



HAL
open science

Machine learning-based prediction of impulse control disorders in Parkinson's disease from clinical and genetic data

Johann Faouzi, Samir Bekadar, Fanny Artaud, Alexis Elbaz, Graziella Mangone, Olivier Colliot, Jean-Christophe Corvol

► To cite this version:

Johann Faouzi, Samir Bekadar, Fanny Artaud, Alexis Elbaz, Graziella Mangone, et al.. Machine learning-based prediction of impulse control disorders in Parkinson's disease from clinical and genetic data. IEEE Open Journal of Engineering in Medicine and Biology, 2022, 3, pp.96 - 107. 10.1109/OJEMB.2022.3178295 . hal-03680714v2

HAL Id: hal-03680714

<https://inria.hal.science/hal-03680714v2>

Submitted on 16 Jun 2022






HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Machine Learning-Based Prediction of Impulse Control Disorders in Parkinson's Disease From Clinical and Genetic Data

Johann Faouzi , Samir Bekadar, Fanny Artaud , Alexis Elbaz , Graziella Mangone, Olivier Colliot , *Member, IEEE*, and Jean-Christophe Corvol 

Abstract—Goal: Impulse control disorders (ICDs) are frequent non-motor symptoms occurring during the course of Parkinson's disease (PD). The objective of this study was to estimate the predictability of the future occurrence of these disorders using longitudinal data, the first study using cross-validation and replication in an independent cohort. **Methods:** We used data from two longitudinal PD cohorts (training set: PPMI, Parkinson's Progression Markers Initiative; test set: DIGPD, Drug Interaction With Genes in Parkinson's Disease). We included 380 PD subjects from PPMI and 388 PD subjects from DIGPD, with at least two visits and with clinical and genetic data available, in our analyses. We trained three logistic regressions and a recurrent neural network to predict ICDs at the next visit using clinical risk factors and genetic variants previously associated

with ICDs. We quantified performance using the area under the receiver operating characteristic curve (ROC AUC) and average precision. We compared these models to a trivial model predicting ICDs at the next visit with the status at the most recent visit. **Results:** The recurrent neural network (PPMI: 0.85 [0.80 – 0.90], DIGPD: 0.802 [0.78 – 0.83]) was the only model to be significantly better than the trivial model (PPMI: ROC AUC = 0.75 [0.69 – 0.81]; DIGPD: 0.78 [0.75 – 0.80]) on both cohorts. We showed that ICDs in PD can be predicted with better accuracy with a recurrent neural network model than a trivial model. The improvement in terms of ROC AUC was higher on PPMI than on DIGPD data, but not clinically relevant in both cohorts. **Conclusions:** Our results indicate that machine learning methods are potentially useful for predicting ICDs, but further works are required to reach clinical relevance.

Manuscript received February 8, 2022; revised April 14, 2022, May 20, 2022, and May 24, 2022; accepted May 24, 2022. Date of publication May 27, 2022; date of current version June 15, 2022. This work was supported in part by the Program Investissements d'avenir under Grant ANR-10-IAIHU-06, in part by the French Government under management of Agence Nationale de la Recherche as part of the Investissements d'avenir Program under Grant ANR-19-P3IA-0001 (PRAIRIE 3IA Institute), in part by the ERA-Net – ERACoSysMed Program through PD-Strat Project, in part by the ICM Big Brain Theory Program through Project PredictICD, in part by the Inria Project Lab Program through Project Neuromarkers, and in part by the European Union H2020 Programme part under Grant 826421, through Project TVB-Cloud. The PPMI – a public-private partnership – was supported by the Michael J. Fox Foundation for Parkinson's Research and funding partners (list of the full names of all of the PPMI funding partners can be found at www.ppmi-info.org/fundingpartners). The DIGPD cohort was supported in part by Assistance Publique Hôpitaux de Paris, and in part by grants from the French Ministry of Health through Programme Hospitalier de Recherche Clinique under Grant AOR0810. The review of this article was arranged by Associate Editor Anat Mirelman. (*Corresponding author: Jean-Christophe Corvol.*)

Johann Faouzi and Olivier Colliot are with the Sorbonne Université, Institut du Cerveau - Paris Brain Institute - ICM, CNRS, Inria, Inserm, AP-HP, Hôpital de la Pitié Salpêtrière, F-75013 Paris, France (e-mail: johann.faouzi@gmail.com; olivier.colliot@cnrs.fr).

Samir Bekadar, Graziella Mangone, and Jean-Christophe Corvol are with the Department of Neurology, Paris Brain Institute, Inserm, CNRS, Sorbonne Université, Assistance Publique Hôpitaux de Paris, Centre d'Investigation Clinique Neurosciences, Hôpital Pitié-Salpêtrière, F-75013 Paris, France (e-mail: samir.bekadar@icm-institute.org; graziella.mangone@icm-institute.org; jean-christophe.corvol@aphp.fr).

Fanny Artaud and Alexis Elbaz are with the Université Paris-Saclay, UVSQ, Université Paris-Sud, Inserm, Gustave Roussy, Équipe "Exposome et Hérité", CESP, 94807 Villejuif, France (e-mail: fanny.artaud@inserm.fr; alexis.elbaz@inserm.fr).

This article has supplementary downloadable material available at <https://doi.org/10.1109/OJEMB.2022.3178295>, provided by the authors. Digital Object Identifier 10.1109/OJEMB.2022.3178295

Index Terms—Impulse control disorders, machine learning, Parkinson's disease, precision medicine.

Impact Statement—Impulse control disorders, leading to high impairment of the quality of life of subjects with Parkinson's disease, are difficult to predict. Further studies are required to try to improve the predictive performance of machine learning-based models.

I. INTRODUCTION

ALTHOUGH Parkinson's disease (PD) is mostly known for its motor symptoms, numerous non-motor symptoms have been reported to occur during the course of the disease [1]. Impulse control disorders (ICDs) are psychiatric disorders characterized by the failure to resist an impulse, and unsuccessful attempts to control specific behaviors [2]. ICDs and related disorders are frequent in PD with a prevalence ranging from 15–20% in cross-sectional studies [3], [4], an incidence estimated to around 10% per year [5], [6], and a cumulative incidence reaching almost 50% after 5 years of disease duration in longitudinal studies [5]. PD patients with disease duration greater than 5 years are also subject to these disorders [7]. The four most common ICDs in PD are pathological gambling, compulsive eating, hypersexuality, and compulsive shopping, but other frequent ICDs include punding and hobbyism, and the prevalence of each ICD, in particular pathological gambling, highly varies between different cultures [8]. ICDs are associated with reduced quality of life, strained interpersonal relationships, increased caregiver burden, and require prompt

addressing [9]–[11]. Several case reports suggest that partial and total discontinuations of dopamine agonist (DA) treatment leads to a resolution of ICDs [12], [13].

Many factors have been associated with ICDs in PD, including socio-demographic, clinical and genetic biomarkers [14]. In particular, men tend to develop more pathological gambling and hypersexuality disorders while women develop more compulsive buying and eating disorders [15]. A younger age has been associated with ICDs in PD in numerous studies [4], [16]–[18]. Anxiety [18]–[20], depression [16], [20], and rapid eye movement (REM) sleep behavior disorders [21], [22] have also been correlated to ICDs. Dopamine replacement therapy has been shown to be the main risk factor for ICD. Both levodopa and dopamine agonists have been associated with ICDs, but with a stronger and higher association with dopamine agonists. Finally, associations between ICDs and several single-nucleotide polymorphisms (SNPs) in dopamine signaling pathway genes have been suggested [23]–[28].

The predictive performance of these factors altogether has been underexplored. Only three studies reported predictions at the patient level [24], [29], [30]. In all three studies, authors trained a logistic regression using clinical and genetic data, and measured its predictive performance using the area under receiver operating characteristic (ROC) curve (ROC AUC). None of these studies had cross-validation or a replication cohort, altering the confidence in the reported performance [31].

Our main objective was to predict ICDs from clinical and genetic data using machine learning approaches. We utilized two longitudinal cohorts to train and cross-validate the models on one cohort, but also assess the generalization capability of these models on the other cohort. The objective was to predict the risk of ICDs at the next visit, knowing the clinical history of the patient and their genotyping data.

II. MATERIALS AND METHODS

A. Populations

We used data from two research cohorts: the Parkinson's Progression Markers Initiative (PPMI) database and the Drug Interaction With Genes in Parkinson's Disease (DIGPD) study.

PPMI (<https://www.ppmi-info.org>) is a multicenter observational clinical study using advanced imaging, biologic sampling, and clinical and behavioral assessments to identify biomarkers of PD progression [32]. Data was gathered during face-to-face visits every 6–12 months. PD subjects were de novo and drug-naive at baseline. We downloaded the clinical and genetic data from the PPMI database (<https://www.ppmi-info.org/data>) on the 17th of October, 2019.

DIGPD is a French multicenter longitudinal cohort with annual follow-up of PD patients [5]. Eligible criteria consist in recent PD diagnosis (UK Parkinson's Disease Society Brain Bank criteria) with disease duration less than 5 years at recruitment. Data was gathered during face-to-face visits every 12 months following standard procedures.

Both studies were conducted according to good clinical practice, obtained approval from local ethic committees and regulatory authorities, and all patients provided informed consent prior to inclusion.

B. Participants and Clinical Measurements

Inclusion criteria consisted of having:

- 1) a PD diagnosis,
- 2) a baseline visit and at least another visit,
- 3) clinical and genetic data available, and
- 4) PD medication taken available.

We included socio-demographics and clinical variables that have been associated with ICDs in the literature: age of PD onset, length of follow-up, sex, past ICDs, continuous scales of anxiety, depression and REM sleep, and the motor exam (part III) of the Movement Disorders Society-sponsored revision of the Unified Parkinson's Disease Rating Scale (MDS-UPDRS). ICDs were assessed at each visit using the Questionnaire for Impulsive-Compulsive Disorders in Parkinson's Disease - Rating Scale [33] in PPMI, and through semi-structured interviews by a movement disorder specialist in DIGPD. We standardized each feature since some of them were assessed with different scales and because it is a common requirement for most machine learning estimators.

We took into account PD medication with three binary variables corresponding to the main classes of treatment (levodopa, dopamine agonists, others) and we derived more specific variables for dopamine agonists: mean daily, maximum daily and total doses (expressed in levodopa equivalent) and cumulative duration.

C. Genetic Variants

In absence of genome-wide association study on ICDs in PD, we considered 50 genetic variants selected as previously described: 20 variants from 16 genes involved in dopamine, serotonin, glutamate, norepinephrine and opioid systems and previously associated with ICD in PD or in the general population [23]; 30 additional variants from 10 genes differentially expressed after an acute challenge of levodopa in the striatum in a mouse model of dopamine denervation [34].

Genotyping data were collected using NeuroX [35] arrays in PPMI (267,607 variants measured), and Illumina Multi-Ethnic Genotyping Arrays in DIGPD (1,779,819 variants). We excluded variants with missing rates greater than 2% and variants deviating from Hardy-Weinberg equilibrium ($p < 10^{-8}$). We excluded related individuals (third-degree family relationships), individuals with mismatch between reported sex and genetically determined sex, and individuals with outlying heterozygosity (± 3 standard deviation). We imputed missing SNPs using the Michigan Imputation Server [36] for PPMI and the Sanger Imputation Server [37] for DIGPD, using the reference panel of the Haplotype Reference Consortium (release 1.1) [37]. We filtered variants based on their imputation quality ($r^2 > 0.6$ for PPMI, INFO score > 0.9 for DIGPD) in order to only include variants imputed with high quality.

D. Data Processing

Processing genetic data and extracting variants of interest matching inclusion criteria was performed using the PLINK [38] software. Processing of the different text-like files was performed using the pandas [39] and NumPy [40] Python packages. Missing values were imputed in a forward-fill fashion: for a given subject and a given feature, missing values were imputed using the most recent non-missing value for this subject and this feature. Baseline missing values were imputed using the mean baseline values on the training set. We used this simple approach because only a small percentage of data from a small subset of variables (anxiety, depression, REM sleep, and motor exam) was imputed and because it can be applied to any scale without any training.

E. Machine Learning Algorithms

We investigated five standard machine learning algorithms implemented in the scikit-learn [41] and XGBoost [42] Python packages: logistic regression, support vector machines with a linear kernel and with a radial basis function kernel [43], [44], random forest [45] and gradient tree boosting [46], [47]. These algorithms expect a fixed number of features as input. In order to deal with varying numbers of visits, we reduced all the previous visits into one “summary” visit using a convex combination. A convex combination is a linear combination such that the weights are all non-negative and sum to one. The weights indicate how much each visit contributes to this “summary” visit. A weight of 1 for the first visit means that the “summary” visit is simply the baseline visit, while a weight of 1 for the latest visit means that the “summary” visit is simply the most recent visit. One can also give uniform weights, so that each visit contributes equally to this “summary” visit, or higher weights to most recent visits if they are assumed to be more important than older visits.

As the prediction task is longitudinal, we also investigated the use of recurrent neural networks. Recurrent neural networks are a class of artificial neural networks dedicated to sequential data. We employed a simple architecture (Fig. 1) with a Gated Recurrent Unit [48] to extract information from the clinical measurements, followed by a concatenation of this vector with the socio-demographic and genetic data, followed by a Fully Connected layer with a sigmoid activation function. We used the PyTorch [49] Python package to build and train the recurrent neural network.

The recurrent neural network model is the only algorithm that learns to transform the variable-length sequence of inputs into a vector. All the machine learning models are trained end-to-end and there is no separate estimation of the $\mathbf{h}^{(t)}$ vector in the recurrent neural network model.

All the machine learning models are trained end-to-end and take as input clinical (age of PD onset, sex, past ICDs, anxiety, depression, REM sleep, types of PD medication and specific variables for the use of dopamine agonists, as described in Section II-B) and genetic data (as described in Section II-C), except in Section III-C in which we compare the models with the same machine learning algorithms trained with clinical data only.

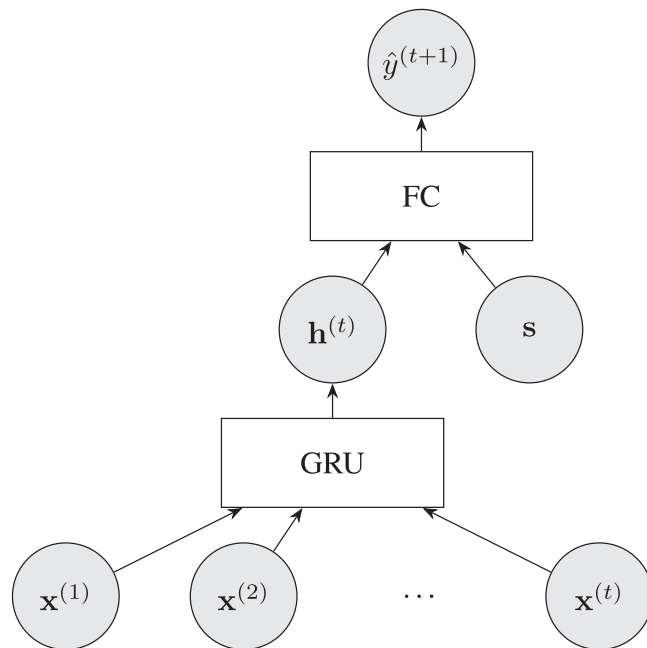


Fig. 1. Architecture of the recurrent neural network. The clinical features assessed at several visits, denoted as $(\mathbf{x}^{(1)}, \dots, \mathbf{x}^{(t)})$, are used as input of the Gated Recurrent Unit (GRU). The GRU extracts information from these clinical features into a vector $\mathbf{h}^{(t)}$. This vector and the time-independent variables, namely the socio-demographic and genetic data denoted as \mathbf{s} , are used as input of a Fully Connected (FC) layer followed by a sigmoid activation function, returning the probability of having an impulse control disorder at the next visit, denoted as $\hat{y}^{(t+1)}$.

Since the task is longitudinal, we defined a trivial model as the one that predicts the ICD status of a patient at the next visit with the ICD status at the most recent visit.

F. Cross-Validation

We used PPMI as the training (discovery) cohort, and DIGPD as the test (replication) cohort. To unbiasedly estimate the predictive performance of the models, we employed a nested cross-validation procedure that is illustrated in Fig. 2. In the outer loop, we randomly split 80% of the PPMI subjects into the training set and the remaining 20% into the test set. In the inner loop, we performed a 5-fold subject-level cross-validation procedure to optimize the hyper-parameters of the models on the training set. These hyper-parameters control how the algorithms fit the training data. For instance, these hyper-parameters included the type (ℓ_1 or ℓ_2 penalty) and amount (λ parameter) of regularization for the linear models. In particular, logistic regression models were regularized. The regularization applies to all the input data (clinical and genetic). After finding the optimal values for the hyper-parameters, each model was evaluated on the test set. Finally, we evaluated the performance of each model on the whole DIGPD cohort.

G. Statistical Analysis

Baseline characteristics in both cohorts were compared with chi-squared tests for categorical variables and t-tests for continuous variables using the SciPy [50] Python package. Predictive

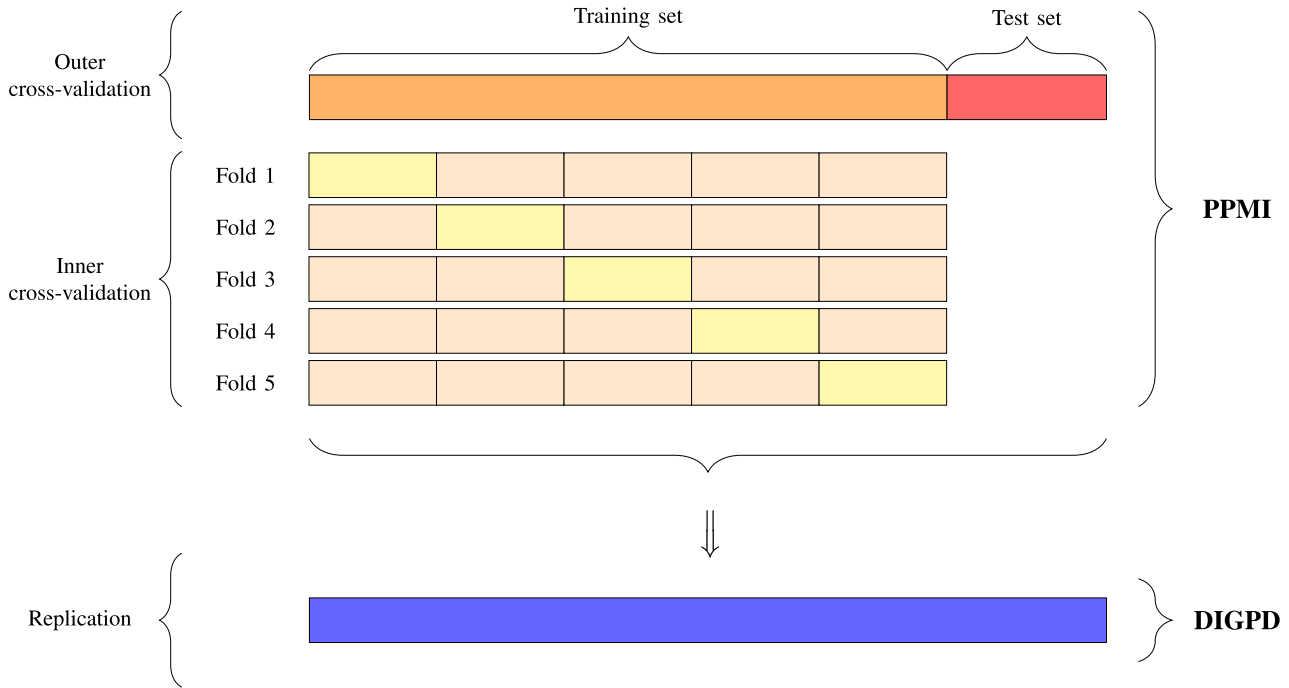


Fig. 2. Cross-validation procedure. We employed a nested cross-validation procedure on the PPMI cohort. In the outer loop, we split the PPMI subjects into training and test sets, while the inner loop was a 5-fold subject-level cross-validation to optimize the hyper-parameters of the model. The model with the optimal values for the hyper-parameters was evaluated on the test set of PPMI and on the whole DIGPD cohort.

performance was mainly evaluated using the area under the receiver operating characteristic curve and average precision (AP). AP summarizes a precision-recall curve as the weighted mean of precisions achieved at each threshold, with the increase in recall from the previous threshold used as the weight. The precision-recall curve is similar to the ROC curve, but plots the precision (positive predictive value) against the recall (sensitivity). The precision-recall curve does not take into account the true negatives, and is particularly useful when the positives are more important than the negatives (false negatives are more serious than false positives). Other metrics included accuracy, balanced accuracy (BA), sensitivity and specificity. ROC and precision-recall curves were plotted using the `matplotlib` [51] Python package, and all the metrics were computed using the `scikit-learn` [41] package. Comparison between ROC AUC was measured using the DeLong test [52]. P-values were adjusted for multiple comparisons using Bonferroni correction. We did not perform hypothesis testing to compare AP scores as we were not aware of a relevant statistical test to do so.

III. RESULTS

A. Population Characteristics

Out of the 423 PD subjects in PPMI, we excluded 1 subject for not having a baseline visit, 2 for not having medication records and 40 for not having genetic data. Out of the 415 PD subjects in DIGPD, we excluded 27 for having only a baseline visit. No subjects were excluded based on their genetic data. Thus, we included 380 PD subjects from PPMI and 388 PD subjects from DIGPD in our analyses. The 380 PPMI subjects had a total of

2,728 visits, while the 388 DIGPD subjects had a total of 2,101 visits. Since our objective was to predict the occurrence of ICDs at the next visit, the number of observations for a given subject is equal to their number of visits minus 1. Thus, the total number of observations was equal to 2,348 in PPMI and 1,713 in DIGPD.

Clinical characteristics are presented in Table I. Age and sex in both cohorts were not significantly different. PPMI subjects had significantly more visits and smaller intervals between back-to-back visits, as well as longer follow-ups. DIGPD subjects had significantly lower scores in the motor exam of the MDS-UPDRS. The prevalence of ICDs at baseline was significantly higher in DIGPD than in PPMI, as well as their lifetime prevalence. Both differences might be explained by the fact that PD subjects are *de novo* and drug-naive at baseline in PPMI whereas they are not in DIGPD. Other phenotypes (anxiety, depression, and REM sleep disorders) were not statistically compared due to the different scales used.

Concerning genetic data, we excluded 1 genetic variant for being a variable number of tandem repeat polymorphism. Furthermore, we excluded 18 SNPs for having too low imputation quality scores. Finally, 31 SNPs were included in our analyses (Supplementary Table 1).

B. Predictive Performance

Table II presents the predictive performance for the five main models: the trivial model, logistic regression models using the baseline visit, the most recent visit, and the mean over all the past visits respectively, and the recurrent neural network model. The logistic regression model using the baseline visit had some

TABLE I

BASELINE CHARACTERISTICS. FOR CONTINUOUS VARIABLES, MEAN \pm STANDARD DEVIATION IS REPORTED. FOR BINARY VARIABLES, THE COUNT FOR BOTH CATEGORIES IS REPORTED AS WELL AS THE PROPORTION OF THE FIRST CATEGORY. STATISTICAL DIFFERENCES WERE ASSESSED USING INDEPENDENT T TESTS FOR CONTINUOUS VARIABLES AND CHI-SQUARED TESTS FOR BINARY VARIABLES. GDS: GERIATRIC DEPRESSION SCALE; HAD: HOSPITAL ANXIETY AND DEPRESSION SCALE; RBDSQ: RAPID EYE MOVEMENT SLEEP BEHAVIOR DISORDER SCREENING QUESTIONNAIRE; STAI: STATE-TRAIT ANXIETY INVENTORY

Characteristic	PPMI	DIGPD	<i>p</i> -value
Age (in years)	60.67 \pm 9.71	58.99 \pm 9.75	1.71 $\times 10^{-2}$
Sex (F/M)	127/253 (33%)	155/233 (40%)	7.16 $\times 10^{-2}$
Length of follow-up (in years)	5.86 \pm 1.95	4.82 \pm 1.83	7.07 $\times 10^{-14}$
Number of visits per subject	7.18 \pm 2.96	5.41 \pm 1.66	4.21 $\times 10^{-35}$
Interval between visits (in years)	0.95 \pm 0.35	1.09 \pm 0.33	3.24 $\times 10^{-39}$
Anxiety	STAI: 93.55 \pm 7.96	HAD: 6.82 \pm 3.77	
Depression	GDS: 5.25 \pm 1.47	HAD: 4.59 \pm 3.16	
REM sleep	RBDSQ: 4.17 \pm 2.71	1/0: 86/302(22%)	
MDS-UPDRS III	20.87 \pm 8.86	9.91 \pm 5.33	3.77 $\times 10^{-72}$
Baseline ICD (1/0)	42/338 (13%)	76/312 (20%)	5.34 $\times 10^{-3}$
Lifetime ICD (1/0)	143/237 (38%)	192/196 (49%)	1.12 $\times 10^{-3}$

TABLE II

RESULTS OF THE FOUR MAIN MODELS. PREDICTIVE PERFORMANCE FOR THE FIVE MAIN MODELS ON BOTH COHORTS ARE REPORTED

		Trivial model	Logistic regression using only the baseline visit	Logistic regression using only the previous visit	Logistic regression using the mean over past visits	Recurrent neural network
ROC AUC	PPMI	0.752	0.753	0.795	0.838	0.850
	DIGPD	0.776	0.666	0.802	0.797	0.802
Average precision	PPMI	0.406	0.441	0.453	0.603	0.608
	DIGPD	0.555	0.429	0.644	0.615	0.623
Accuracy	PPMI	0.877	0.774	0.819	0.841	0.860
	DIGPD	0.833	0.566	0.592	0.645	0.785
Balanced accuracy	PPMI	0.752	0.691	0.764	0.765	0.759
	DIGPD	0.776	0.603	0.668	0.695	0.758
Sensitivity	PPMI	0.571	0.571	0.686	0.657	0.614
	DIGPD	0.651	0.684	0.838	0.805	0.699
Specificity	PPMI	0.932	0.810	0.843	0.873	0.904
	DIGPD	0.900	0.522	0.499	0.584	0.817

of the lowest scores on both cohorts (ROC AUC = 0.75 and AP = 0.44 in PPMI, ROC AUC = 0.67 and AP = 0.43 in DIGPD). The trivial model reached a ROC AUC = 0.75 of in PPMI and of 0.78 in DIGPD. The recurrent neural network yielded the highest scores in PPMI (ROC AUC = 0.85, AP = 0.61), while the logistic regression using the most recent visit yielded the highest scores in DIGPD (ROC AUC = 0.80, AP = 0.64). Fig. 3 and Fig. 4 show the ROC and precision-recall curves for the four main models compared to the trivial model in PPMI and in DIGPD respectively. The recurrent neural network models had sensitivities of 61% and 70% and specificities of 90% and 82% in PPMI and DIGPD respectively, at the default threshold (probability > 0.5).

The recurrent neural network model was the only model to be significantly better than the trivial model on both cohorts, and two logistic regression models were significantly better than the trivial model on one cohort (Fig. 5).

Although AP scores for the three best models were higher in DIGPD than in PPMI, the prevalence of ICDs, computed over all the (patient, visit) pairs, was twice higher in DIGPD than in PPMI (27% in DIGPD, 14% in PPMI). As AP scores of random guess are equal to the prevalence of the positive class,

the differences between AP scores in both cohorts should be interpreted with much caution.

The other machine learning algorithms (support vector machines with linear and radial basis function kernels, random forest, and gradient tree boosting) and other reduction approaches (giving positive weights to all the past visits, but higher weights to more recent visits) yielded comparable results (Supplementary Table 2).

To evaluate the impact of the splitting of PPMI into training and test sets on the predictive performance, we repeated the cross-validation procedure 10 times and also evaluated the 10 models on DIGPD. All iterations yielded comparable results (Supplementary Table 3).

To evaluate the impact of the choice of the training and replication cohorts, we used DIGPD as the training cohort and PPMI as the replication cohort and obtained similar results (Supplementary Tables 4 and 5).

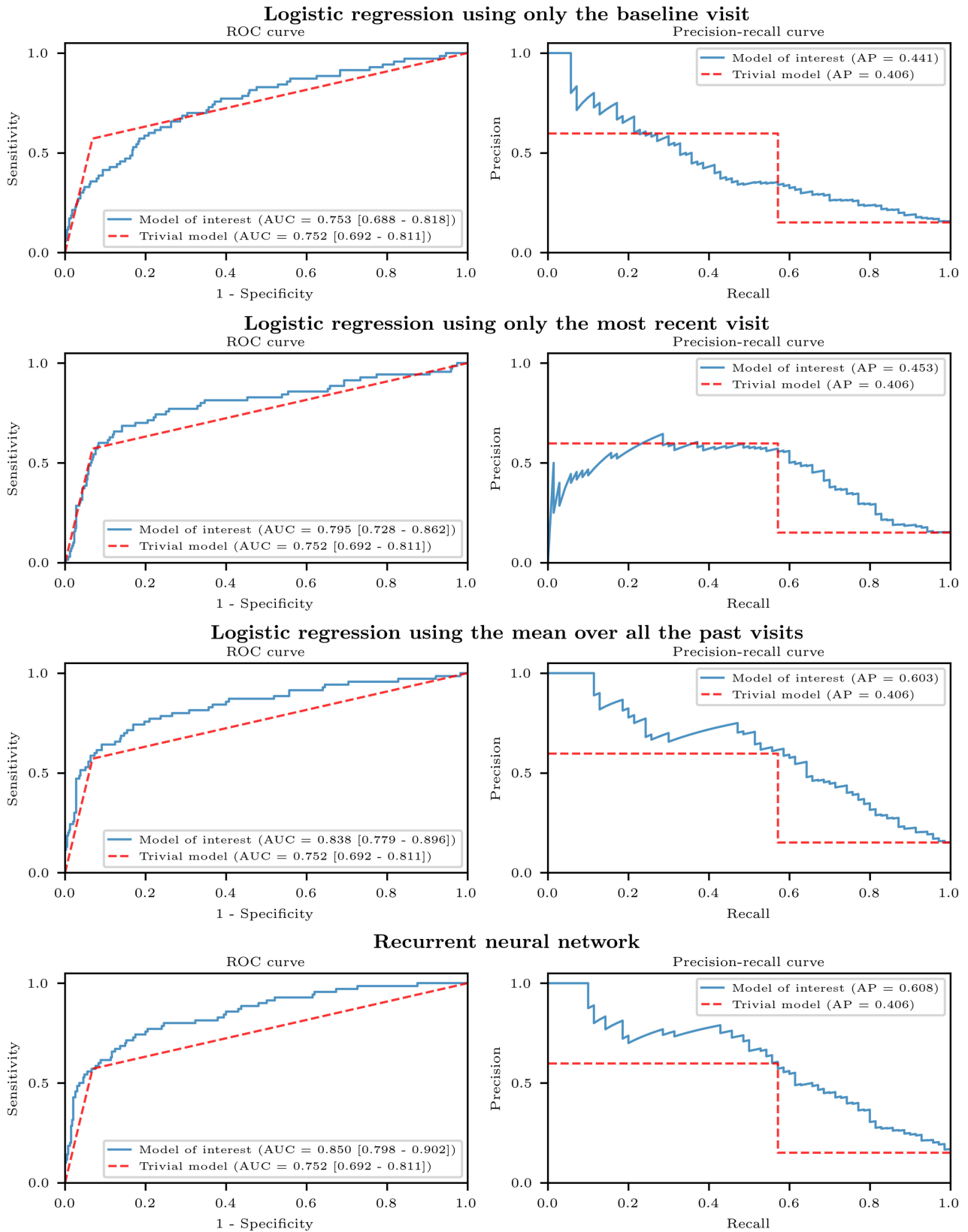


Fig. 3. ROC and Precision-recall curves on PPMI. The models, trained on a subset of subjects in PPMI, were evaluated on an independent subset of subjects in PPMI.

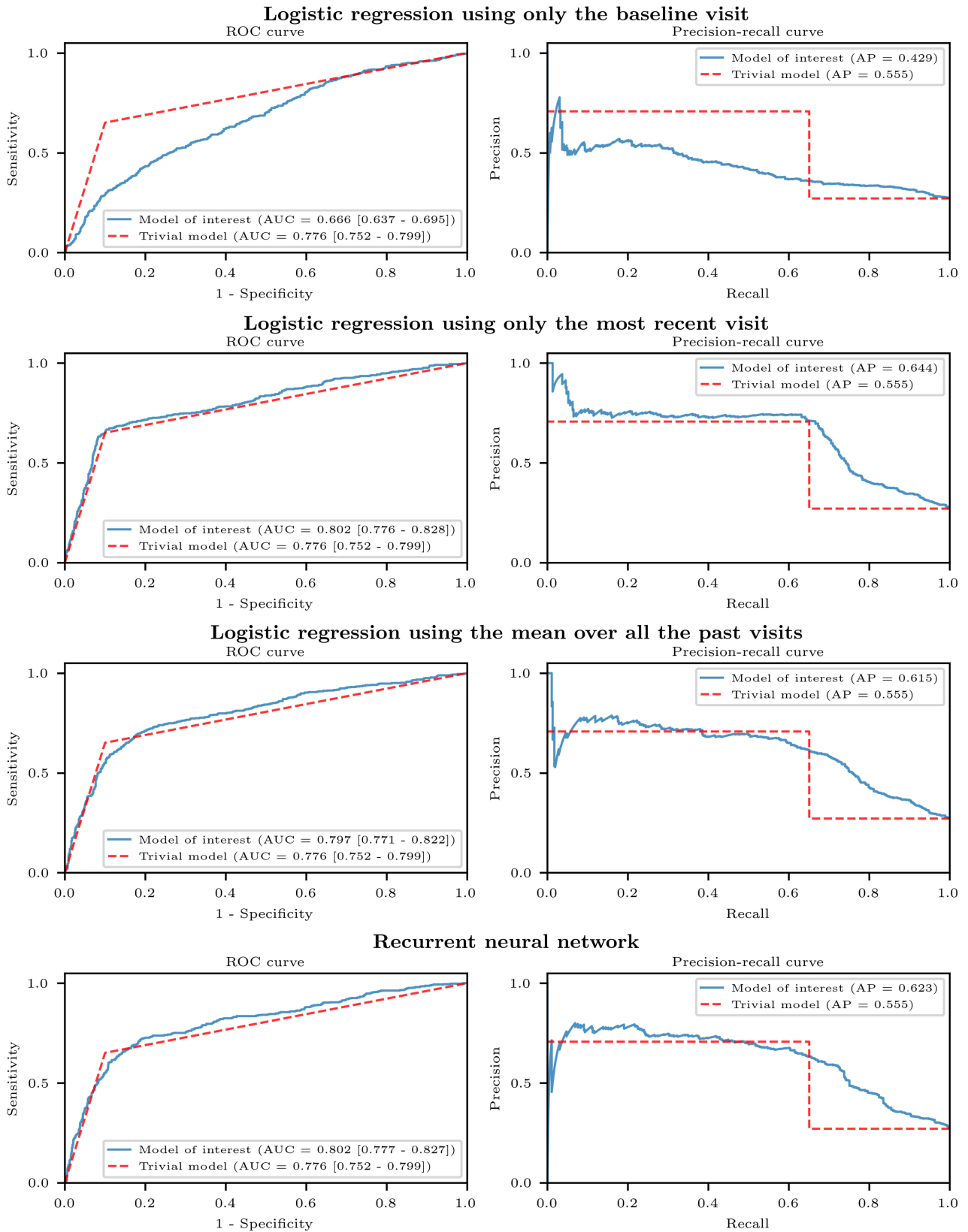


Fig. 4. ROC and Precision-recall curves on DIGPD. The models, trained on a subset of subjects in PPMI, were applied on all the subjects in DIGPD.

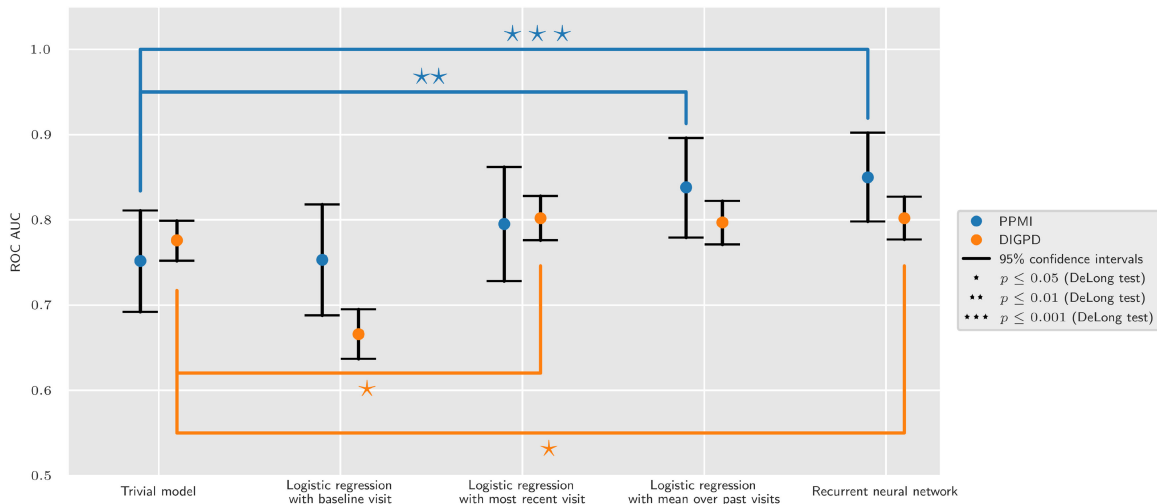


Fig. 5. Statistical comparison of ROC AUC for the five main models. 95% confidence intervals were computed using the asymptotic normal distributions of the ROC AUC scores. P-values were computed using the DeLong test. P-values below the 0.05 threshold after adjustment for multiple comparison using Bonferroni correction are highlighted with at least one asterisk: $p \leq 0.05$ (*), $p \leq 0.01$ (**), $p \leq 0.001$ (***) .

TABLE III

STATISTICAL COMPARISON OF ROC AUC FOR THE FOUR MAIN MODELS WITH AND WITHOUT GENETIC VARIANTS. DIFFERENCES IN ROC AUC BETWEEN THE MODELS WITH AND WITHOUT GENETIC VARIANTS WERE ASSESSED WITH THE DELONG TEST. SIGNIFICANT DIFFERENCES AFTER BONFERRONI CORRECTION ARE HIGHLIGHTED IN BOLD FONT

		Logistic regression using only the baseline visit	Logistic regression using only the previous visit	Logistic regression using the mean over past visits	Recurrent neural network
PPMI	With genetic variants	0.753 [0.688 – 0.818]	0.795 [0.728 – 0.862]	0.838 [0.779 – 0.896]	0.850 [0.798 – 0.902]
	Without genetic variants	0.751 [0.687 – 0.816]	0.807 [0.745 – 0.868]	0.843 [0.789 – 0.899]	0.845 [0.792 – 0.899]
	<i>p</i> -value	0.885	0.0971	0.637	0.451
DIGPD	With genetic variants	0.666 [0.637 – 0.695]	0.802 [0.776 – 0.828]	0.797 [0.771 – 0.822]	0.802 [0.777 – 0.827]
	Without genetic variants	0.682 [0.654 – 0.711]	0.786 [0.758 – 0.813]	0.788 [0.762 – 0.813]	0.803 [0.778 – 0.828]
	<i>p</i> -value	0.0143	0.000252	0.109	0.757

C. Contribution of the Different Features

Since the genetic factors of ICDs in PD are mostly unknown and genotyping data is not usually collected in clinical routine, we investigated the predictive performance of the same algorithms without the genetic variants as input, in order to assess their added value in the models. Table III presents the ROC AUC of the models with and without genetic variants and their statistical comparison. Only one comparison was statistically different: the logistic regression model using the most recent visit had a higher ROC AUC with genetic variants than without genetic variants on DIGPD (ROC AUC = 0.80 with genetic variants, ROC AUC = 0.79 without genetic variants, $p < 0.001$). The genetic variants did not seem to be major contributors to the decision function of the logistic regression models.

We also investigated the coefficients of the three logistic regression models with and without genetic variants as input (Supplementary Tables 6 and 7). As the logistic regression model using the baseline visit did not perform better than the trivial model, and the variables for PD medication were all null (PD patients in PPMI are de novo drug-naïve at baseline, and the medical history of PD patients in DIGPD was not available before their

baseline visit), we only interpreted the other two models. The following features had positive coefficients: sex, past ICDs, depression, REM sleep, motor exam, being on other PD medication than levodopa and dopamine agonists, and maximum dose and cumulative duration of dopamine agonists. On the other hand, the following features had negative coefficients: age, anxiety, being on levodopa, and mean daily and total dose of dopamine agonists. The features corresponding to being on dopamine agonists and time to prediction had coefficients close to zero. The variables with the largest absolute values were past ICDs, and total dose and cumulative duration of dopamine agonists.

IV. DISCUSSION

To the best of our knowledge, this study is the first one evaluating the predictability of ICDs in PD in an unbiased manner using two longitudinal cohorts, including one independent replication cohort.

Three previous studies reported ROC AUC for a prediction task of ICDs in PD [24], [29], [30]. Kraemmer and colleagues reported ROC AUC of 0.65 [0.58 – 0.73] with clinical variables only and of 0.76 [0.70 – 0.83] with clinical and genetic variables,

while Erga and colleagues reported ROC AUC of 0.68 [0.59 – 0.78] with clinical features only and of 0.70 [0.61 – 0.79] with clinical and genetic features, and Jesus and colleagues reported ROC AUC of 0.69 with clinical features only and of 0.80 with clinical and genetic features. However, the methods and prediction tasks were different. Erga and colleagues performed a cross-sectional analysis of 119 PD patients from the Norwegian ParkWest study, Jesus and colleagues also performed a cross-sectional analysis of 353 PD patients recruited from the Movement Disorder Clinic of the University Hospital Virgen del Rocío in Seville, Spain, while Kraemmer and colleagues performed a longitudinal analysis of 276 PD patients from PPMI. In their studies, each patient corresponds to a unique observation, leading to much lower sample sizes. Moreover, all three studies did not use an independent test set nor cross-validation and did not have a replication cohort, which might lead to overly optimistic reported results [31]. By contrast, our study uses cross-validation on the training cohort (PPMI), an independent test set from PPMI and a replication cohort (DIGPD). Results were overall comparable in both cohorts, even though the performance was slightly lower in DIGPD than in PPMI, and the difference with the trivial model was also less substantial, although statistically significant. This may be explained by the fact that both cohorts have different characteristics (de novo drug-naive patients in PPMI, already-treated patients in DIGPD) and some variables (anxiety, depression, REM sleep) were not measured with the same instruments.

The best performing model was the recurrent neural network. It achieved a statistically higher performance than the trivial model in both cohorts. Nevertheless, while the difference in performance was substantial (10 percentage points of ROC AUC) for the PPMI dataset, it was much weaker for DIGPD dataset. This may be partly due to the difficulty of generalizing to a different cohort with different inclusion criteria and different measurement scales. Nevertheless, the results obtained when using DIGPD for training and PPMI for replication (see Supplementary Tables 4 and 5) suggest that prediction may be inherently slightly more difficult in the DIGPD cohort.

The logistic regression coefficients were overall consistent with the literature. For the socio-demographic variables, sex and age have respectively a positive and negative coefficients, in accordance with a younger age and a male sex previously associated with ICDs in PD [15]. Depression, REM sleep and motor exam scores also had positive coefficients, consistent with their positive association [14]. Anxiety scores had negative coefficients although previously reported to be positively associated with ICD. The maximum dose and cumulative duration of dopamine agonists had positive coefficients, confirming the important role of the dose and the duration of dopamine agonist therapy in the risk to develop ICDs in PD [5]. Interestingly, the types of PD medication sparsely contributed to the decision function of the models, with very small coefficients. The mean daily and total doses of dopamine agonists had negative coefficients, although these coefficients were almost null for the mean daily dose. These derived features have rarely been investigated altogether, making the comparison with the literature difficult. It should be noted that the coefficients are estimated altogether

and that the logistic regression models were regularized, so interpretation should be performed with caution.

We used features that have been associated with ICDs in PD as input of our models, but there are probably more unknown risk factors to be discovered. In addition, as a class of psychiatric disorders, ICDs are particularly complex, with qualitative environmental factors that might play important roles, are difficult to measure, and are not captured by clinical scales used in PD. Assessment of ICDs may also be noisy (e.g. patients hiding or not aware of their behavior), and thus ICDs are probably less predictable in practice than other comorbidities in PD, such as dementia [53]. Finally, little is known about the genetic factors of ICDs in PD. In our study, the inclusion of genetic variants did not lead to a substantial improvement over clinical data alone for the best performing models. In absence of genome-wide association study and genetic risk scores for ICDs in PD, we used associated genetic variants from candidate gene analyses [23], [24], [29]. As variation in complex traits is caused by numerous genetic variants, such analyses have important limitations and many association studies could not be replicated, particularly in psychiatric conditions like schizophrenia [54]. More studies, in particular genome-wide association studies, are needed to better understand the genetic landscape of ICDs in PD.

Being able to predict ICDs is of critical importance due to their potential medical, financial, and/or legal medical complications. Identifying patients at high risk to develop ICDs at the next visit may lead to changes in the dopaminergic treatment strategy (e.g. decrease the dose of dopamine agonists and increase levodopa) and/or recommend a closer monitoring of behavioral changes by the caregiver. The efficacy of such preventive strategies based on a predictive model remains however to be evaluated. In this perspective, the model may be adapted depending on the relative importance for identifying positives (patients who will develop ICDs) or negatives (patients who will not develop ICDs). The balanced accuracy scores were equal to 76% for the recurrent neural network models in both cohorts, but the sensitivities (61% vs 70%) and specificities (90% vs 82%) differed, which might be explained by the different prevalences in both cohorts. Using the default threshold (probability > 0.5) made the models more specific than sensitive, which might be a limitation if finding the positives is more important than the negatives. On the other hand, models being more specific than sensitive might be more relevant if the main objective is to propose treatment changes only to patients who are at strong risk, and avoid unnecessary modifications in more patients. The threshold can still be adjusted depending on the main objective. Prospective studies are required to validate the models and allow their relevance in clinical routine. Another clinically relevant task would be to predict the first onset of ICDs in PD, but is left for future work.

Our study has several limitations. A first important limitation is that the performances of the best models were only marginally better than that of a trivial model on the replication cohort DIGPD. While the difference was statistically significant, its magnitude is too small for the tool to be useful in clinical practice in its current state. They nevertheless set a methodologically solid basis for the development of improved models. Second, as mentioned in Section II-B, the definition of ICDs differ between

cohorts. This may partly explain why our machine learning models struggle to generalize in the independent replication cohort. Third, the sample sizes are relatively small, in particular on the test set of PPMI due to the use of cross-validation. Fourth, each observation is a (subject, visit) pair and thus the observations are not independent (the intra-subject observations are not independent, but the inter-subject observations are independent), which could lead to underestimating p-values when assessing the statistical difference between ROC AUC. Fifth, in absence of genome-wide association study and genetic risk scores for ICDs in PD, we used associated genetic variants from candidate gene analyses. Genetic risk scores are more robust estimators of the genetic liability of a phenotype and should be preferred when available [55]. We previously investigated the genetics of ICDs in PD by computing genetic risk scores for other phenotypes and investigating their statistical association with ICDs, but we could not report any statistical association [56]. Sixth, we did not include several risk factors of ICDs in PD as input of the algorithms, such as dyskinesia [57] and sleep disorders other than REM sleep. Nonetheless, the added value of known risk factors that were not included remains to be proven, as these risk factors were discovered in univariate analyses. Moreover, adding more features as input of machine learning algorithms does not necessarily lead to better predictive performance due to the correlation between the features and the risk of overfitting.

V. CONCLUSION

Our study shows the feasibility of prediction of impulse control disorders in Parkinson's disease. Nevertheless, the improvements obtained compared to a trivial model are not sufficient to support clinical utility at this stage of research. Nonetheless, our study highlights a sound methodology and sets a baseline that future studies can compare to. Further studies including other risk factors and investigating the first onset of ICDs are required to obtain clinically relevant models.

ACKNOWLEDGMENT

DIGPD Study Group

Steering committee: Jean-Christophe Corvol, MD, PhD (Pitié-Salpêtrière Hospital, Paris, principal investigator of DIGPD), Alexis Elbaz, MD, PhD (CESP, Villejuif, member of the steering committee), Marie Vidailhet, MD (Pitié-Salpêtrière Hospital, Paris, member of the steering committee), Alexis Brice, MD (Pitié-Salpêtrière Hospital, Paris, member of the steering committee and PI for genetic analysis).

Statistical analyses: Alexis Elbaz, MD, PhD (CESP, Villejuif, PI for statistical analyses), Fanny Artaud, PhD (CESP, Villejuif, statistician).

Principal investigators for sites (alphabetical order): Frédéric Bourdain, MD (CH Foch, Suresnes, PI for site), Jean-Philippe Brandel, MD (Fondation Rothschild, Paris, PI for site), Jean-Christophe Corvol, MD, PhD (Pitié-Salpêtrière Hospital, Paris, PI for site), Pascal Derkinderen, MD, PhD (CHU Nantes, PI for site), Franck Durif, MD (CHU Clermont-Ferrand, PI for

site), Richard Levy, MD, PhD (CHU Saint-Antoine, Paris, PI for site), Fernando Pico, MD (CH Versailles, PI for site), Olivier Rascol, MD (CHU Toulouse, PI for site).

Co-investigators (alphabetical order): Anne-Marie Bonnet, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Cecilia Bonnet, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Christine Brefel-Courbon, MD (CHU Toulouse, site investigator), Florence Cormier-Dequaire, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Bertrand Degos, MD, PhD (Pitié-Salpêtrière Hospital, site investigator), Bérangère Debilly, MD (CHU Clermont-Ferrand, site investigator), Alexis Elbaz, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Monique Galitsky (CHU de Toulouse, site investigator), David Grabli, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Andreas Hartmann, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Stephan Klebe, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Julia Kraemmer, MD (Pitié-Salpêtrière Hospital, site investigator), Lucette Lacomblez, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Sara Leder, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Graziella Mangone, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Louise-Laure Mariani, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Ana-Raquel Marques, MD (CHU Clermont Ferrand, site investigator), Valérie Mesnage, MD (CHU Saint Antoine, Paris, site investigator), Julia Muellner, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Fabienne Ory-Magne, MD (CHU Toulouse, site investigator), Violaine Planté-Bordeneuve, MD (Henri Mondor Hospital, Créteil, site investigator), Emmanuel Roze, MD, PhD (Pitié-Salpêtrière Hospital, Paris, site investigator), Melissa Tir, MD (CH Versailles, site investigator), Marie Vidailhet, MD (Pitié-Salpêtrière Hospital, Paris, site investigator), Hana You, MD (Pitié-Salpêtrière Hospital, Paris, site investigator);

Neuropsychologists: Eve Benchetrit, MS (Pitié-Salpêtrière Hospital, Paris, neuropsychologist), Julie Socha, MS (Pitié-Salpêtrière Hospital, Paris, neuropsychologist), Fanny Pineau, MS (Pitié-Salpêtrière Hospital, Paris, neuropsychologist), Tiphaine Vidal, MS (CHU Clermont-Ferrand, neuropsychologist), Elsa Pomies (CHU de Toulouse, neuropsychologist), Virginie Bayet (CHU de Toulouse, neuropsychologist); Genetic core: Alexis Brice (Pitié-Salpêtrière Hospital, Paris, PI for genetic studies), Suzanne Lesage, PhD (INSERM, ICM, Paris, genetic analyses), Khadija Tahiri, PhD (Inserm, ICM, Paris, lab technician) Hélène Bertrand, MS (Inserm, ICM, Paris, lab technician), Graziella Mangone, MD, PhD (Pitié-Salpêtrière Hospital, Paris, genetic analyses); Sponsor activities and clinical research assistants: Alain Mallet, PhD (Pitié-Salpêtrière Hospital, Paris, sponsor representative), Coralie Villeret (Hôpital Saint Louis, Paris, Project manager), Merry Mazmanian (Pitié-Salpêtrière Hospital, Paris, project manager), Hakima Manseur (Pitié-Salpêtrière Hospital, Paris, clinical research assistant), Mostafa Hajji (Pitié-Salpêtrière Hospital, Paris, data manager), Benjamin Le Toullec, MS (Pitié-Salpêtrière Hospital, Paris, clinical research assistant), Vanessa Brochard, PhD (Pitié-Salpêtrière Hospital, Paris, project manager), Monica Roy, MS (CHU de Nantes, clinical research assistant), Isabelle Rieu, PhD

(CHU Clermont-Ferrand, clinical research assistant), Stéphane Bernard (CHU Clermont-Ferrand, clinical research assistant), Antoine Faurie-Grepon (CHU de Toulouse, clinical research assistant).

SUPPLEMENTARY MATERIALS

We list the different reduction approaches that we investigated and the genetic variants included in our analyses. We also provide additional experiments with other machine learning algorithms and other reduction approaches, with several cross-validation runs, and the same experiments but with swapping the role of both cohorts, and the coefficients of the best logistic regression models.

REFERENCES

- [1] J. P. Hiseman and R. Fackrell, "Caregiver burden and the nonmotor symptoms of Parkinson's disease," *Int. Rev. Neurobiol.*, vol. 133, pp. 479–497, 2017.
- [2] A. H. Evans, A. P. Strafella, D. Weintraub, and M. Stacy, "Impulsive and compulsive behaviors in Parkinson's disease," *Movement Disord.: Official J. Movement Disord. Soc.*, vol. 24, no. 11, pp. 1561–1570, 2009.
- [3] V. Voon *et al.*, "Prevalence of repetitive and reward-seeking behaviors in Parkinson disease," *Neurology*, vol. 67, no. 7, pp. 1254–1257, 2006.
- [4] D. Weintraub *et al.*, "Impulse control disorders in Parkinson disease: A cross-sectional study of 3090 patients," *Arch. Neurol.*, vol. 67, no. 5, pp. 589–595, May 2010.
- [5] J.-C. Corvol *et al.*, "Longitudinal analysis of impulse control disorders in Parkinson disease," *Neurology*, vol. 91, no. 3, pp. e189–e201, Jul. 2018.
- [6] K. M. Smith, S. X. Xie, and D. Weintraub, "Incident impulse control disorder symptoms and dopamine transporter imaging in Parkinson disease," *J. Neurol. Neurosurg. Psychiatry*, vol. 87, no. 8, pp. 864–870, 2016.
- [7] A. H. Erga, G. Alves, O. B. Tysnes, and K. F. Pedersen, "Evolution of impulsive-compulsive behaviors and cognition in Parkinson's disease," *J. Neurol.*, vol. 267, no. 1, pp. 259–266, 2020.
- [8] P. Parra-Díaz *et al.*, "Does the country make a difference in impulse control disorders? A systematic review," *Movement Disord. Clin. Pract.*, vol. 8, no. 1, pp. 25–32, 2021.
- [9] A. L. Phu *et al.*, "Effect of impulse control disorders on disability and quality of life in Parkinson's disease patients," *J. Clin. Neurosci.*, vol. 21, no. 1, pp. 63–66, 2014.
- [10] I. Leroi, V. Harbisetar, M. Andrews, K. McDonald, E. J. Byrne, and A. Burns, "Carer burden in apathy and impulse control disorders in Parkinson's disease," *Int. J. Geriatr. Psychiatry*, vol. 27, no. 2, pp. 160–166, 2012.
- [11] A. H. Erga, G. Alves, O. B. Tysnes, and K. F. Pedersen, "Impulsive and compulsive behaviors in Parkinson's disease: Impact on quality of life and satisfaction with life, and caregiver burden," *Parkinsonism Related Disord.*, vol. 78, pp. 27–30, 2020.
- [12] E. Mamikonyan *et al.*, "Long-term follow-up of impulse control disorders in Parkinson's disease," *Movement Disord.*, vol. 23, no. 1, pp. 75–80, 2008.
- [13] M. J. Nirenberg and C. Waters, "Compulsive eating and weight gain related to dopamine agonist use," *Movement Disord.*, vol. 21, no. 4, pp. 524–529, 2006.
- [14] M. Grall-Bronnec *et al.*, "Dopamine agonists and impulse control disorders: A complex association," *Drug Saf.*, vol. 41, no. 1, pp. 19–75, Jan. 2018.
- [15] D. Weintraub and D. O. Claassen, "Impulse control and related disorders in Parkinson's disease," *Int. Rev. Neurobiol.*, vol. 133, pp. 679–717, 2017.
- [16] M. B. Callesen, D. Weintraub, M. F. Damholdt, and A. Møller, "Impulsive and compulsive behaviors among Danish patients with Parkinson's disease: Prevalence, depression, and personality," *Parkinsonism Related Disord.*, vol. 20, no. 1, pp. 22–26, Jan. 2014.
- [17] M. Poletti *et al.*, "A single-center, cross-sectional prevalence study of impulse control disorders in Parkinson disease: Association with dopaminergic drugs," *J. Clin. Psychopharmacol.*, vol. 33, no. 5, pp. 691–694, Oct. 2013.
- [18] F. E. Pontieri *et al.*, "Sociodemographic, neuropsychiatric and cognitive characteristics of pathological gambling and impulse control disorders NOS in Parkinson's disease," *Eur. Neuropsychopharmacology: J. Eur. College Neuropsychopharmacol.*, vol. 25, no. 1, pp. 69–76, Jan. 2015.
- [19] I. Leroi *et al.*, "Apathy and impulse control disorders in Parkinson's disease: A direct comparison," *Parkinsonism Related Disord.*, vol. 18, no. 2, pp. 198–203, Feb. 2012.
- [20] V. Voon *et al.*, "Impulse control disorders in Parkinson disease: A multicenter case-control study," *Ann. Neurol.*, vol. 69, no. 6, pp. 986–996, Jun. 2011.
- [21] M. L. Fantini *et al.*, "Increased risk of impulse control symptoms in Parkinson's disease with REM sleep behaviour disorder," *J. Neurol. Neurosurg., Psychiatry*, vol. 86, no. 2, pp. 174–179, Feb. 2015.
- [22] C. C. Ramírez Gómez *et al.*, "A multicenter comparative study of impulse control disorder in Latin American patients with Parkinson disease," *Clin. Neuropharmacol.*, vol. 40, no. 2, pp. 51–55, Apr. 2017.
- [23] F. Cormier-Dequaire *et al.*, "Suggestive association between OPRM1 and impulse control disorders in Parkinson's disease," *Movement Disord.: Official J. Movement Disord. Soc.*, vol. 33, no. 12, pp. 1878–1886, 2018.
- [24] A. H. Erga *et al.*, "Dopaminergic and opioid pathways associated with impulse control disorders in Parkinson's disease," *Front. Neurol.*, vol. 9, 2018, Art. no. 109.
- [25] X. H. Castro-Martínez *et al.*, "Behavioral addictions in early-onset Parkinson disease are associated with DRD3 variants," *Parkinsonism Related Disord.*, vol. 49, pp. 100–103, 2018.
- [26] S. Zainal Abidin *et al.*, "DRD and GRIN2B polymorphisms and their association with the development of impulse control behaviour among Malaysian Parkinson's disease patients," *BMC Neurol.*, vol. 15, p. 59, Apr. 2015.
- [27] S. Krishnamoorthy *et al.*, "Dopamine D3 receptor Ser9Gly variant is associated with impulse control disorders in Parkinson's disease patients," *Parkinsonism Related Disord.*, vol. 30, pp. 13–17, 2016.
- [28] J.-Y. Lee *et al.*, "Association of DRD3 and GRIN2B with impulse control and related behaviors in Parkinson's disease," *Movement Disord.: Official J. Movement Disord. Soc.*, vol. 24, no. 12, pp. 1803–1810, Sep. 2009.
- [29] J. Kraemmer *et al.*, "Clinical-genetic model predicts incident impulse control disorders in Parkinson's disease," *J. Neurol. Neurosurg., Psychiatry*, vol. 87, no. 10, pp. 1106–1111, Oct. 2016.
- [30] S. Jesús *et al.*, "Integrating genetic and clinical data to predict impulse control disorders in Parkinson's disease," *Eur. J. Neurol.*, vol. 28, pp. 459–468, Feb. 2021.
- [31] A. Koul, C. Becchio, and A. Cavallo, "Cross-validation approaches for replicability in psychology," *Front. Psychol.*, vol. 9, 2018, Art. no. 1117.
- [32] K. Marek *et al.*, "The Parkinson progression marker initiative (PPMI)," *Prog. Neurobiol.*, vol. 95, no. 4, pp. 629–635, Dec. 2011.
- [33] D. Weintraub *et al.*, "Questionnaire for impulsive-compulsive disorders in Parkinson's disease-rating scale," *Movement Disord.: Official J. Movement Disord. Soc.*, vol. 27, no. 2, pp. 242–247, Feb. 2012.
- [34] F. Charbonnier-Beaupel *et al.*, "Gene expression analyses identify Narp contribution in the development of l-DOPA-induced dyskinesia," *J. Neurosci.*, vol. 35, no. 1, pp. 96–111, Jan. 2015.
- [35] M. A. Nalls *et al.*, "NeuroX, a fast and efficient genotyping platform for investigation of neurodegenerative diseases," *Neurobiol. Aging*, vol. 36, no. 3, pp. 1605.e7–1605.12, Mar. 2015.
- [36] S. Das *et al.*, "Next-generation genotype imputation service and methods," *Nature Genet.*, vol. 48, no. 10, pp. 1284–1287, 2016.
- [37] S. McCarthy *et al.*, "A reference panel of 64,976 haplotypes for genotype imputation," *Nature Genet.*, vol. 48, no. 10, pp. 1279–1283, 2016.
- [38] C. C. Chang, C. C. Chow, L. C. Tellier, S. Vattikuti, S. M. Purcell, and J. J. Lee, "Second-generation PLINK: Rising to the challenge of larger and richer datasets," *GigaScience*, vol. 4, no. 1, Feb. 2015, Art. no. s13742-015-0047-8.
- [39] W. McKinney, "Data structures for statistical computing in Python," in *Proc. 9th Python Sci. Conf.*, 2010, pp. 56–61.
- [40] C. R. Harris *et al.*, "Array programming with NumPy," *Nature*, vol. 585, no. 7825, pp. 357–362, Sep. 2020.
- [41] F. Pedregosa *et al.*, "Scikit-learn: Machine learning in Python," *J. Mach. Learn. Res.*, vol. 12, no. 85, pp. 2825–2830, 2011.
- [42] T. Chen and C. Guestrin, "XGBoost: A scalable tree boosting system," in *Proc. 22nd ACM SIGKDD Int. Conf. Knowl. Discov. Data Mining*, 2016, pp. 785–794.
- [43] B. E. Boser, I. M. Guyon, and V. N. Vapnik, "A training algorithm for optimal margin classifiers," in *Proc. 5th Annu. Workshop Comput. Learn. Theory*, 1992, pp. 144–152.
- [44] C. Cortes and V. Vapnik, "Support-vector networks," *Mach. Learn.*, vol. 20, no. 3, pp. 273–297, Sep. 1995.
- [45] L. Breiman, "Random Forests," *Mach. Learn.*, vol. 45, no. 1, pp. 5–32, Oct. 2001.

- [46] L. Mason, J. Baxter, P. L. Bartlett, and M. R. Frean, "Boosting algorithms as gradient descent," in *Advances in Neural Information Processing Systems 12*, S. A. Solla, T. K. Leen, and K. Müller, Eds. Cambridge, MA, USA: MIT Press, 2000.
- [47] J. H. Friedman, "Greedy function approximation: A gradient boosting machine," *Ann. Statist.*, vol. 29, no. 5, pp. 1189–1232, 2001.
- [48] K. Cho *et al.*, "Learning phrase representations using RNN encoder-decoder for statistical machine translation," in *Proc. Conf. Empirical Methods Natural Lang. Process.*, 2014, pp. 1724–1734.
- [49] A. Paszke *et al.*, "PyTorch: An imperative style, high-performance deep learning library," in *Advances in Neural Information Processing Systems 32*, H. Wallach, H. Larochelle, A. Beygelzimer, F. D. Alché-Buc, E. Fox, and R. Garnett, Eds. Red Hook, NY, USA: Curran Associates, Inc., 2019, pp. 8026–8037.
- [50] P. Virtanen *et al.*, "SciPy 1.0: Fundamental algorithms for scientific computing in Python," *Nature Methods*, vol. 17, no. 3, pp. 261–272, Mar. 2020.
- [51] J. D. Hunter, "Matplotlib: A 2D graphics environment," *Comput. Sci. Eng.*, vol. 9, no. 3, pp. 90–95, May 2007.
- [52] E. R. DeLong, D. M. DeLong, and D. L. Clarke-Pearson, "Comparing the areas under two or more correlated receiver operating characteristic curves: A nonparametric approach," *Biometrics*, vol. 44, no. 3, pp. 837–845, Sep. 1988.
- [53] G. Liu *et al.*, "Prediction of cognition in Parkinson's disease with a clinical-genetic score: A longitudinal analysis of nine cohorts," *Lancet Neurol.*, vol. 16, no. 8, pp. 620–629, 2017.
- [54] E. C. Johnson *et al.*, "No evidence that schizophrenia candidate genes are more associated with schizophrenia than non-candidate genes," *Biol. Psychiatry*, vol. 82, no. 10, pp. 702–708, Nov. 2017.
- [55] N. R. Wray, M. E. Goddard, and P. M. Visscher, "Prediction of individual genetic risk to disease from genome-wide association studies," *Genome Res.*, vol. 17, no. 10, pp. 1520–1528, Oct. 2007.
- [56] J. Faouzi, B. Couvy-Duchesne, S. Bekadar, O. Colliot, and J.-C. Corvol, "Exploratory analysis of the genetics of impulse control disorders in Parkinson's disease using genetic risk scores," *Parkinsonism Related Disord.*, vol. 86, pp. 74–77, 2021.
- [57] R. Biundo *et al.*, "Impulse control disorders in advanced Parkinson's disease with dyskinesia: The ALTHEA study," *Movement Disord.: Official J. Movement Disord. Soc.*, vol. 32, no. 11, pp. 1557–1565, Nov. 2017.