus uptake [63,143,144]. When the N/P ratio is lower than the optimal maximum for biomass production, phosphorus removal decreases due to nitrogen limitation [79,145,146]. Thus, in MBC systems where the nutrients are shared (Figure 5), since many nitrogen removal mechanisms can take place (e.g., nitrification or ammonia volatilization), it is of utmost importance to guarantee the availability of NH_4^+ for efficient PO_4^- removal [147].



Equation 1

Concerning the microalgae luxury uptake of phosphorus, it has been reported that during light conditions and a preceding phosphorus starvation, the accumulation of larger polyphosphate granules is triggered [148]. The maximum content of phosphorus in microalgae can be up to 3.8 mg P mg^{-1} of dry biomass, about four times the maximum standard content under balanced microalgal metabolism (Equation 1) [149,150]. The most common microalgae performing luxury uptake in waste stabilization ponds and HRAPs have been reported to be *Chlamydomonas/Cryptomonas*, *Micractinium/Microcystis*, and *Scenedesmus* [149,151]. However, the luxury phosphorus uptake mechanisms must be further explored, considering their complexity.

Another mechanism for PO_4^{3-} removal is the one involving the activity of polyphosphate-accumulating organisms (PAOs), which has not been widely explored in MBC systems (Figure 5.). The accumulation of phosphate through PAOs is achieved in two environments, one anaerobic followed by one aerobic/anoxic. The PAOs uptake and store organic carbon such as acetate and propionate anaerobically as polyhydroxyalkanoate (PHA), with polyphosphate as the energy source [152]. In anaerobic environments, organic carbon decreases while soluble PO_4^{3-} increases. In the subsequent aerobic/anoxic environment PAOs oxidize the stored PHA, which is used as energy for growth and restore internal polyphosphate. During aerobic/anoxic environment, the PO_4^{3-} decreases; through this cyclical process, more P is stored than was released, resulting in the net removal of phosphorus [153,154]. In MBC systems, Huang et al.

[155] studied the effect of algae growth on aerobic granulation and nutrient removal. They observed inhibition of PAOs due to free nitrous acid (FNA) accumulation ($6.3\text{--}9.8 \times 10^{-3} \text{ mg HNO}_2\text{-N L}^{-1}$), obtaining a total phosphorus removal not higher than 44%. Furthermore, PAOs activity may be limited due to the high NO_3^- concentrations typically found in MBC systems, which hamper the occurrence of anaerobic conditions affecting the release of PO_4^{3-} and subsequently the uptake of phosphate [156].

In addition, recent works have reported indicating that a low carbon/nitrogen ratio will provide low organic carbon to HB growth, including PAOs [58,157]. Then, just under some specific environmental conditions, PAOs may play a significant role in phosphorus removal when the MBC system is used [158]. Due to the complex environmental condition where PAOs act as the main actor in phosphorus removal, some researchers have found them playing a role as phosphorus assimilation and growth promoters in microalgae [139,159].

5.2 Nitrogen Removal

During the first and second stages illustrated in **Figure 1**, the most common nitrogen removal mechanisms reported were the assimilation by microalgal biomass, nitrification/denitrification, and NH_3 volatilization. García et al. [160] reported 73% of total nitrogen (TN) removal at HRT of 10 days and 57% at HRT of 4 days. In both systems, ammonia stripping was the dominant mechanism over microalgal assimilation on TN removal. Moreover, denitrification was not considered, and nitrification was identified mainly in the HRAP working at lower HRT (4 days), contributing to NH_4^+ removal. Nurdogan and Oswald (1995) reported 90% of TN removal, which was mainly attributed to volatilization, especially in summer, when pH reached a maximum value of 11. The predominance of ammonia stripping mechanism has generally been found in HRAP systems as a result of high pH, usually higher than 9, due to the activity of microalgae [161,162]. In addition, some authors have highlighted that controlling pH at a lower level by supplying CO_2 can reduce the ammonia stripping in HRAP by favoring the ammonium form. Moreover, reducing inorganic carbon limitation, enhanced nitrogen recovery [26,163–165]. However, Ruas et al. [26] evaluated the domestic wastewater treatment with the supply of synthetic gas mixture (30% v/v CO_2), and the pH was maintained in the system between 6.8 to 7.7. The TN removal obtained was below 40% due to high nitrification activity combined with a lower pH, preventing ammonia stripping. Similarly, Karya et al. [13] reported low TN removal when treating synthetic wastewater using activated sludge and *Scenedesmus Quadricauda* as inoculum (pH-controlled at 7.5). Under the tested conditions, nitrification was the most relevant ammonia removal mechanism (81% - 85%), contributing to the low the TN removal obtained.

Furthermore, pH has indirect effects on the availability of toxic compounds such as free ammonia (FA) and FNA, where the role of pH indirectly determines the magnitude of inhibition by establishing the degree of dissociation of non-toxic NH_4^+ to toxic NH_3 . For example, Valdivelu et al. [168] found that *Nitrobacter* was more sensitive than *Nitrosomonas* to FA. For *Nitrobacter*, the growth was completely inhibited at above $6.0 \text{ mg NH}_3\text{-N L}^{-1}$, and the capacity to oxidize NO_2^- to NO_3^- was reduced by 12 % at concentration of $6.0\text{--}10 \text{ mg NH}_3\text{-N L}^{-1}$; meanwhile, *Nitrosomonas* did not experience any inhibition at FA up to $16 \text{ mg NH}_3\text{-N L}^{-1}$. Indeed, nitrification process (NO_2^- to NO_3^-) seems to be the more sensitive to ammonia concentrations. Meanwhile, FA can also affect photosynthetic activity. For instance, a FA concentrations of $73 \text{ mg NH}_3\text{-N L}^{-1}$ in MBC with dominant species of *Chlorella sp.* and *Scenedesmus sp.* can generate 50% inhibition of the photosynthetic activity and photooxygenation [169,170].

Likewise, FNA has a greater impact on *Nitrobacter* by completely stopping the growth process at concentrations of 0.02 mg HNO₂-N L⁻¹ and inhibiting the energy yielding metabolisms at concentrations greater than 0.024 mg HNO₂-N L⁻¹ [168]. Huang et al. [155] found that microalgae inhibit NB growth by competing for NH₄⁺ substrate, which led to an accumulation of NO₂ and FNA (>28 mg HNO₂-N L⁻¹) in the effluent, thus limiting the nitrification and denitrification process. This resulted in low levels of TN removal (40.7% - 45.4%).

The key role of the influent alkalinity regulation to avoid strong inorganic carbon limitation has been recently highlighted in MBC systems [171]. CO₂ is the substrate of the Calvin cycle and strongly determines the microalgal growth rate [48,166,167]. It also determines growth of NB, since it is also their carbon source. Inorganic carbon limitation can affect nitrogen removal and is more often observed for wastewaters characterized by low biodegradable organic carbon contents (e.g., digestate). A low biodegradability will limit HB growth, reducing the CO₂ production [48,157]. Conditions of very low alkalinity results in intense competition for inorganic carbon between algae and nitrifiers (limited growth of photoautotrophic microorganisms), incomplete nitrification (high level of ammoniacal nitrogen in the effluent) and favorable conditions for N₂O production and emissions [171,174–176]. N₂O is a very potent greenhouse gas (GHG), with a global warming potential about 298 times higher than that of CO₂ [173].

A well-known mechanism for the oxidation of ammonium nitrogen by bacteria is complete or partial nitrification (nitritation), which requires further denitrification to achieve an effective total nitrogen removal. New processes have been identified in MBC when nitrogen removal is considered, starting by the end of the second stage (**Figure 1**). Keywords such as “*nitrification-denitrification*” and “*shortcut nitrogen removal*” were the most representative in the same time-lapse. Wang et al. [100] developed a novel shortcut nitrogen removal in a photo-sequencing batch reactor (PSBR) using an HRT of 4 days. Researchers obtained a TN removal of 90% without external aeration. As a result of the high concentrations of FA (2.4 mg L⁻¹) and FNA (0.013 mg L⁻¹) and low O₂, 80% of the TN removal was achieved by nitritation/denitritation with the addition of acetate as an external carbon source (ECS) during the dark period. Other researchers have used methanol as ECS for the same purpose [95]. In addition, Alcantará et al. [177] reported a new anoxic-aerobic photobioreactor process with biomass recycling that provided TN removal in the range 68-79%, when treating synthetic wastewater at an HRT of 2 days. In this system, the authors found that NH₄⁺ nitritation was dominant in the aerobic photobioreactor reaching concentrations up to 20 mg NO₂-N L⁻¹ at constant pH of 7.7. TN removal was mainly attributed to nitritation in the photobioreactor and denitritation in the anoxic tank. This unusual NO₂⁻ accumulation and inhibition of NO₃⁻ production was probably due to the high incident light (400 μmol m⁻²L⁻¹), which might have partially inhibited bacterial nitrifiers [178]. Manser et al. [179] proposed the “*algamox*” process, which consisted of TN removal through the conventional Anaerobic Ammonium Oxidation (anammox) process. The anammox process is based on the anaerobic oxidation of ammonia using nitrite as an electron acceptor, giving molecular nitrogen N₂ as a final product. The *algamox* process differs from conventional anammox in that the oxygen for NO₂⁻ production is provided by microalgae [179,180]. The *algamox* process was carried out in a photo-sequencing batch reactor, alternating periods of light and dark, favouring low DO concentrations, since anammox is inhibited by DO above 1.5 mg L⁻¹ [181]. The results showed that ammonia was converted to nitrite during periods of light at a rate of 168 mg NH₄ L⁻¹d⁻¹, and subsequently, an average of 82% of nitrite was reduced to N₂ during the dark (anoxic) period due to anammox activity.

6. MODELLING OF MICROALGAE BACTERIA CONSORTIA

Recently, mathematical modelling has taken an essential role in describing and predicting biological processes such as the MBC system, as we found in the analysis of the frequency of all keywords in Figure 3. Mathematical models are extremely powerful instruments that can be applied for managing existing biotechnologies processes, but also for studying, understanding, and optimizing new processes. Mathematical models have become increasingly attractive since, once calibrated and validated on a long-time scale, they make feasible the investigation of several scenarios and operational conditions. Therefore, the design and operation of MBC systems using models may involve several advantages, such as the reduction of operating costs, the increase in nutrients removal, and the valorization of the nutrients and energy contents of wastewaters.

The classic reference models describing biological processes for waste treatment are Activated Sludge Models (ASMs) [182] and the Anaerobic Digestion Model n°1 (ADM1) [183], which have been widely implemented [184,185]. To date, a large number of models describing microalgae-based bioprocess are available in the literature [186]. However, only a minority of them were developed to represent microalgae and bacteria behavior for wastewater treatment. International Water Association (IWA) introduced a standardized notation for models, including the use of a Petersen matrix. This standardized notation could favor the integration of MBC models with those describing existing water resource recovery facilities (ASMs and ADM) [187]. However, among the microalgae/bacteria reported models, there are only 10 that follows this standardized notation [93,94,96,188–195]. **Table 1** summarizes a comparison between the most recent MBC models available in the literature, for wastewater treatment. Some of the proposed models were developed with the purpose of estimating the biofuels/bioproducts production [196–202]. Among these models, only a few implemented both the phototrophic and heterotrophic metabolisms for the algal biomass, such as the ones proposed by Adesanya et al. [203] and Figueroa-Torres et al. [204]

On the other hand, the calibration and validation of mathematical models are crucial steps to produce reliable simulation results [186,205]. However, there is a lack of long-term validation dataset covering yearly seasonal variation (temperature, incident light, etc.) when working with MBC models for wastewater treatment. Only two works involves long-term seasonal study, the works published by Broekhuizen et al. [192] and by Casagli et al. [93].

Table 1. comparison between the most recent algae-bacteria models available in the literature for wastewater remediation

	RWQM1	PHOBIA	Modified RWQM1 ^(D)	Modified ASM3	Bioalgae1	Bioalgae2	ALBA ^(D)	ABACO
Reference	[189]	[190]	[192]	[195]	[188]	[94]	[93]	Z[96]
State variable (n°)	24	16	24	16	19	19	17	19
Biological processes (n°)	22	13	22	21	18	18	19	18
Parameters (n°)	120	75	138	47	94	108	72*	108
Growth kinetic type	Multiplicative	Minimum	Multiplicative	Multiplicative	Multiplicative	Multiplicative	Multiplicative/ Minimum**	Multiplicative
Dependence on organic and inorganic carbon	C _{ORG}	C _{ORG} , CO ₂ , HCO ₃ ⁻	C _{ORG} , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻	C _{ORG}	C _{ORG} , CO ₂ , HCO ₃ ⁻	C _{ORG} , CO ₂ , HCO ₃ ⁻	C _{ORG} , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻	C _{ORG} , CO ₂ , HCO ₃ ⁻
Considered N-forms	NH ₃ , NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻	NH ₃ ⁺ , NO ₃ ⁻	NH ₃ , NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻ , N ₂	NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻	NH ₃ , NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻	NH ₃ , NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻	N _{org} , NH ₃ , NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻ , HNO ₂ , HNO ₃ , N ₂	NH ₄ ⁺ , NO ₃ ⁻ , NO ₂ ⁻
Considered P-forms	H ₂ PO ₄ ⁻ , HPO ₄ ²⁻	-	H ₂ PO ₄ ⁻ , HPO ₄ ²⁻	-	SPO4 (***)	SPO4 (****)	H ₃ PO ₄ , H ₂ PO ₄ ⁻ , HPO ₄ ²⁻ , PO ₄ ³⁻	SPO4 (****)
Continuity check (mass conservation)	C, O, N, P	(n.s.)	C, O, N, P	(COD, N, P.)	(n.s.)	(n.s.)	C, H, O, N, P, COD	(n.s.)
Algal biomass composition	C ₁₀₀ H ₂₃₂ O ₂₆ N ₁₄ P ₄	(n.s.)	C ₁₀₀ H ₂₃₂ O ₂₆ N ₁₄ P	C ₁₀₆ H ₁₈₁ O ₄₅ N ₁₆ P	C ₁₀₀ H ₂₃₂ O ₂₆ N ₁₄ P	C ₁₀₀ H ₂₃₂ O ₂₆ N ₁₄ P	C ₁₀₀ H ₁₈₃ O ₄₈ N ₁₁ P	C ₁₀₀ H ₂₃₂ O ₂₆ N ₁₄ P
Bacterial biomass composition	C ₁₅₀ H ₃₃₅ O ₁₃ N ₃₀ P ₀	(n.s.)	C ₁₅₀ H ₃₃₅ O ₁₃ N ₃₀ P	C ₅ H ₇ O ₂ N	C ₁₅₀ H ₃₃₅ O ₁₃ N ₃₀ P	C ₁₅₀ H ₃₃₅ O ₁₃ N ₃₀ P	C ₆₀ H ₈₇ O ₂₃ N ₁₂ P	C ₁₅₀ H ₃₃₅ O ₁₃ N ₃₀ P
PAR model	Steele	Eilers & Peters	Smith	Poisson	Eilers & Peters	Eilers & Peters	Bernard & Remond	Molina
pH model	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ₂ PO ₄ ⁻ , HPO ₄ ²⁻ , Ca ²⁺ , H ⁺ , OH ⁻	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ⁺ , OH ⁻ , Δ _{CAT,AN}	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ₂ PO ₄ ⁻ , HPO ₄ ²⁻ , Ca ²⁺ , H ⁺ , OH ⁻	-	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ⁺ , OH ⁻	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ⁺ , OH ⁻	NH ₄ ⁺ , NH ₃ , CO ₂ , HCO ₃ ⁻ , CO ₃ ²⁻ , H ₃ PO ₄ , H ₂ PO ₄ ⁻ , HPO ₄ ²⁻ , PO ₄ ³⁻ , NO ₂ ⁻ , HNO ₂ , NO ₃ ⁻ , HNO ₃ , H ⁺ , OH ⁻ , Δ _{CAT,AN} , TA	-

Experimental setup	pH growth dependence	-	-	Gaussian law	-	-	CPMI	CPM	CPMI
	Temperature simulation/growth dependence	-/Arrhenius	-	-/Arrhenius	-	-/Arrhenius	-/CTMI	✓/CTMI	-/CTMI
	Ammonification	-	-	-	-	-	-	✓	-
	DO inhibition	-	-	-	-	✓	✓	✓	✓
	NH ₃ inhibition	-	-	-	-	-	-	✓	-
	Gas-liquid mass transfer	O ₂	-	O ₂	-	O ₂ , CO ₂ , NH ₃	O ₂ , CO ₂ , NH ₃	O ₂ , CO ₂ , NH ₃ , evaporation	-
	Reactor type	River environment	Laboratory incubator	Raceway	Column PBR	Raceway	Column PBR	Raceway	Column PBR
	Reactor installation/Volume	Outdoor	Indoor(Lab)/3L	Outdoor/8 m ³	Indoor(Lab)/2 L	Outdoor/1 m ³	Indoor(Lab)/4 L	Outdoor/17 m ³ , 1 m ³	Indoor(Lab)/1 L
	Influent	Wastewater discharge	MM	MWW	DSC	MWW	MWW	SWW, DSC	DSC
	Calibration dataset	-	-	365 d	✓ (24 h)	✓ (4 d)	✓ (8 d)	✓ (30 d)	✓ (14 d)
	short-term dynamics	-	-		✓ (24 h)	✓ (4 d)	n.s.	✓ 3, 14 d	✓ (14 d)
	long-term dynamics	-	-	330 d	-	✓ (175 d)	-	✓ (413 d, 189 d)	-
	Sensitivity analysis	✓	✓	-	✓	✓	-	✓	-
Validation	Seasonal analysis	-	-	-	-	-	-	✓	-
	Parameter Uncertainty	✓	-	-	✓	-	-	✓	-
	Confidence intervals for model predictions	✓	-	-	-	-	-	✓	-

Abbreviations: ✓: implemented; (n.s.) not specified or provided in the relative publications; IC: Inorganic Carbon; DSC: Diluted Swine Centrate; MM: Mineral Medium; MWW: Municipal Wastewater; SWW: Synthetic Municipal Wastewater; TA: Total Alkalinity; CTMI: Cardinal Temperature Model with Inflection; CPMI: Cardinal pH Model with Inflection; CPM: Cardinal pH Model. Notes: (D) demonstrative scale reactors (> 5 m³); (*) not including chemical constants, their temperature dependence, and stoichiometric coefficients); (**) in the ALBA model, only the Monod limitation terms relative to nutrients availability were implemented in the minimum function, while the dependence on inhibitory and environmental factors are multiplied for the minimum term (in the PHOBIA model, all the multiplicative terms considered are included in the minimum function); (***) P limitation term only on algae; (****) P limitation term on algae and bacteria

7. BIOMASS RETENTION TECHNOLOGIES

The bibliometric analysis highlights two factors driving the research oriented towards the development or application of biomass retention technologies:

1. The low settling capacity of microalgae, which still represents a major challenge when they are used as a technological platform, due to the high separation and recovery costs, and the need to guarantee a good effluent quality during wastewater treatment [27].
2. The consolidation of solid retention time (SRT) as a key operational parameter, since it affects the type of microorganisms developing in the system, and consequently the nutrient removal [99,195,206].

As shown in **Figure 3**, the research targeting the development of new reactor configurations has boosted from 2014 onwards, mainly to address biomass retention [62,86]. MBC biomass is typically separated by a secondary settler or by applying the sequential batch operation mode, as highlighted by the keywords “SBR,” which showed the highest average frequency in 2018. Indeed, biomass is separated from the liquid phase through gravity sedimentation during a non-aerated or non-mixed phase under SBR operation mode. However, a higher level of automation is required in SBR systems, and separation strongly depends on the biomass aggregation level (flocs). Keywords such as “biofilm”, “immobilization”, “membrane photobioreactor”, and “aerobic granular sludge” increased during the last years, thus resulting in new reactor designs (**Figure 3**). De Godos et al. [176] explored a new configuration based on the concept of denitrification-nitrification CAS with a low C/N ratio, internal recycling (NO_3^-), and external recycling (biomass) that allowed the decoupling of the HRT and SRT. The system consisted of two interconnected tanks: an anoxic tank followed by an aerobic photobioreactor inoculated with cyanobacteria-bacteria. Results showed high organic carbon (>95%) and TN (>90%) removal using the photobioreactor alone at HRT of 3.5 days and SRT of 31 d^{-1} , and using the two-stage anoxic-aerobic configuration at HRT 4.5 days and SRT of 20 d^{-1} . Furthermore, the settler system located after the photobioreactor led to an effluent with $25 \pm 19 \text{ mg L}^{-1}$ of volatile suspended solids, concentration below the European discharge limits. On the other hand, Steen et al. [207] also applied the concept of the CAS, coupling a mixed photobioreactor inoculated with microalgae-bacteria with a settler, providing biomass recirculation to the reactor. The system was able to remove 85% of the applied ammonium load ($66 \text{ mg L}^{-1} \text{ d}^{-1}$) at HRT of 1 day and SRT of 15 d.

Several biofilm-based microalgal bacterial cultivation systems have been reported in recent years, for environmental remediation and biofuel production purposes [208–210]. De Godos et al. [87] engineered an innovative enclosed tubular biofilm photobioreactor inoculated with *Chlorella Sorokiniana* and activated sludge to treat pre-treated swine slurry (7 days of HRT). The system supported removal efficiencies for nitrogen and phosphorus ranging from 94% to 100% and 70% to 90%. The retained biomass was greater than 92% of the generated biomass. Interestingly, *Chlorella Sorokiniana* was dominant throughout the entire experiment. On the other hand, designs such as the rocker system using polystyrene foam as support material [210], the rotating algal disk [211], the rotating rope reactor with spool harvester partially submerged in wastewater [212] and the attached cultivation reactor using glass plate and filter paper as a substratum [213] are among the biofilm-based system’s configuration described in the literature. The

biofilm approach seems to improve the sustainability and the productivity of the microalgae production systems [214]. Compared to conventional open ponds, enhanced biomass productivities have been shown using biofilm-based microalgal cultivation. For instance, Christenson and Sims [212] reported the tertiary wastewater treatment in an enhanced raceway bioreactor using five rotating biological contactors (RBC), close to the paddlewheel of the bioreactor. The five RBC used cotton cords as biofilm support and were 40% submerged in the reactor. With an HRT ranging from 3 to 6 days, an average removal of TN and total phosphorus were 76 % and 23 %, respectively. The achieved biomass productivity was 5.5 g m⁻² d⁻¹ at laboratory scale and 31 g m⁻² d⁻¹ at pilot scale, values higher than those for conventional raceway ponds. In addition, other studies that explored the configuration of biofilm-based systems have reported productivities up to 302% [208], 697% [215] and 700% [216] of the ones expected for traditional raceway ponds.

Researchers from Brazil have worked with hybrid HRAP systems, consisting of the combination of HRAP with a biofilm unit [217,218]. In hybrid HRAP systems, additional carbon source supplementation (CO₂) was not necessary since biofilm provided better conditions for atmospheric CO₂ capture [217]. In addition, hybrid systems followed by a settling tank provided a biomass production 2.6 greater than that of a conventional HRAP followed by a settling tank. Moreover, biomass harvesting in the hybrid system was facilitated [218]. Recently, attached biofilms in carriers have been explored for MBC [59,60,112]. Gou et al. [59] observed a 90% COD and ammonia removal, reducing the HRT to 12 hours, applying a three-dimensional carrier that improved the efficiency by promoting the mass transfer from the wastewater to the biofilm.

Microalgal-bacterial granules have been lately explored to improve biomass settleability and enhance biomass harvesting. Granules formation depends on several factors, such as the inoculum, the composition and concentration of EPS as polysaccharides and proteins, wastewater characteristics, and environmental factors such as seasonal variability that contributes to the change of incident light and temperature [55,219,220]. For instance, Quijano et al. [219] evaluated the influence of light on EPS and granules formation while treating municipal wastewater, at an HRT of 10 days. Low irradiances levels (< 3800 Wh m⁻² d⁻¹) promoted the formation of microalgae-bacteria flocs and granules with high settleability (432 m d⁻¹) and relatively high removal efficiencies of TN (60 %) and total COD (89 %). Conversely, phosphorus removal did not experience the same trend at low irradiances, exhibiting removals of 92 % at 6213 Wh m⁻² d⁻¹ and 28% at incident light lower than 3800 Wh m⁻² d⁻¹. Ji et al. (2020) used microalgal-bacterial granules for synthetic wastewater treatment and reported removal efficiencies of 93 %, 97% and 87% for organic matter, ammonia, and phosphorus, respectively; operating in cycles of 6 hours under non-aeration conditions. The most abundant microalgae found in this system was *Pantalaninema Rosaneae*.

The most recent bioreactor configuration explored for MBC is membrane bioreactors (MBR) [145]. MBRs have been widely explored for aerobic wastewater treatment as an alternative to biomass settling. In MBRs, the cultivation and removal of contaminants are carried out in a reactor, with a membrane system providing complete solids retention and providing a clarified effluent. HRTs as low as 8-24 hours have been reported, with fluxes in the range of 168-360 L m⁻² d⁻¹, achieving high pollutant removal. In addition, easier operation, amelioration of microbial activity, and better pollutants removal have been achieved in MBR systems inoculated with microalgae and bacteria than in MBR systems

inoculated with only activated sludge [62,63]. Furthermore, lower membrane fouling has been reported when operating with microalgae and bacteria consortia, in comparison with a system worked only with activated sludge [98]. In fact, the presence of microalgae in the inoculum is used to improve the biomass quality by transforming the EPS composition and thus improving floc filterability and also decreasing the cake-layer formation in the membrane [63,98].

All these biomass retention technologies have become an essential issue for efficient microalgae valorization. Regardless of the technology used for biomass retention in MBC systems, some researchers have emphasized the importance of carefully selecting the SRT and subsequently the biomass concentration, avoiding growth limitation by self-shading and thus low light availability for microalgae in the reactor [48,206].

8. FUTURE PERSPECTIVES

MBC has been already studied for more than 70 years, and the knowledge consolidation is getting faster. This literature research identified the current trends in the main research streams in MBC for wastewater treatment. The first identified trend is the elucidation of nutrient removal mechanisms in the MBC, under the influence of factors such as incident light, pH, temperature, and dissolved oxygen (DO). Future research studies should not only focus on the conditions that can guarantee the removal of nutrients and organic matter, but also establish the fluxes and symbiotic mechanisms determining process performance. For instance, the environmental and operational conditions that promote nitrogen assimilation over nitrification, and vice versa. In addition, more research is needed in new nitrogen removal mechanisms such as the “shortcut nitrogen removal” or “algammox”. Likewise, more attention should be paid to the removal mechanisms that promote “luxury” uptake of phosphorus by microalgae or the proliferation of PAOs. On the other hand, although MBC has the potential to reduce the direct and indirect emission of CO₂, microalgal-bacterial photobioreactors can lead to the release of NH₃ or the production of N₂O, because of inorganic carbon competition between nitrifiers and microalgae. Thus, these removal mechanisms must further be understood and evaluated together with the potential emissions of GHG.

Mathematical modeling represents an extremely powerful tool for studying, understanding, and optimizing microalgae-bacteria processes. The application of mathematical models must be promoted, since models are especially useful for managing the complexity and nonlinearities arising from the interactions among microalgae and bacteria and physical-chemical processes. involved in nutrients removal mechanisms and GHG emissions. On the other hand, long-term dataset validation must become a standard, to promote reliable mathematical representations that can represent the process over a wide range of potential scenarios and operational conditions.

Low microalgae settling capacity is another issue identified during literature analysis. This issue can hinder biomass separation by settling, which is the most used solid-liquid separation operation in wastewater treatment. Combination of membranes and photobioreactors is expected to bring new opportunities for biomass retention and an effluent clarification. Moreover, the evaluation of biomass properties affecting membrane fouling membrane is key issue. Furthermore, the use of photosynthetic biofilms represents an interesting alternative to facilitate biomass harvesting and valorization. Moreover, the application of different biofilm-based reactor configurations has shown potential for wastewater treatment and the enhancement of biomass productivity. Future

works must focus on the strategies and operational conditions of these systems. Finally, more studies are necessary for both membrane and biofilm photobioreactors, optimizing the design and operating strategy and implementing these technologies at pilot and/or full scale.

9. CONCLUSIONS

- The mechanisms involved in the removal of nutrients and COD depend mainly on the interactions of microorganisms and, at the same time, on factors affecting these interactions, such as pH, light, dissolved oxygen, C/N/P ratio. These factors directly impact the chemical equilibria of pollutants and the presence of toxic compounds (e.g., free NH_3 , HNO_2), and inhibitors. Further research is needed to assess the influence of these factors on interactions and removal mechanisms.
- Novel nutrient removal mechanisms such as "short-cut nitrogen removal," "algammox," and "phosphorus luxury uptake" have been proposed in the past decade. These could present an alternative to improve nutrients removal during wastewater treatment. However, more research is needed to evaluate the specific conditions in which these mechanisms are predominant.
- The application of mathematical modelling has been explored in MBC systems. This tool presents a high potential for the optimization of MBC systems. However, models need to be correctly calibrated and validated.
- Different reactor configurations have been explored to decouple HRT from SRT, and to keep slow-growing microorganisms such as nitrifiers in the reactor. The membrane bioreactor and biofilm photobioreactors have shown high potential, presenting high removal levels of nutrients and organic matter, while facilitating biomass harvesting and reducing systems footprint. These emerging technologies must be thoroughly explored and evaluated, to determine potential full-scale applicability.
- More research efforts must be devoted to assessing the performance of MBC for wastewater treatment at a large scale, including the revalorization of biomass and reuse of treated water, along with its socio-environmental and economic implications.
- MBC has been highlighted as a prominent wastewater technology that could compete with conventional systems. However, the main challenges associated are a high footprint, harvesting costs, and biomass processing to produce high-value products.

Declaration of Conflicting Interests

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Author contribution statements

Jineth Arango conceived the structure, wrote the manuscript, designed the SQ and the analysis with permanent feedback from David Jeison. Francesca Casagli, contributed to the structure of modelling section. Olivier Bernard, Francesca Casagli, Raúl Muñoz, and Andrés Donoso contributed to the technical aspects and the critical review of the document, as well as contributed to the structure and conceptualization of the document. All authors provided critical feedback and helped shaping the research, analysis, and manuscript writing.

REFERENCES

- [1] WWAP (United Nations World Water Assessment Programme), The United Nations World Water Development Report 2017. Wastewater: The Untapped Resource., París, UNESCO, 2017. www.unwater.org.
- [2] E. Corcoran, C. Nellemann, E. Baker, R. Bos, D. Osborn, H. Savelli, Sick Water? The Central Role of Wastewater Management in Sustainable Development. A Rapid Response Assessment, The Hague, UN-Habitat/ UNEP/GRID-Arendal, 2010.
- [3] P.M. Nyenje, J.W. Foppen, S. Uhlenbrook, R. Kulabako, A. Muwanga, Eutrophication and nutrient release in urban areas of sub-Saharan Africa - A review, *Sci. Total Environ.* 408 (2010) 447–455. <https://doi.org/10.1016/j.scitotenv.2009.10.020>.
- [4] WWAP (UNESCO World Water Assessment Programme), The United Nations World Water Development Report 2019: Leaving No One Behind., Paris, UNESCO, 2019.
- [5] A. Noyola, A. Padilla-Rivera, J.M. Morgan-Sagastume, F. Herna, Typology of Municipal Wastewater Treatment Technologies in Latin America, 40 (2012) 926–932. <https://doi.org/10.1002/clen.201100707>.
- [6] Y.J. Chan, M.F. Chong, C.L. Law, D.G. Hassell, A review on anaerobic-aerobic treatment of industrial and municipal wastewater, *Chem. Eng. J.* 155 (2009) 1–18. <https://doi.org/10.1016/j.cej.2009.06.041>.
- [7] J. Drewnowski, A. Remiszewska-Skwarek, S. Duda, G. Lagod, Aeration Process in Bioreactors as the Main Energy Consumer in a Wastewater Treatment Plant . Review of Solutions and Methods of process optimization, *Processes.* (2019).
- [8] Oilgae., Oilgae Guide to Algae-based Wastewater Treatment A Sample Report, Retrived from [http:// repository.uobabylon.edu.iq/2010_2011/4_6558_416.pdf](http://repository.uobabylon.edu.iq/2010_2011/4_6558_416.pdf),

- Chennai, India, 2009.
- [9] H.Y. Dong, Z.M. Qiang, W.D. Wang, H. Jin, Evaluation of rural wastewater treatment processes in a county of eastern China, *J. Environ. Monit.* 14 (2012) 1906–1913. <https://doi.org/10.1039/c2em10976j>.
- [10] I. Lee, H. Lim, B. Jung, M.F. Colosimo, H. Kim, Evaluation of aeration energy saving in two modified activated sludge processes, *Chemosphere.* (2015). <https://doi.org/10.1016/j.chemosphere.2014.03.086>.
- [11] D. Rosso, L.E. Larson, M.K. Stenstrom, Aeration of large-scale municipal wastewater treatment plants: State of the art, *Water Sci. Technol.* (2008). <https://doi.org/10.2166/wst.2008.218>.
- [12] W.J. Oswald, H.B. Gotaas, H.F. Ludwig, V. Lynch, S. Sewage, I. Wastes, N. Jun, I. Wastes, J. Oswald, F. Ludwig, *Algae Symbiosis in Oxidation Ponds : III . Photosynthetic Oxygenation*, 25 (1953) 692–705.
- [13] N.G.A.I. Karya, N.P. van der Steen, P.N.L. Lens, Photo-oxygenation to support nitrification in an algal-bacterial consortium treating artificial wastewater, *Bioresour. Technol.* 134 (2013) 244–250. <https://doi.org/10.1016/j.biortech.2013.02.005>.
- [14] F.J. Humenik, G.P. Hanna, P. Hanna, *Algal-Bacterial Symbiosis for Removal and Conservation of Wastewater Nutrients*, *Water.* 43 (1971) 580–594.
- [15] G. Mujtaba, K. Lee, Advanced treatment of wastewater using symbiotic co-culture of microalgae and bacteria, *Appl. Chem. Eng.* 27 (2016). <https://doi.org/10.14478/ace.2016.1002>.
- [16] R. Muñoz, B. Guieysse, Algal – bacterial processes for the treatment of hazardous contaminants : A review, *Water Res.* 40 (2006) 2799–2815. <https://doi.org/10.1016/j.watres.2006.06.011>.
- [17] C.C. Tang, W. Zuo, Y. Tian, N. Sun, Z.W. Wang, J. Zhang, Effect of aeration rate on performance and stability of algal-bacterial symbiosis system to treat domestic wastewater in sequencing batch reactors, *Bioresour. Technol.* 222 (2016) 156–164. <https://doi.org/10.1016/j.biortech.2016.09.123>.
- [18] H. Khaldi, M. Maatoug, C.S. Dube, M. Ncube, R. Tandlich, H. Heilmeyer, R.K. Laubscher, A. Dellal, Efficiency of wastewater treatment by a mixture of sludge and microalgae, *J. Fundam. Appl. Sci.* 9 (2017) 1454. <https://doi.org/10.4314/jfas.v9i3.13>.
- [19] G. Gutzeit, D. Lorch, A. Weber, M. Engels, U. Neis, Bioflocculent algal-bacterial biomass improves low-cost wastewater treatment, *Water Sci. Technol.* (2015). <https://doi.org/10.2166/wst.2005.0415>.
- [20] D. Kang, K. Kim, Y. Jang, H. Moon, D. Ju, G. Kwon, D. Jahng, Enhancement of wastewater treatment efficiency through modulation of aeration and blue light on wastewater-borne algal-bacterial consortia, *Int. Biodeterior. Biodegrad.* 135 (2018) 9–18. <https://doi.org/10.1016/j.ibiod.2018.07.008>.
- [21] A.L. Gonçalves, J.C.M. Pires, M. Simões, A review on the use of microalgal consortia for wastewater treatment, *Algal Res.* 24 (2017) 403–415. <https://doi.org/10.1016/j.algal.2016.11.008>.
- [22] T. Essam, M. ElRakaiby, A. Hashem, Photosynthetic based algal-bacterial

- combined treatment of mixtures of organic pollutants and CO₂ mitigation in a continuous photobioreactor, *World J. Microbiol. Biotechnol.* 29 (2013) 969–974. <https://doi.org/10.1007/s11274-013-1254-z>.
- [23] A. Anbalagan, A. Toledo-Cervantes, E. Posadas, E.M. Rojo, R. Lebrero, A. González-Sánchez, E. Nehrenheim, R. Muñoz, Continuous photosynthetic abatement of CO₂ and volatile organic compounds from exhaust gas coupled to wastewater treatment: Evaluation of tubular algal-bacterial photobioreactor, *J. CO₂ Util.* 21 (2017) 353–359. <https://doi.org/10.1016/j.jcou.2017.07.016>.
- [24] Wang, Y. Li, N. Wu, C.Q. Lan, CO₂ bio-mitigation using microalgae, *Appl. Microbiol. Biotechnol.* 79 (2008) 707–718. <https://doi.org/10.1007/s00253-008-1518-y>.
- [25] Á. Robles, G. Capson-Tojo, A. Galès, M.V. Ruano, B. Sialve, J. Ferrer, J.P. Steyer, Microalgae-bacteria consortia in high-rate ponds for treating urban wastewater: Elucidating the key state indicators under dynamic conditions, *J. Environ. Manage.* 261 (2020). <https://doi.org/10.1016/j.jenvman.2020.110244>.
- [26] G. Ruas, M.L. Serejo, P.L. Paulo, M.Á. Boncz, Evaluation of domestic wastewater treatment using microalgal-bacterial processes: effect of CO₂ addition on pathogen removal, *J. Appl. Phycol.* 30 (2018) 921–929. <https://doi.org/10.1007/s10811-017-1280-6>.
- [27] E.J. Olguín, Dual purpose microalgae-bacteria-based systems that treat wastewater and produce biodiesel and chemical products within a Biorefinery, *Biotechnol. Adv.* 30 (2012) 1031–1046. <https://doi.org/10.1016/j.biotechadv.2012.05.001>.
- [28] J. Yang, W. Shi, F. Fang, G. Guo, Jinsong, L. Lu, Y. Xiao, X. Jiang, Exploring the feasibility of sewage treatment by algal–bacterial consortia, *Crit. Rev. Biotechnol.* 0 (2020) 1–11. <https://doi.org/10.1080/07388551.2019.1709796>.
- [29] J.A. Moral-Muñoz, E. Herrera-Viedma, A. Santisteban-Espejo, M.J. Cobo, Software tools for conducting bibliometric analysis in science: An up-to-date review, *El Prof. La Inf.* 29 (2020) 1–20.
- [30] Elaine Gauthier, *Bibliometric analysis of scientific and technological research: A user's guide to the methodology*, (1998) 81.
- [31] Alan Pritchard, *Statistical Bibliography or Bibliometrics?*, *J. Doc.* (1969) 348–349.
- [32] L. Castell, Trends in scientific research on climate change in agriculture and forestry subject areas (2005 e 2014), 147 (2017) 406–418. <https://doi.org/10.1016/j.jclepro.2017.01.112>.
- [33] F. León-Vargas, J. Arango-Oviedo, H.J. Luna Wandurraga, Two Decades of Research in Artificial Pancreas : Insights from a Bibliometric Analysis, *J. Dia.* (2021). <https://doi.org/10.1177/19322968211005500>.
- [34] D.C. Calvo, H.J. Luna Wandurraga, J. Arango-Oviedo, C.I. Torres, B.E. Rittmann, Determining global trends in syngas fermentation research through a bibliometric analysis, *J. Environ. Manage.* 307 (2022) 114522. <https://doi.org/10.1016/j.jenvman.2022.114522>.
- [35] H. Chen, W. Jiang, Y. Yang, Y. Yang, X. Man, State of the art on food waste research : a bibliometrics study from 1997 to 2014, 140 (2017) 840–846.

- 320 <https://doi.org/10.1016/j.jclepro.2015.11.085>.
- 321 [36] W. Gao, Y. Chen, Y. Liu, H. cheng Guo, Scientometric analysis of phosphorus
322 research in eutrophic lakes, *Scientometrics*. (2015).
323 <https://doi.org/10.1007/s11192-014-1500-7>.
- 324 [37] J.M. Merigó, J. Miranda, N.M. Modak, G. Boustras, Forty years of Safety
325 Science : A bibliometric overview, *Saf. Sci.* 115 (2019) 66–88.
326 <https://doi.org/10.1016/j.ssci.2019.01.029>.
- 327 [38] O. Persson, R. Danell, J.W. Schneider, How to use Bibexcel for various types of
328 bibliometric analysis, *Celebr. Sch. Commun. Stud. A Festschrift Olle Persson His*
329 *60th Birthd.* (2009).
- 330 [39] S. Zhong, Y. Geng, W. Liu, C. Gao, W. Chen, A bibliometric review on natural
331 resource accounting during 1995 e 2014, *J. Clean. Prod.* 139 (2016) 122–132.
332 <https://doi.org/10.1016/j.jclepro.2016.08.039>.
- 333 [40] A. Yataganbaba, B. Ozkahraman, I. Kurtbas, Worldwide trends on encapsulation
334 of phase change materials: A bibliometric analysis (1990–2015), *Appl. Energy*.
335 185 (2017) 720–731. <https://doi.org/10.1016/j.apenergy.2016.10.107>.
- 336 [41] J.E. Hirsch, An index to quantify an individual’s scientific research output, *Proc.*
337 *Natl. Acad. Sci. U. S. A.* (2005). <https://doi.org/10.1073/pnas.0507655102>.
- 338 [42] R. Salvador, M.V. Barros, L.M. da Luz, C.M. Piekarski, A.C. de Francisco,
339 Circular business models: Current aspects that influence implementation and
340 unaddressed subjects, *J. Clean. Prod.* 250 (2020) 119555.
341 <https://doi.org/10.1016/j.jclepro.2019.119555>.
- 342 [43] Z. Li, Y.S. Ho, Use of citation per publication as an indicator to evaluate
343 contingent valuation research, *Scientometrics*. (2008).
344 <https://doi.org/10.1007/s11192-007-1838-1>.
- 345 [44] N.J. Van Eck, L. Waltman, Software survey: VOSviewer, a computer program
346 for bibliometric mapping, *Scientometrics*. 84 (2010) 523–538.
347 <https://doi.org/10.1007/s11192-009-0146-3>.
- 348 [45] N.J. Van Eck, L. Waltman, Visualizing Bibliometric Networks, in: *Meas. Sch.*
349 *Impact Methods Pract.*, 2014: pp. 285–320. [https://doi.org/10.1007/978-3-319-](https://doi.org/10.1007/978-3-319-10377-8_13)
350 [10377-8_13](https://doi.org/10.1007/978-3-319-10377-8_13).
- 351 [46] Y. Qi, X. Chen, Z. Hu, C. Song, Y. Cui, Bibliometric Analysis of Algal-Bacterial
352 Symbiosis in Wastewater Treatment, *Int. J. Environ. Res. Public Health*. (2019).
353 <https://doi.org/10.3390/ijerph16061077>.
- 354 [47] Y. Su, A. Mennerich, B. Urban, Synergistic cooperation between wastewater-
355 born algae and activated sludge for wastewater treatment: Influence of algae and
356 sludge inoculation ratios, *Bioresour. Technol.* 105 (2012) 67–73.
357 <https://doi.org/10.1016/j.biortech.2011.11.113>.
- 358 [48] A. Sepehri, M.-H. Sarrafzadeh, M. Avateffazeli, Interaction between *Chlorella*
359 *Vulgaris* and nitrifying-enriched activated sludge in the treatment of wastewater
360 with low C/N ratio, *J. Clean. Prod.* 247 (2020).
361 <https://doi.org/10.1016/j.jclepro.2019.119164>.
- 362 [49] S.F. Mohsenpour, S. Hennige, N. Willoughby, A. Adeloye, T. Gutierrez,
363 Integrating micro-algae into wastewater treatment: A review, *Sci. Total Environ.*

364 752 (2021) 142168. <https://doi.org/10.1016/j.scitotenv.2020.142168>.

365 [50] N. Renuka, A. Guldhe, R. Prasanna, P. Singh, F. Bux, Microalgae as multi-
366 functional options in modern agriculture: current trends, prospects and
367 challenges, *Biotechnol. Adv.* 36 (2018) 1255–1273.
368 <https://doi.org/10.1016/j.biotechadv.2018.04.004>.

369 [51] X. Ji, M. Jiang, J. Zhang, X. Jiang, Z. Zheng, The interactions of algae-bacteria
370 symbiotic system and its effects on nutrients removal from synthetic wastewater,
371 *Bioresour. Technol.* 247 (2018) 44–50.
372 <https://doi.org/10.1016/j.biortech.2017.09.074>.

373 [52] B. Zhang, P.N.L. Lens, W. Shi, R. Zhang, Z. Zhang, Y. Guo, X. Bao, F. Cui,
374 Enhancement of aerobic granulation and nutrient removal by an algal–bacterial
375 consortium in a lab-scale photobioreactor, *Chem. Eng. J.* 334 (2018) 2373–2382.
376 <https://doi.org/10.1016/j.cej.2017.11.151>.

377 [53] Y. Wang, S.-H. Ho, C.-L. Cheng, W.-Q. Guo, D. Nagarajan, N.-Q. Ren, D.-J.
378 Lee, J.-S. Chang, Perspectives on the feasibility of using microalgae for industrial
379 wastewater treatment, *Bioresour. Technol.* 222 (2016) 485–497.
380 <https://doi.org/10.1016/j.biortech.2016.09.106>.

381 [54] J. Liu, Y. Wu, C. Wu, K. Muylaert, W. Vyverman, H.-Q. Yu, R. Muñoz, B.
382 Rittmann, Advanced nutrient removal from surface water by a consortium of
383 attached microalgae and bacteria: A review, *Bioresour. Technol.* 241 (2017)
384 1127–1137. <https://doi.org/10.1016/j.biortech.2017.06.054>.

385 [55] L. Liu, H. Fan, Y. Liu, C. Liu, X. Huang, Development of algae-bacteria granular
386 consortia in photo-sequencing batch reactor, *Bioresour. Technol.* 232 (2017) 64–
387 71. <https://doi.org/10.1016/j.biortech.2017.02.025>.

388 [56] F. Meng, L. Xi, D. Liu, W. Huang, Z. Lei, Z. Zhang, W. Huang, Effects of light
389 intensity on oxygen distribution, lipid production and biological community of
390 algal-bacterial granules in photo-sequencing batch reactors, *Bioresour. Technol.*
391 (2019). <https://doi.org/10.1016/j.biortech.2018.10.059>.

392 [57] L. Liu, Y. Hong, X. Ye, L. Wei, J. Liao, X. Huang, C. Liu, Biodiesel production
393 from microbial granules in sequencing batch reactor, *Bioresour. Technol.* (2018).
394 <https://doi.org/10.1016/j.biortech.2017.10.105>.

395 [58] Z. Zhao, X. Yang, W. Cai, Z. Lei, K. Shimizu, Z. Zhang, M. Utsumi, D.J. Lee,
396 Response of algal-bacterial granular system to low carbon wastewater: Focus
397 Response of algal-bacterial granular system to low carbon wastewater: Focus on
398 granular, *Bioresour. Technol.* (2018).
399 <https://doi.org/10.1016/j.biortech.2018.07.114>.

400 [59] Y. Gou, J. Yang, F. Fang, J. Guo, H. Ma, Feasibility of using a novel algal-
401 bacterial biofilm reactor for efficient domestic wastewater treatment, *Environ.*
402 *Technol. (United Kingdom)*. 41 (2020) 400–410.
403 <https://doi.org/10.1080/09593330.2018.1499812>.

404 [60] C.C. Tang, Y. Tian, Z.W. He, W. Zuo, J. Zhang, Performance and mechanism of
405 a novel algal-bacterial symbiosis system based on sequencing batch suspended
406 biofilm reactor treating domestic wastewater, *Bioresour. Technol.* 265 (2018)
407 422–431. <https://doi.org/10.1016/j.biortech.2018.06.033>.

408 [61] L. Sun, Y. Tian, J. Zhang, L. Li, J. Zhang, J. Li, A novel membrane bioreactor

- inoculated with symbiotic sludge bacteria and algae: Performance and microbial community analysis, *Bioresour. Technol.* 251 (2018) 311–319. <https://doi.org/10.1016/j.biortech.2017.12.048>.
- [62] J. Yang, Y. Gou, F. Fang, J. Guo, L. Lu, Y. Zhou, H. Ma, Potential of wastewater treatment using a concentrated and suspended algal-bacterial consortium in a photo membrane bioreactor, *Chem. Eng. J.* 335 (2018) 154–160. <https://doi.org/10.1016/j.cej.2017.10.149>.
- [63] L. Sun, Y. Tian, J. Zhang, H. Cui, W. Zuo, J. Li, A novel symbiotic system combining algae and sludge membrane bioreactor technology for wastewater treatment and membrane fouling mitigation: performance and mechanisms, *Bioresour. Technol.* 251 (2018) 311–319.
- [64] L. Sun, J. Ma, L. Li, Y. Tian, Z. Zhang, H. Liao, J. Li, W. Tang, D. He, Exploring the essential factors of performance improvement in sludge membrane bioreactor technology coupled with symbiotic algae, *Water Res.* (2020). <https://doi.org/10.1016/j.watres.2020.115843>.
- [65] M.B. Paddock, J.D. Fernández-Bayo, J.S. VanderGheynst, The effect of the microalgae-bacteria microbiome on wastewater treatment and biomass production, *Appl. Microbiol. Biotechnol.* (2020). <https://doi.org/10.1007/s00253-019-10246-x>.
- [66] J. Fan, Y. Chen, T.C. Zhang, B. Ji, L. Cao, Performance of *Chlorella sorokiniana*-activated sludge consortium treating wastewater under light-limited heterotrophic condition, *Chem. Eng. J.* (2020). <https://doi.org/10.1016/j.cej.2019.122799>.
- [67] S. Rossi, F. Casagli, M. Mantovani, V. Mezzanotte, E. Ficara, Selection of photosynthesis and respiration models to assess the effect of environmental conditions on mixed microalgae consortia grown on wastewater, *Bioresour. Technol.* 305 (2020). <https://doi.org/10.1016/j.biortech.2020.122995>.
- [68] S. Petrini, P. Foladori, L. Donati, G. Andreottola, Comprehensive respirometric approach to assess photosynthetic, heterotrophic and nitrifying activity in microalgal-bacterial consortia treating real municipal wastewater, *Biochem. Eng. J.* 161 (2020) 107697. <https://doi.org/10.1016/j.bej.2020.107697>.
- [69] S. Rossi, E. Sforza, M. Pastore, M. Bellucci, F. Casagli, F. Marazzi, E. Ficara, Photo-respirometry to shed light on microalgae-bacteria consortia—a review, *Rev. Environ. Sci. Biotechnol.* 19 (2020) 43–72. <https://doi.org/10.1007/s11157-020-09524-2>.
- [70] S. Rossi, M. Bellucci, F. Marazzi, V. Mezzanotte, E. Ficara, Activity assessment of microalgal-bacterial consortia based on respirometric tests, *Water Sci. Technol.* 78 (2018) 207–215. <https://doi.org/10.2166/wst.2018.078>.
- [71] E. Sforza, M. Pastore, A. Spagni, A. Bertucco, Microalgae-bacteria gas exchange in wastewater: how mixotrophy may reduce the oxygen supply for bacteria, *Environ. Sci. Pollut. Res.* 25 (2018) 28004–28014. <https://doi.org/10.1007/s11356-018-2834-0>.
- [72] P. Foladori, S. Petrini, G. Andreottola, Evolution of real municipal wastewater treatment in photobioreactors and microalgae-bacteria consortia using real-time parameters, *Chem. Eng. J.* 345 (2018) 507–516.

- 454 <https://doi.org/10.1016/j.cej.2018.03.178>.
- 455 [73] M. Mantovani, F. Marazzi, R. Fornaroli, M. Bellucci, E. Ficara, V. Mezzanotte,
456 Outdoor pilot-scale raceway as a microalgae-bacteria sidestream treatment in a
457 WWTP, *Sci. Total Environ.* 710 (2020).
458 <https://doi.org/10.1016/j.scitotenv.2019.135583>.
- 459 [74] A.W.J. Oswald, H.B. Gotaas, C.G. Golueke, W.R. Kellen, E.F. Gloyna, S.
460 Sewage, I. Wastes, N. Apr, J. Oswald, R. Kellen, *Algae in Waste Treatment*,
461 *Sewage Ind. Waste.* 29 (1957) 437–457.
- 462 [75] Y. Nurdogan, W.J. Oswald, Enhanced nutrient removal in high-rate ponds, *Water*
463 *Sci. Technol.* 31 (1995) 33–43. [https://doi.org/10.1016/0273-1223\(95\)00490-E](https://doi.org/10.1016/0273-1223(95)00490-E).
- 464 [76] P.S. Lau, N.F.Y. Tam, Y.S. Wong, Effect of algal density on nutrient removal
465 from primary settled wastewater, *Environ. Pollut.* 89 (1995) 59–66.
466 [https://doi.org/10.1016/0269-7491\(94\)00044-E](https://doi.org/10.1016/0269-7491(94)00044-E).
- 467 [77] M.A. Khan, S.I. Ahmad, Performance evaluation of pilot waste stabilization
468 ponds in subtropical region, *Water Sci. Technol.* 26 (1992) 1717–1728.
469 <https://doi.org/10.2166/wst.1992.0615>.
- 470 [78] M.A. Khan, M.A. Khan, The potential of waste stabilization ponds effluent as a
471 liquid fertilizer, *Pakistan J. Bot.* 39 (2007) 817–829.
- 472 [79] Y. Su, A. Mennerich, B. Urban, Municipal wastewater treatment and biomass
473 accumulation with a wastewater-born and settleable algal-bacterial culture, *Water*
474 *Res.* 45 (2011) 3351–3358. <https://doi.org/10.1016/j.watres.2011.03.046>.
- 475 [80] W.J. Oswald, A.M. ASCE, H.B. Gotaas, M. ASCE, Photosynthesis in sewage
476 treatment, *Am. Soc. Civ. Eng.* 2849 (1985) 33.
- 477 [81] H.F. Ludwig, W.J. Oswald, H.B. Gotaas, V. Lynch, J. Oswald, F. Ludwig, *Algae*
478 *Symbiosis in Oxidation Ponds: I. Growth Characteristics of “Euglena Gracilis”*
479 *Cultured in Sewage*, *Sewage Ind. Waste.* 23 (1951) 1337–1355.
- 480 [82] I. de Godos, V.A. Vargas, S. Blanco, M.C.G. González, R. Soto, P.A. García-
481 Encina, E. Becares, R. Muñoz, A comparative evaluation of microalgae for the
482 degradation of piggery wastewater under photosynthetic oxygenation, *Bioresour.*
483 *Technol.* 101 (2010) 5150–5158. <https://doi.org/10.1016/j.biortech.2010.02.010>.
- 484 [83] E. Tamer, M.A. Amin, E.T. Ossama, M. Bo, G. Benoit, Biological treatment of
485 industrial wastes in a photobioreactor, *Water Sci. Technol.* 53 (2006) 117–125.
486 <https://doi.org/10.2166/wst.2006.344>.
- 487 [84] E. Safonova, K. V. Kvitko, M.I. Iankevitch, L.F. Surgko, I.A. Afti, W. Reisser,
488 Biotreatment of industrial wastewater by selected algal-bacterial consortia, *Eng.*
489 *Life Sci.* 4 (2004) 347–353. <https://doi.org/10.1002/elsc.200420039>.
- 490 [85] J.P. Hoffmann, Wastewater treatment with suspended and nonsuspended algae, *J.*
491 *Phycol.* 34 (1998) 757–763. <https://doi.org/10.1046/j.1529-8817.1998.340757.x>.
- 492 [86] N.C. Boelee, H. Temmink, M. Janssen, C. Buisman, R. Wijffels, Scenario
493 analysis of nutrient removal from municipal wastewater by microalgal biofilms,
494 *Water (Switzerland).* (2012). <https://doi.org/10.3390/w4020460>.
- 495 [87] I. de Godos, C. González, E. Becares, P.A. García-Encina, R. Muñoz,
496 Simultaneous nutrients and carbon removal during pretreated swine slurry
497 degradation in a tubular biofilm photobioreactor, *Appl. Microbiol. Biotechnol.* 82

- 498 (2009) 187–194. <https://doi.org/10.1007/s00253-008-1825-3>.
- 499 [88] B. Ji, M. Zhang, J. Gu, Y. Ma, Y. Liu, A self-sustaining synergetic microalgal-
500 bacterial granular sludge process towards energy-efficient and environmentally
501 sustainable municipal wastewater treatment, *Water Res.* 179 (2020) 115884.
502 <https://doi.org/10.1016/j.watres.2020.115884>.
- 503 [89] L.E. de-Bashan, M. Moreno, J.P. Hernandez, Y. Bashan, Removal of ammonium
504 and phosphorus ions from synthetic wastewater by the microalgae *Chlorella*
505 *Vulgaris* coimmobilized in alginate beads with the microalgae growth-promoting
506 bacterium *Azospirillum* *Brasilense*, *Water Res.* 36 (2001) 2941–2948.
507 [https://doi.org/10.1016/S0043-1354\(01\)00522-X](https://doi.org/10.1016/S0043-1354(01)00522-X).
- 508 [90] L.E. de-Bashan, J.P. Hernandez, T. Morey, Y. Bashan, Microalgae growth-
509 promoting bacteria as “helpers” for microalgae: A novel approach for removing
510 ammonium and phosphorus from municipal wastewater, *Water Res.* 38 (2003)
511 466–474. <https://doi.org/10.1016/j.watres.2003.09.022>.
- 512 [91] A. Sánchez-Zurano, J.A. Garrido-Cárdenas, C. Gómez-Serrano, M. Morales-
513 Amaral, F.G. Acién-Fernández, J.M. Fernández-Sevilla, E. Molina-Grima, Year-
514 long assessment of a pilot-scale thin-layer reactor for microalgae wastewater
515 treatment. Variation in the microalgae-bacteria consortium and the impact of
516 environmental conditions, *Algal Res.* 50 (2020) 101983.
517 <https://doi.org/10.1016/j.algal.2020.101983>.
- 518 [92] L.M. Trebuch, B.O. Oyserman, M. Janssen, R.H. Wijffels, L.E.M. Vet, T. V.
519 Fernandes, Impact of hydraulic retention time on community assembly and
520 function of photogranules for wastewater treatment, *Water Res.* 173 (2020)
521 115506. <https://doi.org/10.1016/j.watres.2020.115506>.
- 522 [93] F. Casagli, G. Zuccaro, O. Bernard, J.P. Steyer, E. Ficara, ALBA: A
523 comprehensive growth model to optimize algae-bacteria wastewater treatment in
524 raceway ponds, *Water Res.* 190 (2021) 1–60.
525 <https://doi.org/10.1016/j.watres.2020.116734>.
- 526 [94] A. Solimeno, C. Gomez-Serrano, F.G. Acién, BIO _ ALGAE 2 : improved model
527 of microalgae and bacteria consortia for wastewater treatment, *Environ. Sci.*
528 *Pollut. Res.* 26(25) (2019) 25855–25868.
- 529 [95] S. Arun, N.A. Manikandan, K. Pakshirajan, G. Pugazhenth, Novel shortcut
530 biological nitrogen removal method using an algae-bacterial consortium in a
531 photo-sequencing batch reactor: Process optimization and kinetic modelling, *J.*
532 *Environ. Manage.* 250 (2019) 109401.
533 <https://doi.org/10.1016/j.jenvman.2019.109401>.
- 534 [96] A. Sánchez-Zurano, E. Rodríguez-Miranda, J.L. Guzmán, F.G. Acién-Fernández,
535 J.M. Fernández-Sevilla, E.M. Grima, Abaco: A new model of microalgae-
536 bacteria consortia for biological treatment of wastewaters, *Appl. Sci.* 11 (2021)
537 1–24. <https://doi.org/10.3390/app11030998>.
- 538 [97] Á. Robles, G. Capson-Tojo, A. Gales, A. Viruela, B. Sialve, A. Seco, J.P. Steyer,
539 J. Ferrer, Performance of a membrane-coupled high-rate algal pond for urban
540 wastewater treatment at demonstration scale, *Bioresour. Technol.* 301 (2020)
541 122672. <https://doi.org/10.1016/j.biortech.2019.122672>.
- 542 [98] L. Sun, Y. Tian, J. Zhang, H. Li, C. Tang, J. Li, Wastewater treatment and

- 543 membrane fouling with algal-activated sludge culture in a novel membrane
544 bioreactor : Influence of inoculation ratios, *Chem. Eng. J.* 343 (2018) 455–459.
545 <https://doi.org/10.1016/j.cej.2018.03.022>.
- 546 [99] A.M. Rada-Ariza, D. Fredy, C.M. Lopez-Vazquez, N.P. Van der Steen, P.N.L.
547 Lens, Ammonium removal mechanisms in a microalgal-bacterial sequencing-
548 batch photobioreactor at different solids retention times, *Algal Res.* 39 (2019)
549 101468. <https://doi.org/10.1016/j.algal.2019.101468>.
- 550 [100] M. Wang, H. Yang, S.J. Ergas, P. van der Steen, A novel shortcut nitrogen
551 removal process using an algal-bacterial consortium in a photo-sequencing batch
552 reactor (PSBR), *Water Res.* 87 (2015) 38–48.
553 <https://doi.org/10.1016/j.watres.2015.09.016>.
- 554 [101] D. Marín, E. Posadas, D. García, D. Puyol, R. Lebrero, R. Muñoz, Assessing the
555 potential of purple phototrophic bacteria for the simultaneous treatment of
556 piggery wastewater and upgrading of biogas, *Bioresour. Technol.* 281 (2019) 10–
557 17. <https://doi.org/10.1016/j.biortech.2019.02.073>.
- 558 [102] G. Guo, J. Guan, S. Sun, J. Liu, Y. Zhao, Nutrient and heavy metal removal from
559 piggery wastewater and CH₄ enrichment in biogas based on microalgae
560 cultivation technology under different initial inoculum concentration, *WATER*
561 *Environ. Res.* 92 (2020) 922–933. <https://doi.org/10.1002/wer.1287>.
- 562 [103] S. Arun, A. Sinharoy, K. Pakshirajan, P.N.L. Lens, Algae based microbial fuel
563 cells for wastewater treatment and recovery of value-added products, *Renew.*
564 *Sustain. Energy Rev.* 132 (2020) 110041.
565 <https://doi.org/10.1016/j.rser.2020.110041>.
- 566 [104] B. Zhang, W. Li, Y. Guo, Z. Zhang, W. Shi, F. Cui, P.N.L. Lens, J.H. Tay,
567 Microalgal-bacterial consortia: From interspecies interactions to biotechnological
568 applications, *Renew. Sustain. Energy Rev.* 118 (2020) 109563.
569 <https://doi.org/10.1016/j.rser.2019.109563>.
- 570 [105] J. Lee, J. Lee, S.K. Shukla, J. Park, T.K. Lee, Effect of algal inoculation on COD
571 and nitrogen removal, and indigenous bacterial dynamics in municipal
572 wastewater, *J. Microbiol. Biotechnol.* 26 (2016) 900–908.
573 <https://doi.org/10.4014/jmb.1512.12067>.
- 574 [106] B. Holmes, M.B. Paddock, J.S. VanderGheynst, B.T. Higgins, Algal
575 photosynthetic aeration increases the capacity of bacteria to degrade organics in
576 wastewater, *Biotechnol. Bioeng.* 117 (2020) 62–72.
577 <https://doi.org/10.1002/bit.27172>.
- 578 [107] M.M.M. Kuypers, H.K. Marchant, B. Kartal, The microbial nitrogen-cycling
579 network, *Nat. Publ. Gr.* (2018). <https://doi.org/10.1038/nrmicro.2018.9>.
- 580 [108] Y. Wang, S. Wang, L. Sun, Z. Sun, D. Li, Screening of a *Chlorella*-bacteria
581 consortium and research on piggery wastewater purification, *Algal Res.* 47
582 (2020). <https://doi.org/10.1016/j.algal.2020.101840>.
- 583 [109] T. Cai, S.Y. Park, Y. Li, Nutrient recovery from wastewater streams by
584 microalgae: Status and prospects, *Renew. Sustain. Energy Rev.* 19 (2013) 360–
585 369. <https://doi.org/10.1016/j.rser.2012.11.030>.
- 586 [110] P. Foladori, S. Petrini, M. Nessenzia, G. Andreottola, Enhanced nitrogen removal
587 and energy saving in a microalgal-bacterial consortium treating real municipal

- wastewater, *Water Sci. Technol.* 78 (2018) 174–182.
<https://doi.org/10.2166/wst.2018.094>.
- [111] A.S. Guerra-Renteria, M.A. García-Ramírez, C. Gómez-Hermosillo, A. Gómez-Guzmán, Y. González-García, O. González-Reynoso, Metabolic pathway analysis of nitrogen and phosphorus uptake by the consortium between *C. Vulgaris* and *P. Aeruginosa*, *Int. J. Mol. Sci.* 20 (2019).
<https://doi.org/10.3390/ijms20081978>.
- [112] C.C. Tang, Y. Tian, H. Liang, W. Zuo, Z.W. Wang, J. Zhang, Z.W. He, Enhanced nitrogen and phosphorus removal from domestic wastewater via algae-assisted sequencing batch biofilm reactor, *Bioresour. Technol.* (2018).
<https://doi.org/10.1016/j.biortech.2017.11.028>.
- [113] J. Zhou, Y. Lyu, M.L. Richlen, D.M. Anderson, Z. Cai, Quorum Sensing Is a Language of Chemical Signals and Plays an Ecological Role in Algal-Bacterial Interactions, *CRC. Crit. Rev. Plant Sci.* 35 (2016) 81–105.
<https://doi.org/10.1080/07352689.2016.1172461>.
- [114] A. Mitsutani, I. Yamasaki, H. Kitaguchi, J. Kato, S. Ueno, Y. Ishida, Analysis of algicidal proteins of a diatom-lytic marine bacterium *Pseudoalteromonas* sp. strain A25 by two-dimensional electrophoresis, *Phycologia*. 40 (2001) 286–291.
<https://doi.org/10.2216/i0031-8884-40-3-286.1>.
- [115] W.C. Fuqua, S.C. Winans, E.P. Greenberg, Quorum sensing in bacteria: The LuxR-LuxI family of cell density- responsive transcriptional regulators, *J. Bacteriol.* 176 (1994) 269–275. <https://doi.org/10.1128/jb.176.2.269-275.1994>.
- [116] S.A. Amin, L.R. Hmelo, H.M. Van Tol, B.P. Durham, L.T. Carlson, K.R. Heal, R.L. Morales, C.T. Berthiaume, M.S. Parker, B. Djunaedi, A.E. Ingalls, M.R. Parsek, M.A. Moran, E. V. Armbrust, Interaction and signalling between a cosmopolitan phytoplankton and associated bacteria, *Nature*. 522 (2015) 98–101.
<https://doi.org/10.1038/nature14488>.
- [117] L.A. Pagnussat, G. Maroniche, L. Curatti, C. Creus, Auxin-dependent alleviation of oxidative stress and growth promotion of *Scenedesmus obliquus* C1S by *Azospirillum brasilense*, *Algal Res.* 47 (2020).
<https://doi.org/10.1016/j.algal.2020.101839>.
- [118] L.E. de-Bashan, Y. Bashan, Increased growth of the microalga *Chlorella Vulgaris* when coimmobilized and cocultured in alginate beads with the plant-growth-promoting bacterium *Azospirillum Brasilense*, *Appl. Environ. Microbiol.* 66 (2000) 1527–1531. <https://doi.org/10.1128/AEM.66.4.1527-1531.2000>.
- [119] H.M. Najdenski, L.G. Gigova, I.I. Iliev, P.S. Pilarski, J. Lukavský, I. V. Tsvetkova, M.S. Ninova, V.K. Kussovski, Antibacterial and antifungal activities of selected microalgae and cyanobacteria, *Int. J. Food Sci. Technol.* 48 (2013) 1533–1540. <https://doi.org/10.1111/ijfs.12122>.
- [120] B. Zhang, Y. Guo, P.N.L. Lens, Z. Zhang, W. Shi, F. Cui, J.H. Tay, Effect of light intensity on the characteristics of algal-bacterial granular sludge and the role of N-acyl-homoserine lactone in the granulation, *Sci. Total Environ.* 659 (2019) 372–383. <https://doi.org/10.1016/j.scitotenv.2018.12.250>.
- [121] D. Zhou, C. Zhang, L. Fu, L. Xu, X. Cui, Q. Li, J.C. Crittenden, Responses of the Microalga *Chlorophyta* sp. to Bacterial Quorum Sensing Molecules (N-

- 633 Acylhomoserine Lactones): Aromatic Protein-Induced Self-Aggregation,
 634 Environ. Sci. Technol. 51 (2017) 3490–3498.
 635 <https://doi.org/10.1021/acs.est.7b00355>.
- 636 [122] B. Zhang, P.N.L. Lens, W. Shi, R. Zhang, Z. Zhang, Y. Guo, X. Bao, F. Cui, The
 637 attachment potential and N-acyl-homoserine lactone-based quorum sensing in
 638 aerobic granular sludge and algal-bacterial granular sludge, Appl. Microbiol.
 639 Biotechnol. 102 (2018) 5343–5353. <https://doi.org/10.1007/s00253-018-9002-9>.
- 640 [123] X. Ji, X. Luo, J. Zhang, D. Huang, Effects of exogenous vitamin B12 on nutrient
 641 removal and protein expression of algal-bacterial consortium, Environ. Sci.
 642 Pollut. Res. 28 (2021) 15954–15965. [https://doi.org/10.1007/s11356-020-11720-](https://doi.org/10.1007/s11356-020-11720-0)
 643 0.
- 644 [124] L.E. de-Bashan, V.K. Lebsky, J.P. Hernandez, J.J. Bustillos, Y. Bashan, Changes
 645 in the metabolism of the microalga *Chlorella vulgaris* when coimmobilized in
 646 alginate with the nitrogen-fixing *Phyllobacterium myrsinacearum*, Can. J.
 647 Microbiol. 46 (2000) 653–659. <https://doi.org/10.1139/w00-041>.
- 648 [125] C. Zhang, Q. Li, L. Fu, D. Zhou, J.C. Crittenden, Quorum sensing molecules in
 649 activated sludge could trigger microalgae lipid synthesis, Bioresour. Technol.
 650 263 (2018) 576–582. <https://doi.org/10.1016/j.biortech.2018.05.045>.
- 651 [126] C. Zhang, S.H. Ho, A. Li, L. Fu, D. Zhou, Co-culture of *Chlorella* and
 652 *Scenedesmus* could enhance total lipid production under bacteria quorum sensing
 653 molecule stress, J. Water Process Eng. 39 (2021) 101739.
 654 <https://doi.org/10.1016/j.jwpe.2020.101739>.
- 655 [127] K. Fukami, T. Nishijima, Y. Ishida, Stimulative and inhibitory effects of bacteria
 656 on the growth of microalgae, Hydrobiologia. 358 (1997) 185–191.
 657 https://doi.org/10.1007/978-94-017-2097-7_29.
- 658 [128] N. Rashid, W.-K. Park, T. Selvaratnam, Binary culture of microalgae as an
 659 integrated approach for enhanced biomass and metabolites productivity,
 660 wastewater treatment, and bioflocculation, Chemosphere. 194 (2018) 67–75.
 661 <https://doi.org/10.1016/j.chemosphere.2017.11.108>.
- 662 [129] L.P. Acurio, D.M. Salazar, A.F. Valencia, D.R. Robalino, A.C. Barona, F.C.
 663 Alvarez, C.A. Rodriguez, Antimicrobial potential of *Chlorella* algae isolated
 664 from stacked waters of the Andean Region of Ecuador, IOP Conf. Ser. Earth
 665 Environ. Sci. 151 (2018). <https://doi.org/10.1088/1755-1315/151/1/012040>.
- 666 [130] X. Chen, Z. Hu, Y. Qi, C. Song, G. Chen, The interactions of algae-activated
 667 sludge symbiotic system and its effects on wastewater treatment and lipid
 668 accumulation, Bioresour. Technol. (2019).
 669 <https://doi.org/10.1016/j.biortech.2019.122017>.
- 670 [131] L.M. González-González, L.E. De-Bashan, Toward the enhancement of
 671 microalgal metabolite production through microalgae–bacteria consortia, Biology
 672 (Basel). 10 (2021) 1–20. <https://doi.org/10.3390/biology10040282>.
- 673 [132] B.T. Higgins, I. Gennity, S. Samra, T. Kind, O. Fiehn, J.S. VanderGheynst,
 674 Cofactor symbiosis for enhanced algal growth, biofuel production, and
 675 wastewater treatment, Algal Res. 17 (2016) 308–315.
 676 <https://doi.org/10.1016/j.algal.2016.05.024>.
- 677 [133] S.N. Li, C. Zhang, F. Li, N.Q. Ren, S.H. Ho, Recent advances of algae-bacteria

- consortia in aquatic remediation, *Crit. Rev. Environ. Sci. Technol.* 0 (2022) 1–25.
<https://doi.org/10.1080/10643389.2022.2052704>.
- [134] M.B. Cooper, A.G. Smith, Exploring mutualistic interactions between microalgae and bacteria in the omics age, *Curr. Opin. Plant Biol.* 26 (2015) 147–153.
<https://doi.org/10.1016/j.pbi.2015.07.003>.
- [135] M.T. Nazari, J.F. Freitag, V.A.F. Cavanhi, L.M. Colla, Microalgae harvesting by fungal-assisted bioflocculation, *Rev. Environ. Sci. Biotechnol.* 19 (2020) 369–388. <https://doi.org/10.1007/s11157-020-09528-y>.
- [136] Z. Ding, I. Bourven, G. Guibaud, E.D. van Hullebusch, A. Panico, F. Pirozzi, G. Esposito, Role of extracellular polymeric substances (EPS) production in bioaggregation: application to wastewater treatment, *Appl. Microbiol. Biotechnol.* 99 (2015) 9883–9905. <https://doi.org/10.1007/s00253-015-6964-8>.
- [137] B. Zhang, W. Li, Y. Guo, Z. Zhang, W. Shi, F. Cui, P.N.L. Lens, J.H. Tay, Microalgal-bacterial consortia: From interspecies interactions to biotechnological applications, *Renew. Sustain. Energy Rev.* 118 (2020).
<https://doi.org/10.1016/j.rser.2019.109563>.
- [138] L. Ferro, M. Colombo, E. Posadas, C. Funk, R. Muñoz, Elucidating the symbiotic interactions between a locally isolated microalga *Chlorella Vulgaris* and its co-occurring bacterium *Rhizobium* sp. in synthetic municipal wastewater, *J. Appl. Phycol.* 31 (2019) 2299–2310. <https://doi.org/10.1007/s10811-019-1741-1>.
- [139] Q. Wang, W. Jin, X. Zhou, S. Guo, S.H. Gao, C. Chen, R. Tu, S.F. Han, J. Jiang, X. Feng, Growth enhancement of biodiesel-promising microalga *Chlorella pyrenoidosa* in municipal wastewater by polyphosphate-accumulating organisms, *J. Clean. Prod.* 240 (2019) 118148.
<https://doi.org/10.1016/j.jclepro.2019.118148>.
- [140] T. Kuba, A. Wachtmeister, M.C.M. Van Loosdrecht, J.J. Heijnen, Effect of nitrate on phosphorus release in biological phosphorus removal systems, in: *Water Sci. Technol.*, 1994. <https://doi.org/10.2166/wst.1994.0277>.
- [141] Y. Hu, X. Hao, M. van Loosdrecht, H. Chen, Enrichment of highly settleable microalgal consortia in mixed cultures for effluent polishing and low-cost biomass production, *Water Res.* 125 (2017) 11–22.
<https://doi.org/10.1016/j.watres.2017.08.034>.
- [142] N.C. Boelee, H. Temmink, M. Janssen, C.J.N. Buisman, R.H. Wijffels, Balancing the organic load and light supply in symbiotic microalgal – bacterial biofilm reactors treating synthetic municipal wastewater, *Ecol. Eng.* 64 (2014) 213–221. <https://doi.org/10.1016/j.ecoleng.2013.12.035>.
- [143] N.C. Boelee, H. Temmink, M. Janssen, C.J.N. Buisman, R.H. Wijffels, Nitrogen and phosphorus removal from municipal wastewater effluent using microalgal biofilms, *Water Res.* 45 (2011) 5925–5933.
<https://doi.org/10.1016/j.watres.2011.08.044>.
- [144] V. Ashok, A. Shriwastav, P. Bose, Nutrient Removal Using Algal-Bacterial Mixed Culture, *Appl. Biochem. Biotechnol.* (2014).
<https://doi.org/10.1007/s12010-014-1229-z>.
- [145] Y. Luo, P. Le-Clech, R.K. Henderson, Simultaneous microalgae cultivation and wastewater treatment in submerged membrane photobioreactors: A review, *Algal*

- 723 Res. 24 (2017) 425–437. <https://doi.org/10.1016/j.algal.2016.10.026>.
- 724 [146] J.S. Arcila, D. Céspedes, G. Buitrón, Influence of wavelength photoperiods and
725 N/P ratio on wastewater treatment with microalgae–bacteria, *Water Sci. Technol.*
726 00 (2021) 1–13. <https://doi.org/10.2166/wst.2021.257>.
- 727 [147] H.J. Choi, S.M. Lee, Effect of the N/P ratio on biomass productivity and nutrient
728 removal from municipal wastewater, *Bioprocess Biosyst. Eng.* 38 (2015) 761–
729 766. <https://doi.org/10.1007/s00449-014-1317-z>.
- 730 [148] S. Eixler, U. Karsten, U. Selig, Phosphorus storage in *Chlorella Vulgaris*
731 (*Trebouxioiophyceae*, *Chlorophyta*) cells and its dependence on phosphate supply,
732 *Phycologia*. 45 (2006) 53–60. <https://doi.org/10.2216/04-79.1>.
- 733 [149] A. Crimp, N. Brown, A. Shilton, Microalgal luxury uptake of phosphorus in
734 waste stabilization ponds - frequency of occurrence and high performing genera,
735 *Water Sci. Technol.* 78 (2018) 165–173. <https://doi.org/10.2166/wst.2017.632>.
- 736 [150] N. Powell, A. Shilton, S. Pratt, Y. Chisti, Luxury uptake of phosphorus by
737 microalgae in full-scale waste stabilisation ponds, *Water Sci. Technol.* (2011).
738 <https://doi.org/10.2166/wst.2011.116>.
- 739 [151] Y. Yang, X. Shi, W. Ballent, B.K. Mayer, Biological Phosphorus Recovery:
740 Review of Current Progress and Future Needs, *Water Environ. Res.* 89 (2017)
741 2122–2135. <https://doi.org/10.2175/106143017x15054988926424>.
- 742 [152] T. Mino, M.C.M. Van Loosdrecht, J.J. Heijnen, Microbiology and biochemistry
743 of the enhanced biological phosphate removal process, *Water Res.* 32 (1998)
744 3193–3207. [https://doi.org/10.1016/S0043-1354\(98\)00129-8](https://doi.org/10.1016/S0043-1354(98)00129-8).
- 745 [153] E.R. Coats, C.K. Brinkman, S. Lee, Characterizing and contrasting the microbial
746 ecology of laboratory and full-scale EBPR systems cultured on synthetic and real
747 wastewaters, *Water Res.* 108 (2017) 124–136.
748 <https://doi.org/10.1016/j.watres.2016.10.069>.
- 749 [154] L. Xing, L. Ren, B. Tang, G. Wu, Y. Guan, Dynamics of intracellular polymers
750 in enhanced biological phosphorus removal processes under different organic
751 carbon concentrations, *Biomed Res. Int.* 2013 (2013).
752 <https://doi.org/10.1155/2013/761082>.
- 753 [155] W. Huang, B. Li, C. Zhang, Z. Zhang, Z. Lei, B. Lu, B. Zhou, Effect of algae
754 growth on aerobic granulation and nutrients removal from synthetic wastewater
755 by using sequencing batch reactors, *Bioresour. Technol.* 179 (2015) 187–192.
756 <https://doi.org/10.1016/j.biortech.2014.12.024>.
- 757 [156] T.J. Belli, J. Karla, B. Bernardelli, R. Emanuely, J.P. Bassin, M. Cristina, S.
758 Amaral, F.R. Lapolli, Effect of solids retention time on nitrogen and phosphorus
759 removal from municipal wastewater in a sequencing batch membrane bioreactor,
760 *Environ. Technol.* 38 (2017) 806–815.
761 <https://doi.org/10.1080/09593330.2016.1212934>.
- 762 [157] S. Zhu, L. Qin, P. Feng, C. Shang, Z. Wang, Z. Yuan, Treatment of low C/N ratio
763 wastewater and biomass production using co-culture of *Chlorella Vulgaris* and
764 activated sludge in a batch photobioreactor, *Bioresour. Technol.* 274 (2019) 313–
765 320. <https://doi.org/10.1016/j.biortech.2018.10.034>.
- 766 [158] N. Artan, R. Tasli, N. Özgür, D. Orhon, The fate of phosphate under anoxic
767 conditions in biological nutrient removal activated sludge systems, *Biotechnol.*

- 768 Lett. 20 (1998) 1085–1090. <https://doi.org/10.1023/A:1005419408097>.
- 769 [159] F. Lananan, S.H. Abdul Hamid, W.N.S. Din, N. Ali, H. Khatoon, A. Jusoh, A.
770 Endut, Symbiotic bioremediation of aquaculture wastewater in reducing
771 ammonia and phosphorus utilizing Effective Microorganism (EM-1) and
772 microalgae (*Chlorella* sp.), *Int. Biodeterior. Biodegrad.* 95 (2014) 127–134.
773 <https://doi.org/10.1016/j.ibiod.2014.06.013>.
- 774 [160] J. García, R. Mujeriego, M. Hernández-Mariné, High rate algal pond operating
775 strategies for urban wastewater nitrogen removal, *J. Appl. Phycol.* 12 (2000)
776 331–339. <https://doi.org/10.1023/A:1008146421368>.
- 777 [161] N. Nirmalakhandan, T. Selvaratnam, D. Tchinda, I.S.A. Abeysiriwardana-
778 Arachchige, H.M.K. Delanka-Pedige, S.P. Munasinghe-Arachchige, Algal
779 wastewater treatment : Photoautotrophic vs . mixotrophic processes, *Algal Res.*
780 41 (2019) 101569. <https://doi.org/10.1016/j.algal.2019.101569>.
- 781 [162] B. Picot, H. El Halouani, C. Casellas, S. Moersidik, J. Bontoux, Nutrient removal
782 by high rate pond system in a Mediterranean climate (France), *Water Sci.*
783 *Technol.* 23 (1991) 1535–1541. <https://doi.org/10.2166/wst.1991.0607>.
- 784 [163] E. Posadas, M. del M. Morales, C. Gomez, F.G. Ación, R. Muñoz, Influence of
785 pH and CO₂ source on the performance of microalgae-based secondary domestic
786 wastewater treatment in outdoors pilot raceways, *Chem. Eng. J.* 265 (2015) 239–
787 248. <https://doi.org/10.1016/j.cej.2014.12.059>.
- 788 [164] R. Craggs, D. Sutherland, H. Campbell, Hectare-scale demonstration of high rate
789 algal ponds for enhanced wastewater treatment and biofuel production, *J. Appl.*
790 *Phycol.* 24 (2012) 329–337. <https://doi.org/10.1007/s10811-012-9810-8>.
- 791 [165] J.B.K. Park, R.J. Craggs, Wastewater treatment and algal production in high rate
792 algal ponds with carbon dioxide addition, *Water Sci. Technol.* 61 (2010) 633–
793 639. <https://doi.org/10.2166/wst.2010.951>.
- 794 [166] J.B.K. Park, R.J. Craggs, Nutrient removal in wastewater treatment high rate
795 algal ponds with carbon dioxide addition, *Water Sci. Technol.* (2011).
796 <https://doi.org/10.2166/wst.2011.114>.
- 797 [167] S. Van Den Hende, H. Vervaeren, S. Desmet, N. Boon, Bioflocculation of
798 microalgae and bacteria combined with flue gas to improve sewage treatment, *N.*
799 *Biotechnol.* 29 (2011) 23–31. <https://doi.org/10.1016/j.nbt.2011.04.009>.
- 800 [168] V.M. Vadivelu, J. Keller, Z. Yuan, Free ammonia and free nitrous acid inhibition
801 on the anabolic and catabolic processes of *Nitrosomonas* and *Nitrobacter*, *Water*
802 *Science and Technology*, 2007. <https://doi.org/10.2166/wst.2007.612>.
- 803 [169] A. Abeliovich, Y. Azov, Toxicity of ammonia to algae in sewage oxidation
804 ponds, *Appl. Environ. Microbiol.* (1976). [https://doi.org/10.1128/aem.31.6.801-](https://doi.org/10.1128/aem.31.6.801-806.1976)
805 [806.1976](https://doi.org/10.1128/aem.31.6.801-806.1976).
- 806 [170] S. Rossi, R. Díez-Montero, E. Rueda, F. Castillo Cascino, K. Parati, J. García, E.
807 Ficara, Free ammonia inhibition in microalgae and cyanobacteria grown in
808 wastewaters: Photo-respirometric evaluation and modelling, *Bioresour. Technol.*
809 305 (2020). <https://doi.org/10.1016/j.biortech.2020.123046>.
- 810 [171] F. Casagli, S. Rossi, J.P. Steyer, O. Bernard, E. Ficara, Balancing Microalgae and
811 Nitrifiers for Wastewater Treatment: Can Inorganic Carbon Limitation Cause an
812 Environmental Threat?, *Environ. Sci. Technol.* 55 (2021) 3940–3955.

- 813 <https://doi.org/10.1021/acs.est.0c05264>.
- 814 [172] B.L. Mellbye, A. Giguere, F. Chaplen, P.J. Bottomley, L.A. Sayavedra-Soto,
815 Steady-state growth under inorganic carbon limitation conditions increases
816 energy consumption for maintenance and enhances nitrous oxide production in
817 *Nitrosomonas europaea*, *Appl. Environ. Microbiol.* 82 (2016) 3310–3318.
818 <https://doi.org/10.1128/AEM.00294-16>.
- 819 [173] S. Solomon, I.P. on C. Change., I.P. on C. Change., W.G. I., *Climate change*
820 *2007 : the physical science basis : contribution of Working Group I to the Fourth*
821 *Assessment Report of the Intergovernmental Panel on Climate Change*,
822 Cambridge University Press, Cambridge; New York, 2007.
- 823 [174] B.-G. Ryu, J. Kim, J.I. Han, J.W. Yang, Feasibility of using a microalgal-
824 bacterial consortium for treatment of toxic coke wastewater with concomitant
825 production of microbial lipids, *Bioresour. Technol.* 225 (2017) 58–66.
826 <https://doi.org/10.1016/j.biortech.2016.11.029>.
- 827 [175] D.L. Sutherland, C. Howard-Williams, M.H. Turnbull, P.A. Broady, R.J. Craggs,
828 The effects of CO₂ addition along a pH gradient on wastewater microalgal
829 photo-physiology, biomass production and nutrient removal, *Water Res.* 70
830 (2015) 9–26. <https://doi.org/10.1016/j.watres.2014.10.064>.
- 831 [176] I. de Godos, V.A. Vargas, H.O. Guzmán, R. Soto, B. García, P.A. García-Encina,
832 R. Muñoz, Assessing carbon and nitrogen removal in a novel anoxic-aerobic
833 cyanobacterial-bacterial photobioreactor configuration with enhanced biomass
834 sedimentation, *Water Res.* 61 (2014) 77–85.
835 <https://doi.org/10.1016/j.watres.2014.04.050>.
- 836 [177] C. Alcántara, J.M. Domínguez, D. García, S. Blanco, R. Pérez, P.A. García-
837 Encina, R. Muñoz, Evaluation of wastewater treatment in a novel anoxic-aerobic
838 algal-bacterial photobioreactor with biomass recycling through carbon and
839 nitrogen mass balances, *Bioresour. Technol.* 191 (2015) 173–186.
840 <https://doi.org/10.1016/j.biortech.2015.04.125>.
- 841 [178] C. Vergara, R. Muñoz, J.L. Campos, M. Seeger, D. Jeison, Influence of light
842 intensity on bacterial nitrifying activity in algal-bacterial photobioreactors and its
843 implications for microalgae-based wastewater treatment, *Int. Biodeterior.*
844 *Biodegrad.* 114 (2016) 116–121. <https://doi.org/10.1016/j.ibiod.2016.06.006>.
- 845 [179] N.D. Manser, M. Wang, S.J. Ergas, J.R. Mihelcic, A. Mulder, J. Van de
846 Vossenberg, J.B. Van Lier, P. Van der Steen, Biological Nitrogen Removal in a
847 Photosequencing Batch Reactor with an Algal-Nitrifying Bacterial Consortium
848 and Anammox Granules, *Environ. Sci. Technol. Lett.* 3 (2016) 175–179.
849 <https://doi.org/10.1021/acs.estlett.6b00034>.
- 850 [180] S.S. Shalini, K. Joseph, Start-up of the SHARON and ANAMMOX process in
851 landfill bioreactors using aerobic and anaerobic ammonium oxidising biomass,
852 *Bioresour. Technol.* 149 (2013) 474–485.
853 <https://doi.org/10.1016/j.biortech.2013.09.104>.
- 854 [181] K. Egli, U. Fanger, P.J.J. Alvarez, H. Siegrist, J.R. Van der Meer, A.J.B.
855 Zehnder, Enrichment and characterization of an anammox bacterium from a
856 rotating biological contactor treating ammonium-rich leachate, *Arch. Microbiol.*
857 175 (2001) 198–207. <https://doi.org/10.1007/s002030100255>.

- [182] M. Henze, W. Guher, T. Mino, M.C.M. van Loosdrecht, Activated sludge models ASM1, ASM2, ASM2D and ASM3, *J. Chem. Inf. Model.* 53 (2008) 287. <https://doi.org/10.1017/CBO9781107415324.004>.
- [183] D.J. Batstone, J. Keller, I. Angelidaki, S. V Kalyuzhnyi, S.G. Pavlostathis, A. Rozzi, W.T.M. Sanders, H. Siegrist, V.A. Vavilin, Industrial applications of the IWA anaerobic digestion model No. 1 (ADM1), *Water Sci. Technol.* 47 (2003) 199–206. <https://doi.org/10.2166/wst.2003.0647>.
- [184] M.C.M. Van Loosdrecht, C.M. Lopez-Vazquez, S.C.F. Meijer, C.M. Hooijmans, D. Brdjanovic, Twenty-five years of ASM1: Past, present and future of wastewater treatment modelling, *J. Hydroinformatics.* 17 (2015) 697–718. <https://doi.org/10.2166/hydro.2015.006>.
- [185] D.J. Batstone, J. Keller, J.P. Steyer, A review of ADM1 extensions, applications, and analysis: 2002–2005, *Water Sci. Technol.* 54 (2006) 1–10. <https://doi.org/10.2166/wst.2006.520>.
- [186] B.D. Shoener, S.M. Schramm, F. Béline, O. Bernard, B.G. Plósz, S. Snowling, J. Steyer, D. Wágner, J.S. Guest, Microalgae and cyanobacteria modeling in water resource recovery facilities: A critical review, *Water Res.* X. (2018) 100024. <https://doi.org/10.1016/j.wroa.2018.100024>.
- [187] P.A. Vanrolleghem, C. Rosen, U. Zaher, J. Copp, L. Benedetti, E. Ayesa, U. Jeppsson, Continuity-based interfacing of models for wastewater systems described by Petersen matrices, *Water Sci. Technol.* 52 (2005) 493–500. <https://doi.org/10.2166/wst.2005.0558>.
- [188] A. Solimeno, L. Parker, T. Lundquist, J. García, Integral microalgae-bacteria model (BIO _ ALGAE): Application to wastewater high rate algal ponds, *Sci. Total Environ.* 601–602 (2017) 646–657. <https://doi.org/10.1016/j.scitotenv.2017.05.215>.
- [189] P. Reichert, D. Borchardt, M. Henze, W. Rauch, P. Shanahan, L. Somlyódy, P. Vanrolleghem, River Water Quality Model no. 1 (RWQM1);, *Water Sci. Technol.* 43 (2001) 11–30. <https://doi.org/10.2166/wst.2001.0241>.
- [190] G. Wolf, C. Picioreanu, M.C.M. van Loosdrecht, Kinetic modeling of phototrophic biofilms: The PHOBIA model, *Biotechnol. Bioeng.* 97(5) (2007) 1064–1079.
- [191] T. Gehring, J.D. Silva, O. Kehl, A.B. Castilhos Jr., R.H.R. Costa, F. Uhlenhut, J. Alex, H. Horn, M. Wichern, Modelling waste stabilisation ponds with an extended version of ASM3, *Water Sci. Technol.* 61 (2010) 713–720. <https://doi.org/10.2166/wst.2010.954>.
- [192] N. Broekhuizen, J.B.K. Park, G.B. McBride, R.J. Craggs, Modification, calibration and verification of the IWA River Water Quality Model to simulate a pilot-scale high rate algal pond, *Water Res.* 46 (2012) 2911–2926. <https://doi.org/10.1016/j.watres.2012.03.011>.
- [193] J.D. Muñoz Sierra, C. Picioreanu, M.C.M. Van Loosdrecht, Modeling phototrophic biofilms in a plug-flow reactor, *Water Sci. Technol.* 70 (2014) 1261–1270. <https://doi.org/10.2166/wst.2014.368>.
- [194] J. Zambrano, I. Krustok, E. Nehrenheim, B. Carlsson, A simple model for algae-bacteria interaction in photo-bioreactors, *Algal Res.* 19 (2016) 155–161.

- 903 <https://doi.org/10.1016/j.algal.2016.07.022>.
- 904 [195] L.T. Arashiro, A.M. Rada-Ariza, M. Wang, P. van der Steen, S.J. Ergas,
905 Modelling shortcut nitrogen removal from wastewater using an algal bacterial
906 consortium, (2017) 1–12. <https://doi.org/10.2166/wst.2016.561>.
- 907 [196] E.M. Grima, F.G. Camacho, J.A.S. Pérez, J.M.F. Sevilla, F.G.A. Fernández, A.C.
908 Gómez, A mathematical model of microalgal growth in light-limited chemostat
909 culture, *J. Chem. Technol. Biotechnol.* 61 (1994) 167–173.
910 <https://doi.org/https://doi.org/10.1002/jctb.280610212>.
- 911 [197] S. Park, W. Bae, J. Chung, S.-C. Baek, Empirical model of the pH dependence of
912 the maximum specific nitrification rate, *Process Biochem.* 42 (2007) 1671–1676.
913 <https://doi.org/https://doi.org/10.1016/j.procbio.2007.09.010>.
- 914 [198] A. Packer, Y. Li, T. Andersen, Q. Hu, Y. Kuang, M. Sommerfeld, Growth and
915 neutral lipid synthesis in green microalgae: A mathematical model, *Bioresour.*
916 *Technol.* 102 (2011) 111–117.
917 <https://doi.org/https://doi.org/10.1016/j.biortech.2010.06.029>.
- 918 [199] B. Palabhanvi, V. Kumar, M. Muthuraj, D. Das, Preferential utilization of
919 intracellular nutrients supports microalgal growth under nutrient starvation:
920 multi-nutrient mechanistic model and experimental validation., *Bioresour.*
921 *Technol.* 173 (2014) 245–255.
- 922 [200] X. Bai, P. Lant, S. Pratt, The contribution of bacteria to algal growth by carbon
923 cycling, *Biotechnol. Bioeng.* 112 (2015) 688–695.
924 <https://doi.org/https://doi.org/10.1002/bit.25475>.
- 925 [201] R.L.L. Ribeiro, J.V.C. Vargas, A.B. Mariano, J.C. Ordonez, The experimental
926 validation of a large-scale compact tubular microalgae photobioreactor model,
927 *Int. J. Energy Res.* 41 (2017) 2221–2235.
928 <https://doi.org/https://doi.org/10.1002/er.3784>.
- 929 [202] R. Rodríguez, J.J. Espada, J. Moreno, G. Vicente, L.F. Bautista, V. Morales, A.
930 Sánchez-Bayo, J. Dufour, Environmental analysis of Spirulina cultivation and
931 biogas production using experimental and simulation approach, *Renew. Energy.*
932 129 (2017) 724–732. <https://doi.org/https://doi.org/10.1016/j.renene.2017.05.076>.
- 933 [203] V.O. Adesanya, M.P. Davey, S.A. Scott, A.G. Smith, Kinetic modelling of
934 growth and storage molecule production in microalgae under mixotrophic and
935 autotrophic conditions, *Bioresour. Technol.* 157 (2014) 293–304.
936 <https://doi.org/https://doi.org/10.1016/j.biortech.2014.01.032>.
- 937 [204] G.M. Figueroa-Torres, J.K. Pittman, C. Theodoropoulos, Kinetic modelling of
938 starch and lipid formation during mixotrophic, nutrient-limited microalgal
939 growth, *Bioresour. Technol.* 241 (2017) 868–878.
940 <https://doi.org/https://doi.org/10.1016/j.biortech.2017.05.177>.
- 941 [205] W. Gujer, *Systems analysis for water technology*, 2008.
- 942 [206] P. Foladori, S. Petrini, G. Andreottola, How suspended solids concentration
943 affects nitrification rate in microalgal-bacterial photobioreactors without external
944 aeration, *Heliyon.* 6 (2020) e03088.
945 <https://doi.org/10.1016/j.heliyon.2019.e03088>.
- 946 [207] P. van der Steen, K. Rahsilawati, A.M. Rada-Ariza, C.M. Lopez-Vazquez, P.N.L.
947 Lens, A new photo-activated sludge system for nitrification by an algal-bacterial

- 948 consortium in a photo-bioreactor with biomass recycle, (2015) 443–450.
949 <https://doi.org/10.2166/wst.2015.205>.
- 950 [208] M. Gross, Z. Wen, Yearlong evaluation of performance and durability of a pilot-
951 scale Revolving Algal Biofilm (RAB) cultivation system, *Bioresour. Technol.*
952 171 (2014) 50–58. <https://doi.org/10.1016/j.biortech.2014.08.052>.
- 953 [209] M. Kesaano, R.C. Sims, Algal biofilm based technology for wastewater
954 treatment, *Algal Res.* 5 (2014) 231–240.
955 <https://doi.org/10.1016/j.algal.2014.02.003>.
- 956 [210] D. Hoh, S. Watson, E. Kan, Algal biofilm reactors for integrated wastewater
957 treatment and biofuel production: A review, *Chem. Eng. J.* 287 (2016) 466–473.
958 <https://doi.org/10.1016/j.cej.2015.11.062>.
- 959 [211] W. Blanken, M. Janssen, M. Cuaresma, Z. Libor, T. Bhaiji, R.H. Wijffels,
960 Biofilm growth of *Chlorella sorokiniana* in a rotating biological contactor based
961 photobioreactor, *Biotechnol. Bioeng.* 111 (2014) 2436–2445.
962 <https://doi.org/10.1002/bit.25301>.
- 963 [212] L.B. Christenson, R.C. Sims, Rotating algal biofilm reactor and spool harvester
964 for wastewater treatment with biofuels by-products, *Biotechnol. Bioeng.* 109
965 (2012) 1674–1684. <https://doi.org/10.1002/bit.24451>.
- 966 [213] J. Shi, B. Podola, M. Melkonian, Application of a prototype-scale twin-layer
967 photobioreactor for effective N and P removal from different process stages of
968 municipal wastewater by immobilized microalgae, *Bioresour. Technol.* 154
969 (2014) 260–266. <https://doi.org/10.1016/j.biortech.2013.11.100>.
- 970 [214] M. Morales, H. Bonnefond, O. Bernard, Rotating algal biofilm versus planktonic
971 cultivation: LCA perspective, *J. Clean. Prod.* 257 (2020) 120547.
972 <https://doi.org/10.1016/j.jclepro.2020.120547>.
- 973 [215] M. Gross, V. Mascarenhas, Z. Wen, Evaluating algal growth performance and
974 water use efficiency of pilot-scale revolving algal biofilm (RAB) culture systems,
975 *Biotechnol. Bioeng.* 112 (2015) 2040–2050. <https://doi.org/10.1002/bit.25618>.
- 976 [216] M. Gross, D. Jarboe, Z. Wen, Biofilm-based algal cultivation systems, *Appl.*
977 *Microbiol. Biotechnol.* 99 (2015) 5781–5789. [https://doi.org/10.1007/s00253-](https://doi.org/10.1007/s00253-015-6736-5)
978 015-6736-5.
- 979 [217] L.R. de Assis, M.L. Calijuri, E. de A. Do Couto, P.P. Assemany, Microalgal
980 biomass production and nutrients removal from domestic sewage in a hybrid
981 high-rate pond with biofilm reactor, *Ecol. Eng.* 106 (2017) 191–199.
982 <https://doi.org/10.1016/j.ecoleng.2017.05.040>.
- 983 [218] L. Rodrigues de Assis, M.L. Calijuri, P.P. Assemany, T.A. Silva, J.S. Teixeira,
984 Innovative hybrid system for wastewater treatment: High-rate algal ponds for
985 effluent treatment and biofilm reactor for biomass production and harvesting, *J.*
986 *Environ. Manage.* 274 (2020) 111183.
987 <https://doi.org/10.1016/j.jenvman.2020.111183>.
- 988 [219] G. Quijano, J.S. Arcila, G. Buitrón, Microalgal-bacterial aggregates: Applications
989 and perspectives for wastewater treatment, *Biotechnol. Adv.* 35 (2017) 772–781.
990 <https://doi.org/10.1016/j.biotechadv.2017.07.003>.
- 991 [220] K. Milferstedt, W.C. Kuo-Dahab, C.S. Butler, J. Hamelin, A.S. Abouhend, K.
992 Stauch-White, A. McNair, C. Watt, B.I. Carbajal-González, S. Dolan, C. Park,

993 The importance of filamentous cyanobacteria in the development of oxygenic
994 photogranules, Sci. Rep. 7 (2017) 1–16. <https://doi.org/10.1038/s41598-017->
995 16614-9.
996
997
998
999
1000
1001