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#### ▶ To cite this version:

Sébastien Benzekry, Mathieu Grangeon, Mélanie Karlsen, Maria Alexa, Isabella Bicalho-Frazeto, et al.. Machine Learning for Prediction of Immunotherapy Efficacy in Non-Small Cell Lung Cancer from Simple Clinical and Biological Data. Cancers, 2021, 13 (24), pp.6210. 10.3390/cancers13246210. hal-03478003v2

## HAL Id: hal-03478003 https://inria.hal.science/hal-03478003v2

Submitted on 14 Dec 2021

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## Machine learning for prediction of immunotherapy efficacy in nonsmall cell lung cancer from simple clinical and biological data

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#### Highlights:

- Machine learning applied to a large set of NSCLC patients could predict efficacy of immunotherapy with a 69% accuracy using simple routine data
- Hemoglobin levels and performance status were the strongest predictors and significantly associated with DCR, PFS and OS
- Neutrophils-to-lymphocyte ratio was also associated with outcome
- Benchmark of 8 machine learning models

#### **ABSTRACT**

#### Background

Immune checkpoint inhibitors (ICIs) are now a therapeutic standard in advanced non-small cell lung cancer (NSCLC), but strong predictive markers for ICIs efficacy are still lacking. We evaluated machine learning models built on simple clinical and biological data to individually predict response to ICIs.

#### **Methods**

Patients with metastatic NSCLC who received ICI in second line or later were included. We collected clinical and hematological data and studied the association of this data with disease control rate (DCR), progression free survival (PFS) and overall survival (OS). Multiple machine learning (ML) algorithms were assessed for their ability to predict response.

#### Results

Overall, 298 patients were enrolled. The overall response rate and DCR were 15.3 % and 53%, respectively. Median PFS and OS were 3.3 and 11.4 months, respectively. In multivariable analysis, DCR was significantly associated with performance status (PS) and hemoglobin level (OR 0.58, p<0.0001; OR 1.8, p<0.001). These variables were also associated with PFS and OS and ranked top in random forest-based feature importance. Neutrophils-to-lymphocytes ratio was also associated with DCR, PFS and OS. The best ML algorithm was a random forest. It could predict DCR with satisfactory efficacy based on these three variables. Ten-fold cross-validated performances were: accuracy  $0.68 \pm 0.04$ , sensitivity  $0.58 \pm 0.08$ ; specificity  $0.78 \pm 0.06$ ; positive predictive value  $0.70 \pm 0.08$ ; negative predictive value  $0.68 \pm 0.06$ ; AUC  $0.74 \pm 0.03$ .

#### Conclusion

Combination of simple clinical and biological data could accurately predict disease control rate at the individual level.

**Keywords:** Blood counts, lung cancer, response, survival, prediction, machine learning

#### INTRODUCTION

Immune checkpoint inhibitors (ICIs) are now a therapeutic standard in several advanced cancers, particularly in stage IV non-small cell lung cancer (NSCLC) without genetic alteration <sup>1,2</sup>. The development of ICIs is leading to treat an increasing number of patients with these expensive drugs. Even if the overall response rate is higher with ICIs than chemotherapy, it is equal to about 20% for ICIs in monotherapy <sup>2</sup>. Consequently, there are still 4 patients out of 5 with no response to single agent ICI. Thus, identification of predictive markers for ICIs efficacy is an important unmet medical need.

Biologically, ICIs mechanism of action rely on the immune system and tumor micro-environment. Tumor-infiltrating lymphocytes (TILs) are known to have different effects on survival<sup>3</sup>. Blood counts may be a surrogate marker of these TILs and reflect inflammation and adaptive immune response in lung cancer 4. In this respect, analysis of blood counts before the start of ICIs showed some interesting correlation with response. In a meta analysis<sup>5</sup>, particularly in melanomas treated with Ipilimumab, a higher lymphocyte count and relative lymphocyte count predicted better overall survival (OS), as for a higher eosinophil count and lower neutrophil count. Neutrophil to lymphocyte ratio (NLR) at baseline has also been investigated, and its decrease was associated with better OS, progression-free survival (PFS) and response <sup>6</sup>. The derived NLR (dNLR = absolute neutrophils count / [white blood count - absolute neutrophils count]) is another ratio which has already been an alternative to NLR in melanomas <sup>7,8</sup> and metastatic colorectal cancer <sup>9</sup>. Furthermore, for lung cancer, in a Chinese meta-analysis published in 2016 <sup>10</sup>, high levels of platelet to lymphocytes ratio (PLR) at baseline were associated with poor OS and PFS, but in all types of treatment. Specifically for NSCLC treated by ICIs, a study showed that a score combining dNLR greater than 3 with elevated LDH was correlated with worse outcome for ICIs 11. Furthermore, an Italian study in NSCLC patients treated by ICIs 12 showed that low NLR and low PLR at baseline were associated with development of immune related adverse events (IRAEs), and low NLR was associated with better outcomes (OS, PFS). However, a comprehensive analysis of all classical blood markers for prediction of efficacy in a large number of patients is still lacking <sup>13</sup>.

In the era of precision medicine, machine learning (ML) has recently developed as an alternative to classical statistical analysis <sup>14</sup>. The main difference is that statistical analysis focuses on inference and association between variable(s) and outcome(s), while ML puts emphasis on predictive performances only <sup>15</sup>. In oncology, ML has demonstrated great success for prediction from large-dimensional 'big' data, such as genomics or imaging <sup>14</sup>. Nevertheless, such data science methods also have relevance to establish predictive models from smaller sets of variables <sup>16</sup>. In addition, successes have mostly been limited to diagnosis and prognosis but seldom for predictive applications in a clinical oncology context (i.e., for therapeutic decision).

We hypothesized that ML could be useful to accurately predict the efficacy of ICIs in NSCLC patients. The present study aimed to develop a ML model for the selection of patients which could benefit from treatment with ICIs, from simple clinical and biological data.

#### MATERIALS AND METHODS

#### **Patients**

In this observational monocentric retrospective study, we analyzed data from all patients older than 18 years of age who were diagnosed with advanced NSCLC and who received at least 1 cycle of ICI (anti-PD-L1, anti PD-1 or anti-CTLA-4), alone or in combination (anti-PD(L)1 and anti-CTLA4) following a first cycle of non-ICI systemic therapy from April 2, 2013 to February 14, 2017. Patients were treated according to available guidelines.

The study protocol and retrospective data collection were approved by the Institutional Review Board of the French Society of Respiratory Diseases (Société de Pneumologie de Langue Française – SPLF), under reference number: CEPRO 2019-007 and patients have signed an informed consent.

#### Data

Data were retrieved from electronic patient records. Clinical and epidemiological data (age, gender, tobacco status, asbestos exposure, performance status, body mass index), disease characteristics (histology, mutation status, TNM stage), treatment data (type, treatment line, toxicity), biological data (last blood count before first infusion of ICI) and outcome data (response and survival) were collected. From the pre-treatment blood counts, we calculated the PLR, NLR, and derived NLR (dNLR) = absolute neutrophils count / (white blood count – absolute neutrophils count). Performance status was dichotomized into < 2 or  $\ge 2$ .

Tumor response was assessed once every 2 months through computed tomography scans, according to the Response Evaluation Criteria in Solid Tumors, version 1.1.<sup>17</sup> Definition of response here is the best response observed. Overall response rate (ORR) included patients with complete or partial responses. Disease control (DCR) included patients with complete, partial responses or stable disease. Overall survival was defined as the time from start of immunotherapy to death from any cause, censored at the date of last follow-up. Progression-free survival was defined as the time from start of immunotherapy to documented disease progression or death from any cause, censored at the date of last follow-up period.

#### Statistical Analysis

In exploratory data analysis, two-tailed Student's t-tests were used for continuous variables and chi-squared tests for categorical variables. Association of clinical and biological data response was assessed using univariable and multivariable logistic regression, as implemented in the *glm* function of the *R* software (version 3.6)<sup>18</sup>. Survival analyses of OS and PFS were performed using univariable and multivariable proportional hazard Cox's regression models <sup>19,20</sup>. Continuous variables were centered and scaled before these analyses.

#### Machine Learning

Feature selection was performed using feature importance given by a random forest classification algorithm applied on the entire data set (randomForest R package, no tuning, 1,000 trees). Once sorted by mean decrease accuracy, incremental logistic regression models were built with increasing number of variables. The selected optimal set of features was the maximal one before observing a decrease in 10-fold crossvalidated accuracy. Predictive machine learning (ML) models were further built and assessed using repeated nested k-fold cross-validation with 5 repeats of 3 outer loops (to assess generalizability) containing each 5 repeats of 3 inner loops (to tune the models). Thus, 15 train and test sets were built to test the predictive performances. The models were implemented under the *tidymodels* framework in *R* version 4.0.4 <sup>21</sup>. They included: logistic regression (glm), random forest (ranger, 1000 trees), single layer neural network (2600 maximum number of weights), naïve Bayes, k-nearest neighbors (kknn, rectangular kernel function) and support vector machines (with linear, polynomial or radial basis kernel function). For each outer fold, models were tuned and trained on the train set and predictions were assessed on the test set. The decision tree was built using the *rpart* engine and tuned for hyper-parameters of tree depth, minimum number of data points required for further splitting (min n) and complexity parameter (cost complexity), using a grid search and a 10-fold cross-validation. The tree was then trained on the entire set.

#### **RESULTS**

#### Patients and Disease Characteristics

Overall, 298 patients treated with ICIs for stage IV or relapsed NSCLC were retrieved from our database and analyzed. Patient and disease characteristics are summarized in Table 1. Regarding the treatments with ICIs, 89% (n = 266) of patients received an anti-PD-1 antibody, with 96.7% (n = 286) patients being pretreated with chemotherapy prior to ICIs. The report of blood counts values at start of treatment with ICIs are reported in Table S1.

#### Statistical Analysis

#### Response

The DCR was 53.4% and ORR was 15.4%. Exploratory data analysis was conducted for association of the considered variables with outcome (Figure 1). Significant associations were found for NLR (p<0.001), derived NLR (p<0.001), hemoglobin (p<0.0001), leukocytes (p<0.01) and neutrophils (p<0.001). These results were confirmed by logistic regression analysis, with additional significance of (Table 2). However, in multivariable analysis, only hemoglobin and PS remained significant. They also remained significant in multivariable analysis controlling for possible additional confounding factors: sex, immunotherapy type and smoking status.

#### Survival analysis

Progression-free and overall survival are reported in Figure S1. The median PFS was 3.27 months (95%CI: 2.63 - 4.07) and median OS was 14.5 months (95%CI: 8.8 – 15.5). Proportional hazard Cox regression confirmed association of hemoglobin and performance status with response. They were significantly correlated with PFS and OS, in univariable and multivariable analysis (Tables S2 and S3). They also remained significant in multivariable analysis controlling for possible additional confounding factors: sex, immunotherapy type and smoking status.

#### Machine Learning

The ML analysis was conducted for prediction of DCR (classification task) and comprised two steps. First, feature selection and then ML classification. The first step was conducted using random forest-based mean decrease in accuracy (Figure 2A) followed by selection of an optimal number of predictors (Figure 2B). The former

revealed hemoglobin level as the strongest predictor of DCR. The second strongest predictor was performance status, followed by NLR. Adding further predictors resulted in a decrease in the cross-validated accuracy of logistic regression models (Figure 2B). Thus, these three variables were selected for further inclusion in ML models.

Multiple machine learning models using this set of variables were then assessed for their predictive abilities of DCR (Figure 3 and Table 3). First, learning curves –which evaluate the models predictive abilities with increasing number of patients demonstrated that convergence to the optimal predictive power had been reached, for each model (Figure S2). Receiver-operator curves were similarly discriminant across the algorithms (Figure 3A), apart from k-nearest neighbors (knn) and the polynomial support vector machine (SVM) with poorer performances. Aside knn, mean areas under the curve ranged 0.72 – 0.74 (Table 3). Similarly, precision (sensitivity) – recall (positive predictive value) curves were comparable (Figure 3B). Best accuracy was 68%, achieved by the random forest model. Sensitivity was generally low (max 0.58, random forest) while specificity was high (0.73 – 0.94, Table 3). Best positive (naive Bayes, 0.72) and negative (random forest, 0.68) predictive values suggested good predictive power (Table 3). Altogether, the ML algorithms performances suggested the random forest algorithm as the most adequate, achieving highest score in the largest number of them (accuracy, area under the ROC curve, sensitivity and negative predictive value, Table 3) and exhibiting the smallest inter-score variability (Figure 3C).

A random forest algorithm being hard to interpret, we also trained a decision tree algorithm (Figure 3D). This confirmed performance status (<2 versus  $\geq$ 2) and hemoglobin level (optimal threshold 13 g/dL) as the most important predictive variable. Then NLR, consistently with our random forest-based importance analysis. This tree could be useful for clinical decision at bedside. For instance, patients with patient status < 2 and hemoglobin  $\geq$  13 g/dL are predicted to have an 83% chance of disease control.

#### DISCUSSION

The selection of patients who will benefit from ICIs therapies is crucial in the era of precision medicine, in order to develop new strategies for those patients who are not likely to respond from current strategies. Clinical examination and blood counts are easily acquired, but their predictive power in combination remains to be determined. While classical statistical methods are appropriate and have been employed for determination of association with outcome <sup>11,22</sup>, ML techniques are more adapted for prediction tasks. Therefore, we decided to analyze our data with the help of such methods.

Although well developed in several areas of science and industry, especially for dealing with 'big data', the use of ML for clinical oncology has remained limited to date <sup>14,23</sup>. In particular, very few studies have investigated ML for prediction of response to immune-checkpoint blockade, and none has focused on the predictive value of blood counts <sup>16,24–26</sup>. In addition, the main limitation of such studies is the small sample size, despite being a critical determinant to ensure the robustness and generalizability of the results <sup>27</sup>. For instance, Wiesweg et al. had 55 patients in the training set and 36 in the test set <sup>25</sup>.

In comparison, we analyzed the data from 298 patients, allowing to have higher statistical power. Training and test sets were thus composed of about 200 and 100 patients in training and tests, respectively. This large number of patients ensured that the models had enough information to learn and predict (Figure S2). Our results even show that 200 patients are sufficient to reach the optimal accuracy, for models with six variables. Importantly, our data was collected from clinical practice (i.e., real-world data), which implies larger heterogeneity but is also more reflective of the reality at bedside <sup>22</sup>. The random forest algorithm emerged as the algorithm with best trade-off over all the metrics considered, resulting in a 68% mean accuracy and 0.74 mean area under the ROC curve, on test sets. Nevertheless, logistic regression, single layer neural network and naïve Bayes models performed almost equally well, suggesting that similar predictive power can be achieved with a standard, linear statistical learning algorithm such as logistic regression. This is in line with a review that suggested no benefit of ML over logistic regression for clinical prediction models <sup>28</sup>.

Associations between blood count and efficacy were consistent with previous studies, in particularly correlations with hemoglobin, NLR, dNLR and PLR, were the same than in previous studies  $^{5,6,10,11,29}$ . This consistency comforts the interest of pretreatment blood counts for prediction of ICIs efficacy. Neutrophils and leukocytes levels were also associated and predictive of response, a finding from our study that has not been reported elsewhere, to our knowledge. We also demonstrated that an ECOG performance status  $\geq 2$  was significantly associated with response, a result that remained to be fully established  $^{30,31}$ . High PS and a high NLR were correlated with worse outcomes, consistently with the physio-pathological immune hypothesis of peripheral lymphocytes stimulation with ICIs, which could lead to redirect TILs to destruct tumor cells through an activation cascade.

The present study focused on the predictive power of blood biomarkers but the model could include additional variables, even if their predictive values are not perfect individually. PD-L1 expression <sup>32</sup>, because of its discordant results, its heterogeneity among histological components <sup>33</sup> and its poor accuracy when assessed in peripheral blood 34, is not an ideal predictive biomarker but would probably increase the performance of our model. Some characteristics based on genomic alterations could also be added. Tumor mutational burden (TMB), defined as the total number of nonsynonymous mutations in the tumor exome 35 and evaluated by next-generationsequencing (NGS), seems to be a potential biomarker <sup>36</sup>. Even if its measure has some limitation such as high cost, large input DNA amount needed and time of analysis, lung cancer is within tumors with the highest TMB <sup>37</sup>, which correlates with ICI efficacy in NSCLC<sup>38</sup>. Importantly, our results compare favorably with the latter study, where predictive power of (durable) response from PD-L1 expression (AUC = 0.646) or TMB (AUC = 0.601) was inferior. K-RAS mutation is a frequent genetic alteration with contrasting implications for ICIs efficacy. It was initially not linked with poor response to ICIs <sup>39</sup>, but a better response when mutated was shown in 2019 <sup>40</sup>, whereas the cooccurring genomic alteration of K-RAS and STK11/LKB1 was associated with a primary resistance to PD1 axis blockade<sup>41</sup>. Other recent work using metabolomics data obtained from blood sampling demonstrated impressive predictive power of response, although with small sample sizes 42,44. The integration of such markers in our model might improve its efficacy. Based on the combination of quantitative imaging and machine learning, radiomics study have also started to emerge for prediction of ICIs efficacy from radiological images <sup>24,42</sup>. Such studies should nevertheless be carefully evaluated for their added predictive power in comparison to simpler biomarkers as the ones used here <sup>43</sup>.

The first limitation of our study is the retrospective design and the limited number of centers involved (2 centers). Another limitation is the lack of tumor PD-L1 assessment. This testing was not systematically performed from 2013 to 2017 as nivolumab in second-line or more did not require the PD-L1 status to be prescribed to NSCLC patients. This status could have helped our algorithm using a quantitative variable, freeing ourselves from a qualitative threshold as currently used: 1% for the refund of Durvalumab in France for the maintenance after a radio-chemotherapy or pembrolizumab in second-line therapy<sup>44</sup> and 50% for the use of Pembrolizumab in first-line therapy<sup>2</sup>. Nevertheless, the PD-L1 determination methods were not the same as nowadays, involving either a different or biased analysis or a new determination of PD-L1 status.

Certain confounding factors such as bacterial and viral infection, demographic variables such as race, recent chemotherapy, or the use of corticosteroids before treatment could have modified the blood counts analyzed in this study. These data were not collected and not analyzed in our study because we wanted a simple tool to help clinical practice, which could benefit to the majority of patients. Furthermore, blood counts were not necessarily sampled the day of first ICIs infusion, but sometimes one or some days before. This period of time could also have modified the values. On the other hand, our study reflects clinical practice in real-life.

Eventually, the majority of studies to date, including ours, have focused on static pre-treatment predictive markers. However, dynamic markers that integrate ontreatment data start to emerge, with promising predictive value <sup>45</sup>. In such context, the use of statistical Bayesian modeling is particularly appropriate and holds great promise <sup>46</sup>. For ICI, a mathematical model-derived kinetic parameters of tumor kinetics regrowth during relapse has been shown to be the best predictor of overall survival in multivariable analysis including baseline clinical markers <sup>47</sup>. More mechanistic mathematical models for tumor-immune-ICI-radiotherapy dynamics have also been proposed <sup>48</sup>. In light of our results, such models should be applied to include dynamics of blood counts.

### **CONCLUSION**

Blood counts prior to ICIs (elevation of hemoglobin, decrease of NLR, leukocytes or neutrophils) and clinical status (good PS) were significantly associated with better DCR in multivariable analysis. The practical application of these associations using machine learning algorithms was able to predict individual response to treatment. This could be improved further by increasing the number of variables in the model and should be further validated in an independent cohort.

### **ACKNOWLEDGEMENTS**

The authors would like to thank Pauline Fleury for valuable input on the work. This work is part of the QUANTIC project funded by ITMO Cancer AVIESAN and French Institut National du Cancer (grant #19CM148-00)

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# Figures and Tables

Table 1: Patients and disease characteristics

Figure 1: Exploratory data analysis

Table 2: Logistic regression analysis for disease control

Figure 2: Variable selection

Figure 3: Machine learning algorithms predictive performances

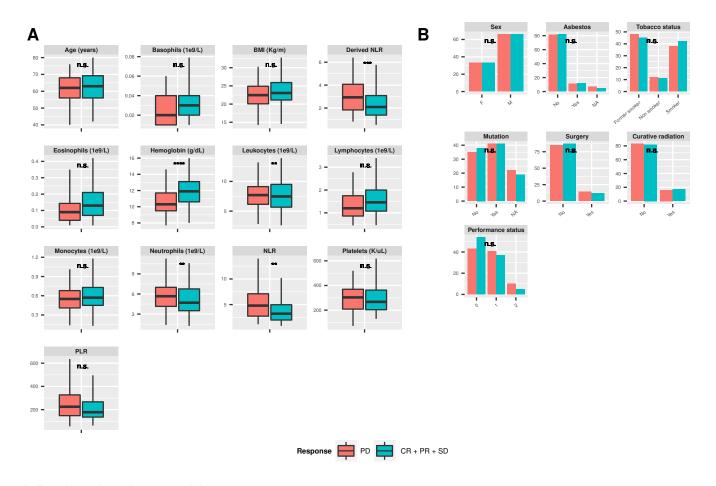
Table 3: Summary of machine learning algorithms predictive performances

**Table 1: Patients and disease characteristics** 

Variable         N = 298¹           Age         62 (55, 69)           Sex         Female         99 (33%)           Male         199 (67%)           Tobacco status         Former Smoker         140 (47%)           Non smoker         36 (12%)           Smoker         122 (41%)           Brain metastases         72 (29%)           Performance status         >=2         26 (8.9%)           0-1         265 (91%)           Mutation profile         ALK         1 (0.8%)           BRAF         9 (7.4%)           EGFR         14 (11%)           KRAS         87 (71%)           Other mutation         5 (4.1%)           ROS1         2 (1.6%)           Wild type         4 (3.3%)           Immunotherapy type           anti-CTLA-4         3 (1.0%)           anti-PD-1         266 (89%)           anti-PD-L1         26 (8.7%)           Combination         3 (1.0%)           Previous treatment         Chemotherapy + immunotherapy         5 (1.7%)           Targeted therapy         11 (3.7%)           Response         Complete response         2 (0.7%)           Progressive disease         131 (45		
Sex           Female         99 (33%)           Male         199 (67%)           Tobacco status         140 (47%)           Former Smoker         140 (47%)           Non smoker         36 (12%)           Smoker         122 (41%)           Brain metastases         72 (29%)           Performance status         72 (29%)           0-1         265 (91%)           Mutation profile         4 (10.8%)           ALK         1 (0.8%)           BRAF         9 (7.4%)           EGFR         14 (11%)           KRAS         87 (71%)           Other mutation         5 (4.1%)           ROS1         2 (1.6%)           Wild type         4 (3.3%)           Immunotherapy type           anti-PD-1         266 (89%)           anti-PD-1         26 (8.7%)           Combination         3 (1.0%)           Previous treatment         2 (8.7%)           Chemotherapy + immunotherapy         5 (1.7%)           Targeted therapy         11 (3.7%)           Response         2 (0.7%)           Complete response         2 (0.7%)           Partial response         44 (15%)           P	Variable	N = 298 <sup>1</sup>
Female       99 (33%)         Male       199 (67%)         Tobacco status       140 (47%)         Former Smoker       140 (47%)         Non smoker       36 (12%)         Smoker       122 (41%)         Brain metastases       72 (29%)         Performance status       →2         ≥=2       26 (8.9%)         0-1       265 (91%)         Mutation profile       ↓1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type       anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       266 (89%)         Combination       3 (1.0%)         Previous treatment       2 (8.7%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Age	62 (55, 69)
Male       199 (67%)         Tobacco status         Former Smoker       140 (47%)         Non smoker       36 (12%)         Smoker       122 (41%)         Brain metastases       72 (29%)         Performance status         >=2       26 (8.9%)         0-1       265 (91%)         Mutation profile         ALK       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       266 (89%)         combination       3 (1.0%)         Previous treatment         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response         Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Sex	
Tobacco status         Former Smoker       140 (47%)         Non smoker       36 (12%)         Smoker       122 (41%)         Brain metastases       72 (29%)         Performance status       -2         >=2       26 (8.9%)         0-1       265 (91%)         Mutation profile	Female	99 (33%)
Former Smoker 140 (47%) Non smoker 36 (12%) Smoker 122 (41%)  Brain metastases 72 (29%)  Performance status  >=2 26 (8.9%) 0-1 265 (91%)  Mutation profile  ALK 1 (0.8%) BRAF 9 (7.4%) EGFR 14 (11%) KRAS 87 (71%) Other mutation 5 (4.1%) ROS1 2 (1.6%) Wild type 4 (3.3%)  Immunotherapy type  anti-CTLA-4 3 (1.0%) anti-PD-1 266 (89%) anti-PD-1 266 (89%) anti-PD-1 266 (89%) Tombination 3 (1.0%)  Previous treatment  Chemotherapy 281 (95%) Chemotherapy + immunotherapy Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Male	199 (67%)
Non smoker       36 (12%)         Smoker       122 (41%)         Brain metastases       72 (29%)         Performance status       265 (91%)         0-1       265 (91%)         Mutation profile       4LK       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type       anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       2 (0.7%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Tobacco status	
Smoker       122 (41%)         Brain metastases       72 (29%)         Performance status       26 (8.9%)         0-1       265 (91%)         Mutation profile       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Former Smoker	140 (47%)
Brain metastases       72 (29%)         Performance status       ≥=2       26 (8.9%)         0-1       265 (91%)         Mutation profile       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Non smoker	36 (12%)
Performance status         >=2       26 (8.9%)         0-1       265 (91%)         Mutation profile	Smoker	122 (41%)
>=2 26 (8.9%) 0-1 265 (91%)  Mutation profile  ALK 1 (0.8%) BRAF 9 (7.4%) EGFR 14 (11%) KRAS 87 (71%) Other mutation 5 (4.1%) ROS1 2 (1.6%) Wild type 4 (3.3%) Immunotherapy type anti-CTLA-4 3 (1.0%) anti-PD-1 266 (89%) anti-PD-L1 26 (8.7%) Combination 3 (1.0%)  Previous treatment Chemotherapy 281 (95%) Chemotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Brain metastases	72 (29%)
0-1       265 (91%)         Mutation profile       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Performance status	
Mutation profile         ALK       1 (0.8%)         BRAF       9 (7.4%)         EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       Chemotherapy         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response         Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	>=2	26 (8.9%)
ALK BRAF 9 (7.4%) EGFR 14 (11%) KRAS 87 (71%) Other mutation FOS1 Wild type 4 (3.3%) Immunotherapy type anti-CTLA-4 anti-PD-1 266 (89%) anti-PD-L1 Combination 3 (1.0%) Previous treatment Chemotherapy Chemotherapy + immunotherapy Targeted therapy 11 (3.7%) Response Complete response Partial response 44 (15%) Progressive disease 131 (45%)	0-1	265 (91%)
BRAF 9 (7.4%) EGFR 14 (11%) KRAS 87 (71%) Other mutation 5 (4.1%) ROS1 2 (1.6%) Wild type 4 (3.3%) Immunotherapy type anti-CTLA-4 3 (1.0%) anti-PD-1 266 (89%) anti-PD-L1 26 (8.7%) Combination 3 (1.0%) Previous treatment Chemotherapy 281 (95%) Chemotherapy 11 (3.7%) Targeted therapy 11 (3.7%) Response Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Mutation profile	
EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       Chemotherapy         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response         Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	ALK	1 (0.8%)
EGFR       14 (11%)         KRAS       87 (71%)         Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       Chemotherapy         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       Complete response         Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	BRAF	9 (7.4%)
Other mutation       5 (4.1%)         ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       281 (95%)         Chemotherapy       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	EGFR	
ROS1       2 (1.6%)         Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       Chemotherapy         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	KRAS	87 (71%)
Wild type       4 (3.3%)         Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment       281 (95%)         Chemotherapy       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Other mutation	5 (4.1%)
Immunotherapy type         anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment         Chemotherapy       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response         Complete response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	ROS1	2 (1.6%)
anti-CTLA-4       3 (1.0%)         anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment         Chemotherapy       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	Wild type	4 (3.3%)
anti-PD-1 266 (89%) anti-PD-L1 26 (8.7%) Combination 3 (1.0%)  Previous treatment Chemotherapy 281 (95%) Chemotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Immunotherapy type	
anti-PD-1       266 (89%)         anti-PD-L1       26 (8.7%)         Combination       3 (1.0%)         Previous treatment         Chemotherapy       281 (95%)         Chemotherapy + immunotherapy       5 (1.7%)         Targeted therapy       11 (3.7%)         Response       2 (0.7%)         Partial response       44 (15%)         Progressive disease       131 (45%)	anti-CTLA-4	3 (1.0%)
Combination 3 (1.0%)  Previous treatment  Chemotherapy 281 (95%) Chemotherapy + immunotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	anti-PD-1	266 (89%)
Previous treatment  Chemotherapy 281 (95%) Chemotherapy + immunotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	anti-PD-L1	26 (8.7%)
Chemotherapy 281 (95%) Chemotherapy + immunotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Combination	3 (1.0%)
Chemotherapy + immunotherapy 5 (1.7%) Targeted therapy 11 (3.7%)  Response Complete response 2 (0.7%) Partial response 44 (15%) Progressive disease 131 (45%)	Previous treatment	
Targeted therapy 11 (3.7%)  Response  Complete response 2 (0.7%)  Partial response 44 (15%)  Progressive disease 131 (45%)	Chemotherapy	281 (95%)
ResponseComplete response2 (0.7%)Partial response44 (15%)Progressive disease131 (45%)	Chemotherapy + immunotherapy	5 (1.7%)
ResponseComplete response2 (0.7%)Partial response44 (15%)Progressive disease131 (45%)	Targeted therapy	11 (3.7%)
Partial response 44 (15%) Progressive disease 131 (45%)		
Progressive disease 131 (45%)	Complete response	2 (0.7%)
,	Partial response	44 (15%)
	Progressive disease	131 (45%)
	Stable disease	113 (39%)

¹Median (IQR); n (%)

Figure 1: Exploratory data analysis



#### A. Boxplots of continuous variables

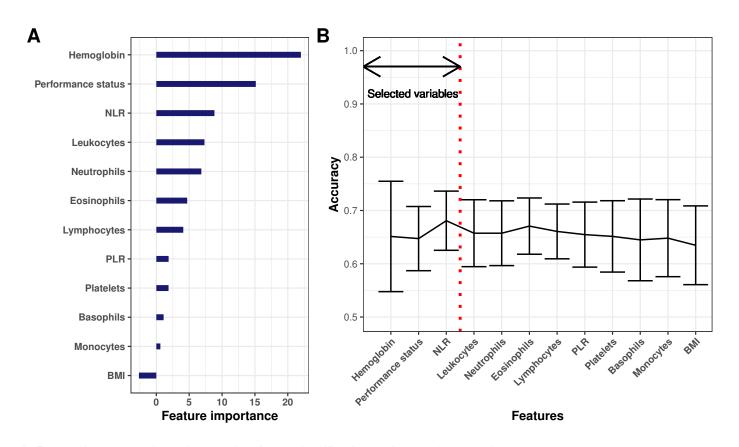
#### B. Barplots of categorical variables

BMI = body mass index, NLR = neutrophils-to-lymphocytes ratio, PLR = platelets-to-lymphocytes ratio, CR = complete response, PR = partial response, SD = stable disease and PD = progressive disease. Stars indicate statistical significance:  $^*$  = p < 0.05,  $^{**}$  = p < 0.01,  $^{***}$  = p < 0.001,  $^{****}$  = p < 0.0001, n.s. = non significant.

Table 2: Logistic regression analysis for disease control

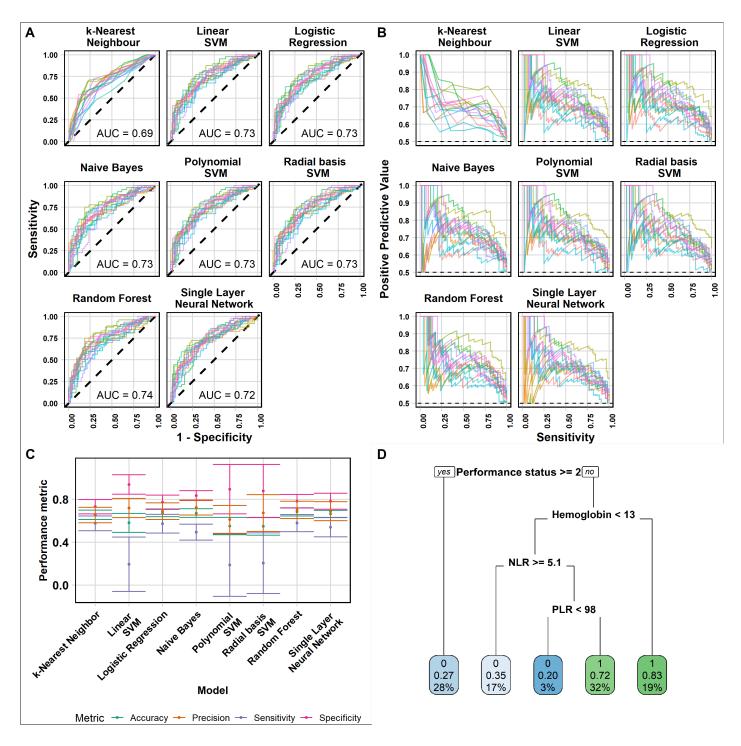
	Univariable logistic regression			Multivariable logistic regression		
Variable	Odds ratio [95% IC]	р	signif	Odds ratio [95% IC]	р	signif
Lymphocytes	1.1 [0.83, 1.4]	0.678		0.98 [0.15, 5.2]	0.984	
NLR	0.49 [0.31, 0.73]	0.000879	***	0.68 [0.098, 1.9]	0.651	
Platelets	1 [0.82, 1.3]	0.762		1.3 [0.72, 2.4]	0.404	
PLR	0.84 [0.64, 1.1]	0.156		1.1 [0.5, 2.4]	0.788	
Leukocytes	0.68 [0.5, 0.89]	0.00791	**	0.6 [0.0022, 3e+02]	0.847	
Hemoglobin	1.9 [1.5, 2.5]	9.26e-07	***	1.8 [1.3, 2.4]	0.000122	***
dNLR	0.63 [0.47, 0.83]	0.00155	**	0.8 [0.33, 2.7]	0.689	
Neutrophils	0.62 [0.45, 0.83]	0.00232	**	1.5 [0.0047, 2.7e+02]	0.863	
Monocytes	0.87 [0.69, 1.1]	0.226		0.86 [0.5, 1.4]	0.545	
Eosinophils	1.3 [0.97, 1.9]	0.139		1.1 [0.75, 1.8]	0.582	
Basophils	1.2 [0.95, 1.8]	0.177		1.2 [0.89, 1.8]	0.321	
ВМІ	1.2 [0.95, 1.5]	0.123		1 [0.76, 1.3]	0.997	
Performance status	0.5 [0.39, 0.64]	6.21e-08	***	0.58 [0.44, 0.75]	7.79e-05	***

Figure 2: Variable selection



- A. Feature importance based on random forest classification and mean decrease in accuracy.
- B. Accuracy score of incremental logistic regression models built on an increasing number of predictors (i.e., the first one contains only hemoglobin, the second hemoglobin and NLR, etc..).
- NLR = neutrophils-to-lymphocytse ratio. PLR = platelets-to-lymphocytes ratio. BMI = body mass index

Figure 3: Machine learning algorithms predictive performances



- A. Receiver-operator curves for prediction on test sets from each fold of the outer cross-validation loop, for each model. AUC = area under the curve.
- B. Precision (positive-predictive value) recall (sensitivity) curves.
- C. Main performance metrics for each algorithm.
- D. Decision tree obtained after tuning and training. Each node shows: the predicted class (0 = PD, 1 = CR+PR+SD), the predicted probability of response and the percentage of total observations in the node. PD = progressive disease, CR = complete response, PR = partial response, SD = stable disease.

Table 3: Summary of machine learning algorithms predictive performances

Model	Accuracy	ROC AUC	PPV	NPV	Sensitivity	Specificity
Random Forest	0.68 ± 0.04	0.74 ± 0.03	0.70 ± 0.08	0.68 ± 0.06	0.58 ± 0.08	0.78 ± 0.06
Logistic Regression	0.67 ± 0.04	0.73 ± 0.03	0.69 ± 0.08	0.67 ± 0.06	0.57 ± 0.09	0.77 ± 0.07
Naive Bayes	0.67 ± 0.04	0.73 ± 0.03	0.72 ± 0.07	0.65 ± 0.06	0.49 ± 0.07	0.83 ± 0.05
Single Layer Neural Network	0.66 ± 0.03	0.72 ± 0.03	0.69 ± 0.09	0.66 ± 0.06	0.54 ± 0.09	0.78 ± 0.07
k-Nearest Neighbour	0.66 ± 0.04	0.69 ± 0.04	0.65 ± 0.07	0.66 ± 0.06	0.58 ± 0.07	0.73 ± 0.07
Linear SVM	0.58 ± 0.09	0.73 ± 0.03	0.72 ± 0.09	0.58 ± 0.10	0.19 ± 0.25	0.94 ± 0.09
Polynomial SVM	0.55 ± 0.08	0.73 ± 0.03	0.61 ± 0.13	0.58 ± 0.13	0.19 ± 0.29	0.89 ± 0.23
Radial basis SVM	0.55 ± 0.08	0.73 ± 0.03	0.67 ± 0.17	0.56 ± 0.06	0.20 ± 0.28	0.88 ± 0.25

# Supplementary figures and tables

Table S1: Blood counts at ICI treatment start

Figure S1: Progression-free and overall survival

Table S2: Cox regression: progression-free survival

Table S3: Cox regression: overall survival

Figure S2: Learning curves

Table S1: Blood counts at ICI treatment start

Variable	Mean	Median	Min	Max
Basophils	0.04	0.03	0.00	0.80
Derived NLR	3.15	2.37	0.41	18.58
Eosinophils	0.16	0.11	0.00	3.26
Hemoglobin	11.32	11.30	7.30	16.40
Leucocytes	8.56	7.99	2.09	49.65
Lymphocytes	1.55	1.35	0.11	19.31
Monocytes	0.63	0.58	0.07	1.96
Neutrophils	6.17	5.52	0.86	46.00
NLR	5.66	3.85	0.46	96.91
Platelets	317	290	57	2,013
PLR	273	214	3	2,309

Figure S1: Progression-free and overall survival

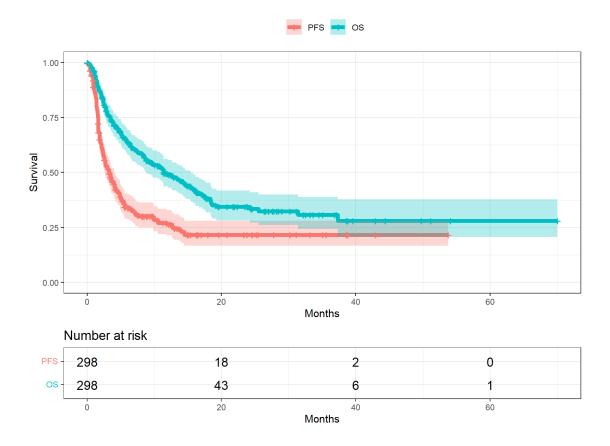


Table S2: Cox regression: progression-free survival

	Univariable cox regression			Multivariable cox regression		
Variable	Hazard ratio [95% IC]	р	signif	Hazard ratio [95% IC]	р	signif
Lymphocytes	0.85 [0.71, 1]	0.0912		1 [0.38, 2.6]	0.996	
NLR	1 [1, 1]	0.00585	**	0.71 [0.33, 1.5]	0.376	
Platelets	1 [1, 1]	0.37		0.81 [0.59, 1.1]	0.206	
PLR	1 [1, 1]	0.881		1 [0.66, 1.5]	0.994	
Leukocytes	1.1 [1, 1.1]	0.00246	**	0.69 [0.044, 11]	0.795	
Hemoglobin	0.9 [0.83, 0.97]	0.00525	**	0.84 [0.72, 0.98]	0.0291	*
dNLR	1.1 [1, 1.1]	0.0017	**	1.2 [0.69, 1.9]	0.596	
Neutrophils	1.1 [1, 1.1]	6.26e-05	***	2 [0.15, 26]	0.603	
Monocytes	1.1 [0.66, 1.8]	0.774		1 [0.78, 1.3]	0.885	
Eosinophils	0.53 [0.23, 1.2]	0.138		0.96 [0.77, 1.2]	0.695	
Basophils	0.91 [0.14, 6.1]	0.923		0.99 [0.87, 1.1]	0.917	
ВМІ	0.96 [0.93, 0.99]	0.0133	*	0.88 [0.76, 1]	0.088	
Performance status	2.1 [1.6, 2.9]	8.15e-07	***	1.3 [1.1, 1.5]	0.000335	***

Table S3: Cox regression: overall survival

	Univariable co	x regression	Multivariable cox regression		
Variable	Hazard ratio [95% IC]	p signif	Hazard ratio [95% IC]	p signif	
Lymphocytes	0.8 [0.64, 0.99]	0.045 *	1.1 [0.4, 3.1]	0.845	
NLR	1 [1, 1.1]	4.54e-08 ***	0.87 [0.43, 1.7]	0.686	
Platelets	1 [1, 1]	0.81	0.83 [0.58, 1.2]	0.305	
PLR	1 [1, 1]	0.331	1 [0.65, 1.6]	0.949	
Leukocytes	1.1 [1, 1.1]	6.63e-07 ***	0.99 [0.054, 18]	0.994	
Hemoglobin	0.83 [0.76, 0.91]	3.58e-05 ***	0.76 [0.64, 0.92]	0.00392 **	
dNLR	1.2 [1.1, 1.2]	8.93e-09 ***	1.4 [0.83, 2.2]	0.225	
Neutrophils	1.1 [1.1, 1.1]	4.05e-10 ***	1.3 [0.087, 19]	0.857	
Monocytes	1.4 [0.85, 2.3]	0.195	1.1 [0.82, 1.5]	0.558	
Eosinophils	0.13 [0.032, 0.54]	0.00509 **	0.78 [0.55, 1.1]	0.166	
Basophils	0.72 [0.071, 7.4]	0.784	0.99 [0.84, 1.2]	0.943	
BMI	0.97 [0.93, 1]	0.0715 .	0.93 [0.8, 1.1]	0.405	
Performance status	2.8 [2, 3.9]	1.12e-09 ***	1.4 [1.2, 1.6]	3.68e-05 ***	

Figure S2: Learning curves

