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Duration of Zika Viremia in Serum

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breakthrough IFI among high-risk hematologic malignancy and organ transplant patients on isavuconazole for prophylaxis and treatment (Table 1). All patients presented with pneumonia, including 2 cases due to *Aspergillus nigri* group, a nonfumigatus *Aspergillus* species that exhibits increased minimum inhibitory concentrations to isavuconazole [10]. Additional breakthrough IFI pathogens included *Rhizopus* spp., *Scedosporium apiospermum*, and *Aspergillus fumigatus*. All patients received oral isavuconazole at standard dosing. In 2 cases, isavuconazole levels were measured and demonstrated adequate absorption. All cases were treated with escalation to liposomal amphotericin B with or without a second antifungal agent (triazole or echinocandin). The risk of mortality with breakthrough IFI was high: 3 of 5 died.

Although isavuconazole is an important therapeutic option in the management of IFIs, the epidemiology and microbiology of breakthrough infections on this agent are not well understood. To help guide optimal empirical management in these difficult cases, additional studies are needed to systematically evaluate breakthrough IFIs on isavuconazole given concern for azole-resistant or emerging fungal pathogens.

Note

Potential conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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References

- Mellinghoff SC, Panse J, Alakel N, et al. Primary prophylaxis of invasive fungal infections in patients with hematological malignancies: 2017 update of the recommendations of the Infectious Diseases Working Party (AGIHO) of the German Society for Hematology and Medical Oncology (DGHO). *Ann Hematol* 2018; 97:197–207.

- Neoh CF, Snell GI, Kotsimbos T, et al. Antifungal prophylaxis in lung transplantation—a world-wide survey. *Am J Transplant* 2011; 11:361–6.
- Lamoth F, Chung SJ, Damonti L, Alexander BD. Changing epidemiology of invasive mold infections in patients receiving azole prophylaxis. *Clin Infect Dis* 2017; 64:1619–21.
- CRESEMBA® (isavuconazonium sulfate) [package insert]. Northbrook, IL: Astellas Pharma US, Inc.
- Maertens JA, Raad II, Marr KA, et al. Isavuconazole versus voriconazole for primary treatment of invasive mould disease caused by *Aspergillus* and other filamentous fungi (SECURE): a phase 3, randomised-controlled, non-inferiority trial. *Lancet* 2016; 387:760–9.
- Marty FM, Ostrosky-Zeichner L, Cornely OA, et al. VITAL and FungiScope Mucormycosis Investigators. Isavuconazole treatment for mucormycosis: a single-arm open-label trial and case-control analysis. *Lancet Infect Dis* 2016; 16:828–37.
- Cornely OA, Böhme A, Schmitt-Hoffmann A, Ullmann AJ. Safety and pharmacokinetics of isavuconazole as antifungal prophylaxis in acute myeloid leukemia patients with neutropenia: results of a phase 2, dose escalation study. *Antimicrob Agents Chemother* 2015; 59:2078–85.
- Dadwal S, Kriengkauykiat J, Tegtmeyer B, Ito J. Breakthrough invasive fungal infections in patients with hematologic malignancy and hematopoietic cell transplantation receiving isavuconazole for empiric or directed antifungal therapy. *Open Forum Infectious Diseases* 2016; 3:1580.
- Rausch CR, Dipippo AJ, Kontoyiannis DP. Breakthrough invasive fungal infections in adult patients with leukemia receiving isavuconazole. *Open Forum Infectious Diseases* 2017; 4:S718.
- Astvad KMT, Hare RK, Arendrup MC. Evaluation of the in vitro activity of isavuconazole and comparator voriconazole against 2635 contemporary clinical *Candida* and *Aspergillus* isolates. *Clin Microbiol Infect* 2017; 23:882–7.
- Clinical and Laboratory Standards Institute. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi: Approved standard, 2nd ed, CLSI document M38-A2. Wayne, PA: Clinical and Laboratory Standards Institute, 2008.
- Segal BH, Herbrecht R, Stevens DA, et al. Defining responses to therapy and study outcomes in clinical trials of invasive fungal diseases: Mycoses Study Group and European Organization for Research and Treatment of Cancer consensus criteria. *Clin Infect Dis* 2008; 47:674–83.

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Duration of Zika Viremia in Serum

TO THE EDITOR—Zika virus emerged in French Guiana in December 2015 shortly after its first occurrence on the American continent [1]. In this context

of emergence, molecular diagnostic approach is essential to confirm first human cases and to monitor the outbreak. Nevertheless, its use must be based on a robust diagnostic algorithm built from data of Zika virus RNA persistence in body fluids with minimal bias.

Here, we prospectively assessed Zika virus RNA persistence by real-time reverse transcription polymerase chain reaction in the serum of 38 infected-patients at restricted time intervals from onset of symptoms until virus clearance [2]. Detection threshold was 1.8 log₁₀ copies/mL. No censored data and no intermittent virus detection were observed. Patients were included based on Zika virus RNA detection in their serum or urine no longer after 5 days after symptom onset. The cohort comprised 13 females and 25 males with an age of 39 ± 8.8 (mean ± standard deviation) years, recruited between December 2015 and November 2016 in French Guiana, and all provided written informed consent. This study was approved by the independent ethics committee “Comité de Protection des Personnes Sud-Méditerranée I” in Marseille, France. Time to event analyses were performed using the packages survival (v 2.41.3), flexsurv (v 1.1) and survminer (v 0.4.1) in the R statistical environment [3]. An accelerated failure-time regression model (AFT) with a log-logistic error distribution was chosen over other distributions to be fitted to the data based on the lowest Akaike information criterion.

The minimum and maximum Zika virus RNA clearance time observed in our cohort were 3 and 16 days after symptoms onset, respectively. Median time from symptoms onset to Zika virus RNA clearance in serum was similar with both models: 5 days (95% confidence interval [CI] 4 days, 6 days) using the non-parametric Kaplan-Meier method and 4.8 days (95% CI 4.1 days, 5.6 days) using parametric AFT regression (Figure 1). According to the AFT regression, 95% of symptomatic patients would experience a

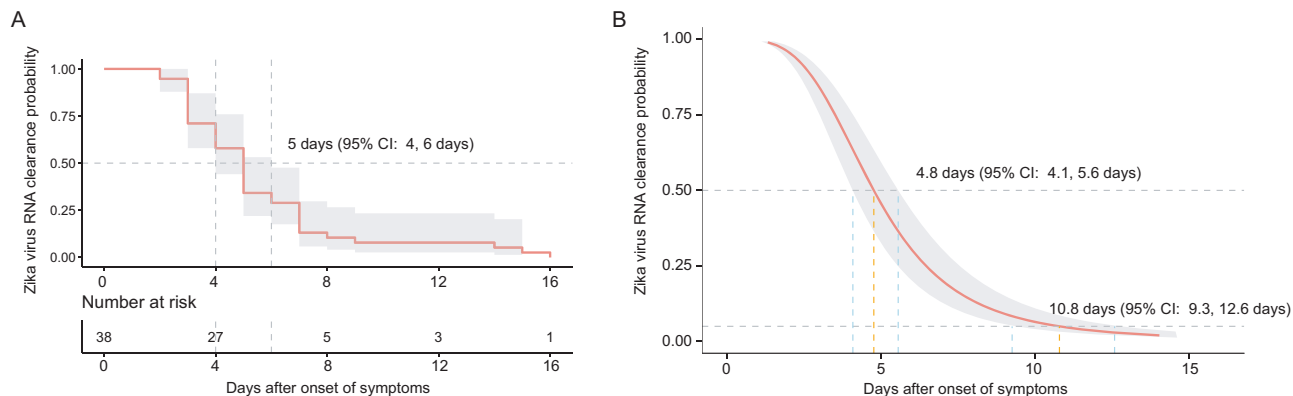


Figure 1. Probabilities of Zika virus RNA clearance according to time after onset of symptoms. Kaplan-Meier estimates (A) and parametric estimates (B) of the Zika virus RNA clearance probability. A table displaying number of individuals at risk of loss of Zika virus RNA detection is provided in panel A. An accelerated failure-time regression model (AFT) with a log-logistic error distribution was used to provide parametric estimates. The medians are indicated for both analyses (A, B) and the 95th percentile is provided for the parametric analysis only (B). Light gray shading denotes 95% confidence intervals. Abbreviation: CI, confidence interval.

Zika RNA clearance after 10.8 days (95% CI 9.3 days, 12.6 days) post symptoms onset. No viral load nor sex effect were observed on time to Zika RNA clearance ($P = .68$ and $P = .45$, log-rank test).

Our results are in accordance with the median estimates assessed by Eui Jeong and colleagues analyzed only from 9 imported Zika cases in South Korea [4]. However, several other studies established a median time to Zika virus clearance after symptom onset that equal or extend 10 days in plasma or whole blood [5–7]. In these works, several biases could have contributed to overestimate the duration of Zika viremia: (i) consideration of asymptomatic patients, (ii) very low sample size, (iii) weekly interval sampling, or (iv) intermittent detection in the follow-up assessment of Zika virus RNA [5–7]. Here we provide estimates of time-dependent probabilities of loss of RNA Zika virus detection after onset of symptoms by analyzing a cohort of an unprecedented sample size. Our results can provide guidance concerning the optimum time window after symptom onset for the implementation of molecular diagnosis tests on serum.

Notes

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References

- de Laval F, Matheus S, Maquart M, et al. Prospective Zika virus disease cohort: systematic screening. *Lancet* 2016; 388:868.
- de Laval F, Matheus S, Labrousse T, Enfissi A, Rousset D, Briolant S. Kinetics of Zika viral load in semen. *N Engl J Med* 2017; 377:697–9.
- R Core Team. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. 2017. Available at <https://www.R-project.org/>
- Jeong YE, Cha GW, Cho JE, Lee EJ, Jee Y, Lee WJ. Viral and serological kinetics in Zika virus-infected patients in South Korea. *Virology* 2017; 14:70.
- Barzon L, Percivalle E, Pacenti M, et al. Virus and antibody dynamics in travelers with acute Zika virus infection. *Clin Infect Dis* 2017; 66:1173–80.
- Mansuy JM, Mengelle C, Pasquier C, et al. Zika virus infection and prolonged viremia in whole-blood specimens. *Emerg Infect Dis* 2017; 23:863–5.

- Paz-Bailey G, Rosenberg ES, Doyle K, et al. Persistence of Zika virus in body fluids—preliminary report. *N Engl J Med* 2017; in press.

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Reply to Fontaine

TO THE EDITOR—We thank Fontaine and colleagues for their report, which estimated the duration of Zika virus (ZIKV) RNA detection in the serum of 38 patients with symptomatic infection, and for their comments [1] on our recent article, which described the dynamics of ZIKV RNA shedding in different body fluids and the antibody response in 30 subjects with acute infection [2]. The median time from symptom onset to ZIKV RNA clearance in serum estimated by Fontaine (5 days, 95% confidence interval [CI] 4.1–5.6) was shorter than the median estimates assessed by our study [2] (11.5 days, 95% CI 9–33) and by the