

Criteria for the Regression of Pediatric Mastocytosis: A Long-Term Follow-Up

Laura Polivka, Julien Rossignol, Antoine Neuraz, Djiba Condé, Julie Agopian, Cécile Méni, Nicolas Garcelon, Patrice Dubreuil, Leïla Maouche-Chrétien, Smail Hadj-Rabia, et al.

▶ To cite this version:

Laura Polivka, Julien Rossignol, Antoine Neuraz, Djiba Condé, Julie Agopian, et al.. Criteria for the Regression of Pediatric Mastocytosis: A Long-Term Follow-Up. Journal of Allergy and Clinical Immunology: In Practice, 2021, 9 (4), pp.1695-1704.e5. 10.1016/j.jaip.2020.12.019 . hal-03476810

HAL Id: hal-03476810 https://inria.hal.science/hal-03476810

Submitted on 24 Apr 2023 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1

Criteria for the regression of pediatric mastocytosis: a long-term follow-up

2

3 Laura Polivka, MD-MSc¹⁻³*, Julien Rossignol, MD-PhD²⁻⁴*, Antoine Neuraz, MD-MSc⁵,

4 Djiba Condé, MSc¹, Julie Agopian, MSc^{3,6,7}, Cécile Méni, MD¹, Nicolas Garcelon, MD-PhD⁵,

5 Patrice Dubreuil, PhD^{3,6}, Leïla Maouche-Chrétien, PhD², Smail Hadj-Rabia, MD-PhD¹,

6 Olivier Hermine, MD-PhD²⁻⁴*, Christine Bodemer, MD-PhD¹*

7 *Equal contribution

8

9 1. Department of Dermatology, Reference Center for Genodermatoses (MAGEC), Necker10 Enfants Malades Hospital (AP-HP5), Paris-Centre University, Imagine Institute, INSERM
11 U1163, Paris, France.

Laboratory of Molecular Mechanisms of Hematologic Disorders and Therapeutic
 Implications, Necker Branch, INSERM U1163 and CNRS ERL 8654, Necker-Enfants

14 Malades Hospital (AP-HP), Paris-Centre University, Imagine Institute, Paris, France.

15 3. CEREMAST, Imagine Institute, INSERM U1163, Paris, France.

16 4. Department of Hematology, Necker-Enfants Malades Hospital (AP-HP5), Paris-Centre

17 University, Imagine Institute, Paris, France.

18 5. Department of Bioinformatics, Necker Branch, Necker-Enfants Malades Hospital (AP-

19 HP5), Paris-Centre University, Imagine Institute, INSERM U1163, Paris, France.

20 6. Centre de Recherche en Cancérologie de Marseille, équipe Labellisée Ligue nationale

- 21 contre le Cancer, INSERM U1068, Marseille, France.
- 22 7. Association Française pour les Initiatives de Recherche sur le Mastocyte et les
- 23 Mastocytoses (AFIRMM), France.
- 24

25	Performed on behalf of the French Reference Center for Mastocytosis (CEREMAST) study
26	group

27

28 **Corresponding authors:**

- 29 Christine Bodemer, Department of Dermatology and Referral Center for Genodermatoses
- 30 (MAGEC), Imagine Institute, Necker-Enfants Malades Hospital, APHP5 and Paris Centre
- 31 University, 149 rue de Sèvres, F-75743 Paris cedex 15, France. christine.bodemer@aphp.fr;
- 32 tel.: +33-144-494-672; fax: +33-144-494-660
- 33 and
- 34 Laura Polivka, Imagine Institute, INSERM U1163, 24 boulevard Montparnasse, F-75015
- 35 Paris, France. laura.polivka@institutimagine.org; tel.: +33-142-754-278; fax: +33-142-754-
- 36 217
- 37
- 38 Word count for text: 3842
- **Figures**: 2
- 40 **Tables:** 6
- 41 **Essential Figures**: 4
- 42 Essential Tables: 4
- 43 Funding sources: This research did not receive any specific funding from agencies or
- 44 organizations in the public, commercial, or not-for-profit sectors.
- 45 **Conflict of Interest**: the authors declare no conflicts of interest for this study.

47 Abstract

Background. Mastocytosis is a neoplastic condition characterized by the accumulation of mast cells (MCs) in one or more organs. Adults tend to have persistent, systemic mastocytosis (SM), whereas MC infiltration in children is usually limited to the skin and typically regresses after several years. Both adults and children could display MC activation symptoms (MCAs) due to MC mediator release. In more than 85% of both adult and pediatric cases, *KIT* mutations are present, the *KIT* D816V mutation being present in most affected adults but in only half of affected children.

55 Objective. To identify the clinical, biological and molecular factors associated with the
56 regression of cutaneous mastocytosis (CM) in children, and to assess the correlation between
57 MCAs and CM regression.

Methods. Patients having suffered from pediatric-onset mastocytosis for at least 8 years were
included in a longitudinal cohort study. Clinical data, the baseline serum tryptase level, the *KIT* sequence, and the progression of MCAs and CM were recorded.

Results. CM regressed in 210 of the 272 included patients (77.2%; mean time to regression: 6.10 years). The rare cases of aggressive SM were symptomatic from the outset. Congenital mastocytosis and the *KIT* D816V mutation were associated with CM regression [odds ratio=0.48 (p=0.031) and 0.173 (p=0.031), respectively]. Aggravation of MCAs over time was correlated with the persistence of skin lesions. However, the MCAs became more intense in 19% of the patients with MCAs at baseline and CM regression - justifying long-term follow-up in this setting.

68 Conclusion. Our results open up new hypotheses with regard to the spontaneous regression of69 CM in pediatric patients.

70	Highlights	box
----	------------	-----

71

73

72 **1. What is already known about this topic?**

In contrast to adult-onset mastocytosis, mast cell infiltration in children is usually limited to
the skin and typically regresses after several years. The underlying mechanisms have not been
yet been identified.

77

78 2. What does this article add to our knowledge?79

In children, congenital disease and the *KIT* D816V mutation were associated with the regression of cutaneous mastocytosis. Symptoms of mast cell activation may worsen over time - even after cutaneous regression. The rare aggressive forms were symptomatic from the outset.

- 84
- 85

87

86 **3.** How does this study impact current management guidelines?

KIT D816V is not associated with a poor prognosis factor in children. Long-term follow-up is
mandatory for children with mastocytosis and symptoms of mast cell activation.

- 90
- 91

92 Keywords

93 Childhood mastocytosis - cutaneous mastocytosis - mast cell - mast cell activation symptom

- 94 *KIT* mutation tryptase prognosis
- 95

96	Abbreviations
97	ASM: aggressive systemic mastocytosis
98	CI: 95% confidence interval
99	CM: cutaneous mastocytosis
100	DCM: diffuse cutaneous mastocytosis
101	MC: mast cell
102	MCAs: mast cell activation symptoms
103	MPCM: maculopapular cutaneous mastocytosis
104	OR: odds ratio
105	SM: systemic mastocytosis

- 106 WHO: World Health Organization
- 107 WT: wild type

109 Introduction

110 Mastocytosis corresponds to a heterogeneous group of neoplastic conditions characterized by the expansion and accumulation of mast cells (MCs) in one or more organs (such as the 111 skin, bone marrow, spleen, lymph nodes, and gastrointestinal tract).¹⁻³ Disease can appear in 112 113 childhood or in adulthood. Pediatric patients generally exhibit cutaneous mastocytosis (CM), 114 in which MC infiltrates are limited to the skin and the resulting skin lesions spontaneously recede after several years in most patients.⁴⁻¹⁰ Congenital mastocytosis is defined by the 115 116 presence of lesions at birth. The clinical manifestations of mastocytosis are related to the 117 clonal accumulation of MCs (depending on which tissues are involved) and the effects of 118 mediators released from the MCs. In fact, over 200 mediators with a role in various 119 physiological and pathological conditions have been identified in MC granules, including heparin and proteases like tryptase and chymase.¹¹ As in cases of adult-onset mastocytosis, 120 121 most of cases of pediatric mastocytosis present symptoms of mast cell activation (MCAs, due to MC mediator release), in addition to skin involvement.^{12,13} Somatic gain-of-function 122 123 mutations in the KIT gene are found in most of patients with mastocytosis. In adult-onset 124 mastocytosis (which is usually persistent and systemic), the KIT D816V mutation (in exon 17) is the most frequent, and is detected in more than 85% of cases.^{14,15} In contrast, a mutation in 125 126 exon 17 is found in only 43% of pediatric cases - the same prevalence as for mutations affecting exons 8, 9, 10 or 11.^{16,17} Although it has been hypothesized that the mutational 127 differences observed between pediatric and adult cases explain (at least in part) the regression 128 of CM, the underlying mechanisms have not been yet identified.^{3,18,19} Nevertheless, a number 129 130 of clinical factors appeared to be associated with spontaneous disease improvement; they include early disease onset (within 12 months of birth) and the type of lesion (e.g. 131 maculopapular cutaneous mastocytosis (MPCM) with large lesions).^{17,20} 132

The often prominent, severe worsening of MCAs can significantly impact the patient's quality of life and can sometimes trigger life-threatening complications; however, the relationship between the regression of MCAs and the regression of skin lesions in pediatric-onset mastocytosis has not been clearly elucidated.

The objective of the present retrospective study of the largest yet cohort of well-characterized patients with pediatric-onset mastocytosis and at least 8 years of follow-up were to (i) identify the clinical, biological and molecular factors associated with spontaneous CM regression in children, and (ii) to analyze the correlation between MCA symptoms and the skin lesion regression.

- 142
- 143
- 144
- 145

146 Material and Methods

147 **Study population**

148 We performed a retrospective study in the dermatology department at Necker Children's 149 Hospital (Paris, France), which coordinates the pediatric branch of the CEREMAST national 150 reference center for mastocytosis. This was an observational study; laboratory tests (KIT gene 151 sequencing and blood chemistry tests) were performed only after the patient's parents had 152 been given a comprehensive explanation and had given their consent (see below). 153 Accordingly, all patients with pediatric-onset mastocytosis who had consulted or been 154 hospitalized between January 1st, 1996, and December 31st, 2018, were considered for 155 eligibility in the study (Figure 1). Mastocytosis was defined and classified according to the 156 World Health Organization (WHO) criteria (see Supplemental Methods). In most cases, the 157 diagnosis of CM was based solely on clear clinical signs (major criteria); for ethical reasons, a 158 skin biopsy (to screen for minor criteria) was not performed. Patients were selected from the 159 CEREMAST national cohort, which was set up as part of a physiopathological study of 160 mastocytosis in which information is stored in electronic medical records. The study protocol 161 was approved by the local investigational review board (CPPRB Groupe Hospitalier de Paris 162 Necker", Paris, France, n°03-01-05; July 7th,2003). Regardless of the type of mastocytosis, 163 KIT sequencing of a skin biopsy was always suggested if it was likely to improve the patient's 164 care. The information extracted from the CEREMAST was supplemented with information 165 from Necker Children's Hospital's data warehouse, which integrates free text data 166 (hospitalization, consultation and prescription reports, etc.) and coded data (laboratory 167 results).²¹ The data warehouse was queried with the following diagnostic codes: 168 "mastocytosis", "cutaneous mastocytosis", "diffuse cutaneous mastocytosis", "mastocytoma", 169 and "urticaria pigmentosum". We next analyzed the characteristics of individual patients by 170 referring to their electronic medical records; erroneous diagnoses, cases of adult-onset 171 mastocytosis, and cases of mastocytosis diagnosed less than 8 years previously were all 172 excluded. Hence, all the included patients presented with confirmed pediatric-onset 173 mastocytosis diagnosed at least 8 years previously. We analyzed the patient's electronic and 174 paper-based medical records, and pictures of their skin.

Patient selection. Between January 1st, 1996, and December 31st, 2018, a total of 687 patients consulted for pediatric-onset mastocytosis at the French National Reference Center. Seventyseven patients were excluded because of an incorrect diagnosis. Overall, 272 of the remaining 610 patients had suffered from pediatric-onset mastocytosis for at least 8 years, and so were selected for long-term follow-up (Figure 1). Twelve patients were lost to follow-up (the changes in CM and MCAs were not documented) and so were excluded from analyses of the regression of CM and MCAs.

182 Data collection. The study variables were assessed twice: at disease onset (during a 183 consultation) and at a study visit (a 'phone interview' or a clinical examination) by 184 dermatologists from the same department at the CEREMAST, using an electronic case report 185 form. For all identified patients, the following items of data were collected: sex; age at onset; 186 age at diagnosis; family history of mastocytosis; the type of cutaneous mastocytosis (MPCM, 187 mastocytoma, or diffuse cutaneous mastocytosis (DCM)), the presence of one or more of the 188 suggested symptoms of MCA: pruritus, flushes, urticaria, diarrhea, abdominal pain, epigastric 189 pain, food reactions, anorexia, nausea, vomiting, loss of consciousness, hypotension, episodes 190 of pallor, a drained feeling, asthenia, headaches, nightmares, sleep disorders, dyspnea, 191 muscle/joint/bone pain, palpitations, polyuria, emotional disturbance or behavioral disorders, 192 the baseline serum tryptase level, KIT gene sequencing data, and the change over time in 193 MCAs and skin lesions. Cases of MPCM were subdivided in three groups: MPCM-large 194 subgroup [patients with at least one large (>1 cm) or polymorphic lesion]; MPCM-small 195 subgroup [patients with small (<5 mm) lesions] and non-classified MPCM (patients for whom

196 the size of the lesions was not known). Congenital mastocytosis was defined by the presence 197 of lesions at birth. The change over time in skin lesions was classified as complete regression 198 (i.e. the healing of lesions, with the absence of Darier's sign), partial regression (a decrease in 199 the number of lesions, and/or lightening of the lesions), or stabilization. The presence or 200 absence of the 26 above-mentioned MCAs was rated (with one point for each symptom) at 201 baseline and at last follow-up. The change over time in MCAs was classified as complete 202 regression, partial regression (a decrease of at least one point), stabilization, or aggravation 203 (an increase of at least one point).

204 Screening for KIT mutations

205 A skin biopsy was collected in 66 of 272 enrolled patient for KIT mutation analysis using a 206 highly sensitive technique (described in the Supplemental Methods) but only if the patient's 207 parents or legal guardian had given his/her informed consent for genetic screening or if the 208 patient had systemic mastocytosis (SM) requiring specific treatment. A bone marrow biopsy 209 with KIT sequencing was performed only in patients (n=7) with signs suggestive of systemic 210 mastocytosis (cytopenia, organomegaly, malabsorption syndrome and bone damage). Three 211 categories of KIT mutation were defined: a point mutation at codon 816, a mutation outside 212 codon 816, and the wild type (WT).

213 Statistical analyses are described in the Supplemental Methods.

214 **Results**

215 Characteristics of the study population

216 The main characteristics of the study population (n=272) are summarized in Table 1. The 217 male-to-female ratio was 1.23, 115 patients (42.3%) had congenital mastocytosis. Twenty-218 three patients (8.5%) had familial mastocytosis (i.e. at least one first-degree parent with 219 mastocytosis). The mean (range) disease duration at the end of our study was 12.93 years 220 (8.1-31.7). We observed 211 (77.6%) patients with MPCM, 46 (16.9%) with mastocytoma, 221 and 15 (5.5%) with DCM. The frequency of congenital forms was higher among patients with 222 mastocytoma (54.3%) or DCM (53.3%) than among patients with MPCM (38.9%), although 223 the intergroup differences were not statistically significant (p=0.069 and p=0.29, 224 respectively). One hundred and ninety-five patients (71.7%) presented with MCAs at 225 diagnosis, with a mean (range) of 2.04 signs (1-11). There were no significant differences 226 between the MPCM, mastocytoma, and DCM groups with regard to the number of MCAs at 227 baseline, except that cutaneous signs of MCA (pruritus and/or flushes) were significantly 228 more frequent in the DCM group (73.3%) than in the MPCM group (40.8%; p=0.016, Table 2 229 and Figure E1). The KIT gene was sequenced (using a skin biopsy sample) in 66 of the 272 230 patients (24.3%): 28 of these 66 (42.4%) carried a mutation in codon 816 (exon 17), 28 231 (42.4%) carried a KIT mutation outside codon 816, and 10 (15.2%) had a WT KIT gene. Fifty of the 66 patients have already been described by Bodemer et al.¹⁶ The different types of *KIT* 232 233 mutation are shown in Table E1. No genotype-phenotype correlations were apparent (data not shown). The mean serum tryptase level at baseline was 7.52 μ g.L⁻¹ [1–60] in the 121 assayed 234 235 patients. As expected, the mean (range) tryptase level was significantly higher in the DCM 236 group [23.99 (2-60) μ g.L⁻¹] than in the MPCM group [7.36 (1 - 45.2) μ g.L⁻¹; p<0.0001] and significantly higher in the MPCM group than in the mastocytoma group $[4.32 \ \mu g.L^{-1}, (1.3 -$ 237 238 11.5), p=0.045] (Figure E1). Apart from the tryptase level, the MPCM, mastocytoma and DCM groups did not differ significantly (data not shown) with regard to the maincharacteristics summarized in Table 1.

241 Within the MPCM group, 85 patients (40.3%) had large MPCM lesions, 78 (37%) had small 242 MPCM lesions and 48 patients (22.7%) had non-classified MPCM lesions. The frequency of 243 familial mastocytosis was significantly higher in the MPCM-small subgroup (17.9%) than in 244 the MPCM-large subgroup (5.9%, p=0.026), whereas the frequency of congenital 245 mastocytosis was higher in the MPCM-large group (48.2% vs. 37.5%, p=0.016). Disease 246 onset after the age of 2 was recorded for 19 patients, all of whom belonged to the MPCM-247 small or -non-classified subgroups. The groups did not differ significantly with regard to the 248 distribution of the KIT mutations (Table 1). The characteristics of the included participants 249 and the screened (source) population were similar (n=610, Table E2). Five of the 610 patients 250 in the source population presented with SM (as defined by the WHO criteria): three of these 251 five had KIT-D816V-positive, congenital, aggressive systemic mastocytosis (ASM) from the 252 onset, with DCM and MC infiltration of the bone marrow, liver, spleen and digestive tract; the 253 remaining two developed acute myeloid leukemia in which a t(8;21) or (1;17) translocation was associated with a KIT D816V or N822Y mutation, respectively (Table E3).¹ 254

255 The main differences between patients for whom mastocytosis was congenital or appeared 256 within one month of birth (the congenital group) and those for whom mastocytosis appeared 257 later than a month after birth (the non-congenital group) are summarized in Table 3. We 258 found that MPCM was significantly more common in the non-congenital group (83.4%) than 259 in the congenital group (70.9%, p=0.019). Mastocytoma and DCM were more common (but 260 not significantly) in the congenital group. The distribution of KIT mutations (notably the 261 frequency of KIT point mutations in codon 816) was similar in the two groups (Figure E2). 262 The distribution of the *KIT* mutations as a function of age of onset is shown in Figure E2.

263 Changes over time in CM

264 Two hundred and ten of the 260 patients (80.8%) showed complete (n=18) or partial (n=192) 265 regression of CM at last follow-up (Figure E3). The mean (range) time to regression was 6.1 266 years (1.48-21.9). Forty-one patients were stable at the last examination, and 9 patients had a 267 greater number of lesions (i.e. worsening). The clinical, biological and molecular variables 268 associated with changes in CM are summarized in Table 4. Congenital mastocytosis was 269 associated with CM regression (odds ratio (OR)=0.476, 95% confidence interval 270 (CI)=[0.236-0.917], p=0.031). The presence of one or more MCAs at baseline was a 271 significant prognostic factor for CM progression (OR=2.20, 95%CI=[1.05 - 5.06], p=0.047). 272 However, each individual MCAs (digestive symptoms, pain, a drained feeling etc.) was not 273 significantly associated with changes over time in CM (data not shown), except for the 274 presence of pruritus or flushes at baseline; surprisingly, these were associated with CM 275 regression (OR=0.19, 95%CI=[0.072-0.43], p=0.000; and OR=0.3, 95%CI=[0.128 - 0.663], 276 p=0.004, respectively). Aggravation or stabilization of MCAs was significantly correlated 277 with worsening CM (OR=3.36, 95%CI=[1.28 - 9.13], p=0.015). Sex, familial mastocytosis, 278 the type of CM, the different types of MPCM and the serum tryptase level at baseline were 279 not significantly associated with the disease outcome.

280 The presence of a KIT point mutation in codon 816 was significantly associated with CM 281 regression (OR=0.173, 95%CI=[0.025-0.719], p=0.031). In contrast, a WT KIT genotype was 282 associated (albeit not significantly) with the aggravation of skin lesions (OR=3, 283 95%CI=[0.666-12.6], p=0.135). Of the 50 stable (n=41) or worsening (n=9) patients, 14 284 patients (12 stable and 2 worsening) had KIT sequence data: two patients presented with a 285 KIT point mutation at codon 816, eight patients presented with other KIT mutations, and four 286 patients were WT (notably two patients with worsening). Two patients developed 287 extracutaneous disease features. A KIT D816V+patient in the MPCM-small subgroup with 288 MCAs from the age of 1 developed a small but painful osteolytic bone lesion with MC infiltrates at puberty but did not meet the criteria for ASM. Another *KIT* D816Y+ patient with
MPCM-large congenital mastocytosis developed hepatomegaly. No liver biopsies were
performed. In both cases, a bone marrow aspirate confirmed the presence of MC infiltrates.
None of the 272 included patients had developed ASM.

293 Changes over time in MCA symptoms

294 Seventy-seven patients (29.6%) were free of MCAs throughout the study. Of the remaining 295 183 patients, 144 (78.7%) showed complete (n=85) or partial (n=59) regression of the MCAs. 296 Twenty-two (12%) patients were classified as stable at the last examination, and the number 297 of MCAs increased in 17 (9.3%) patients (i.e. worsening). The clinical, biological and 298 molecular markers associated with an increase in the number of MCAs are summarized in 299 Table 5. The number of MCAs (and especially the presence of urticaria) at baseline was a 300 prognostic factor for the worsening of MCAs (OR=653, 95%CI=[1.18.10²-1.24.10⁴], p=0.000 301 and OR=4.97, 95%CI=[1.05-26.2], p=0.041, respectively). Taken individually, the other types 302 of MCAs (digestive symptoms, pain, a drained feeling etc.) were not significantly associated 303 with worsening of MCAs (data not shown). Likewise, sex, congenital mastocytosis, familial 304 mastocytosis, the type of CM, the different types of MPCM and the type of KIT mutation 305 were not significantly associated with the course of the MCAs. The serum tryptase level at 306 baseline was not significantly associated with the course of the MCAs (OR=1.05, 307 95%CI=[0.973 – 1.17], p=0.341). In a multivariate analysis, however, a KIT point mutation in 308 exon 816 became significantly associated with the regression of MCAs (Table E4).

309 The correlation between changes in MCAs and CM

Aggravation/stability of MCAs was significantly correlated with worsening/stability of CM (OR=3.36, p=0.015). However, baseline MCAs persisted or even worsened in 27 patients with a favorable CM outcome (Figure 2). The regression of both MCA symptoms and CM was associated with the number of MCAs at baseline and the presence of cutaneous symptoms 314 (pruritus and flushing), relative to patients not showing this correlation (OR=2.5 and 1.97,
315 respectively, p=0.0002, Table 6).

316 Cluster analysis of the different MCAs at diagnosis and changes in CM/MCAs

A cluster analysis of the different MCAs identified four clusters of patients (Figure E4). 317 318 Clinically, cluster 1 was characterized by isolated food intolerance, cluster 2 was not highly 319 specific, cluster 3 was characterized mainly by flushes and pruritus, fatigue with a drained 320 feeling, abdominal pain and intestinal transit disorders, and cluster 4 was characterized by 321 pruritus (but not flushes), abdominal pain, and intestinal transit disorders. The changes over 322 time in each group are depicted in Figure E4. There were no significant intergroup differences 323 with regard to changes over time in CM. Patients in cluster 3 were significantly more likely to 324 experience a decrease in MCAs outcome than patients in cluster 1 (taken as a reference) were 325 (p=0.008, Figure E4). The same association was seen for cluster 4 but was not statistically 326 significant (p=0.08).

328 **Discussion:**

Here, we analyzed a large cohort of patients with pediatric-onset mastocytosis (n=610) and defined prognostic factors in 260 patients with at least 8 years of follow-up (mean duration of disease: 12.93 years). We showed that a spontaneous CM regression was significantly associated with not only congenital disease but also (unexpectedly) a *KIT* D816V mutation. Moreover, even though the worsening of MCAs was correlated with the worsening of CM, MCAs continued to worsen into adulthood in a subset of patients with CM regression. Sex and familial mastocytosis were not associated with changes in CM.

336 The demographic characteristics of our study population were similar to those reported 337 in the literature, namely slight male predominance (male/female ratio: 1.23), disease onset 338 early in life (mean age at onset: 301.2 days), a relatively high frequency of congenital forms 339 (42.3%), a predominance of MPCM (77.6%) relative to mastocytoma (16.9%) and DCM (5.5%), and a mean time to CM regression of 6.1 years (Table 1).⁴ Although aggressive forms 340 341 of SM were rare (n=5 out of 610, 0.82%), the fact that they were always clinically apparent at 342 disease onset meant that we did not systematically have to obtain bone marrow aspirates in 343 children lacking obvious clinical signs of ASM (organomegaly with impaired organ function, pancytopenia, etc.). 344

345 As expected, a large minority (42.4%) of the patients presented with a *KIT* point mutation in 346 codon 816.¹⁶ The present report is the first to show that this mutation is (like congenital CM) 347 significantly associated with CM regression (OR=0.173, p=0.031, Table 4) in pediatric-onset 348 mastocytosis; however, in this study we confirmed the presence of the KIT D816V mutation 349 in only 28 patients. In contrast, the KIT D816V mutation is associated with disease persistence in adult forms of mastocytosis.²² In our study, mutations outside codon 816 did 350 351 not appear to be significantly associated with the change over time in CM; this was probably 352 because the follow-up period was too short.

354 We did not find a KIT mutation in two patients with worsened of cutaneous lesions. As in 355 adult forms of ASM, the absence of a KIT mutation suggests the causal presence of one of 356 more other gene mutations. In the present study, the absence of a KIT mutation appeared to be associated with the worsening of CM (OR=3.0, 95%CI=[0.666 - 12.6], p=0.135, Table 4).²³ 357 358 The distribution of the KIT mutations (notably the KIT point mutation in codon 816) was 359 similar in congenital and non-congenital forms of mastocytosis (Figure E2). Thus, the 360 mutational differences between children and adults with mastocytosis (especially the high 361 prevalence of KIT D816V mutation in adults) cannot alone explain the outcome of CM. Based 362 on Gentek et al.'s research on the ontogeny of MCs, we propose a new hypothesis for the regression of pediatric-onset mastocytosis.²⁴ Gentek et al. showed that in the embryonic 363 364 mouse, MCs have two hematopoietic origins: the yolk sac and the bone marrow. The two MC lineages differ with regard to the phenotype and the gene expression pattern. Interestingly, 365 366 embryonic skin MCs derived from the yolk sac persist after birth and are then gradually 367 replaced by the definitive adult MCs. We therefore hypothesized that regardless of the nature 368 of the KIT gene mutation, mastocytosis will regress over time if the mutation occurs in 369 senescent yolk-sac MCs. Here, we did not find a correlation between the KIT genotype and 370 the patient phenotype; hence, the KIT mutation alone does not explain the heterogeneous 371 symptoms observed in all the various kinds of mastocytosis. This observation strongly 372 suggests that one or more mutations in one or several genes are involved - perhaps in synergy with a *KIT* mutation - in the development of mastocytosis.^{25,26} The association between *KIT* 373 374 mutation and regression may be related to the presence of other beneficial factors such as an increase over time in the expression of MC-inhibiting chondroitin or heparan sulfate.²⁷ 375

377 The aggravation of MCAs was significantly correlated with the worsening of CM (OR=3.36, 378 p=0.015, Table 4). Interestingly, MCAs persisted or even worsened in 27 of the 142 patients 379 (19%) with these symptoms at baseline and a favorable CM outcome (Figure 2) - suggesting 380 that long-term follow-up of these patients might nevertheless be necessary. Thus, we 381 identified two subgroups of patients with CM regression: (i) those in whom both CM and 382 MCAs regressed (n=115), and (ii) those in whom CM regressed despite the worsening or 383 persistence of MCAs (n=27, subgroup 2). The only significant differences between these two 384 groups were the number of MCAs at baseline and the presence of skin symptoms (pruritus and 385 flushing). Surprisingly, pruritus and flushing were more frequent in the "CM and MCA 386 regression" subgroup than in the other subgroup (number of MCAs: 2.5 and 1.97 respectively, 387 p=0.0002, Table 6) - probably because these symptom;s are more strongly associated with the 388 abnormally high MC count than with MC activation per se. The difference between these groups 389 might be related to the presence of mutations or polymorphisms in one or more of the genes 390 involved in MCA; however, addressing this question would require an extensive molecular 391 investigations. The evolution of the levels of serum tryptase, and urinary n-methyl-histamine, 392 urinary 2,3-dinor-11 β -prostaglandin F_{2a}, and urinary leukotriene E4 in this patient population 393 might provide further insights.

394 The present study is the first to have reported on such a large number of cases and to have 395 specified the various MCAs and their changes over time. Symptoms of MCA were frequent in 396 patients with CM (71.7%) and appeared not to be related to the extent of skin involvement 397 (except for pruritus and flushing, Table 2 and Figure E1). Thus, the type and the number of 398 MCAs appear to be more strongly related to a qualitative defect in diseased MCs than to the 399 high MC count per se. Of all the individual MCAs, only the presence of urticaria at diagnosis 400 was significantly associated with a worsening of MCAs in general. At present, clinicians lack 401 a validated scoring system for the qualitative and quantitative evaluation MCAs in children. 402 The development of this type of score might improve patient management in general and the403 assessment of drug efficacy in particular.

404 Our cluster analysis of the MCAs at baseline identified between four clusters of patients 405 (Figure E4). In two clusters (patients presenting with pruritus and flushes at baseline or 406 pruritus without flushes at baseline, respectively), the MCAs decreased over time. These 407 findings will have to be validated in a study with greater statistical power.

408 Although our sample size was large for a rare disease, the study's retrospective design, and 409 the low percentage of patients with *KIT* gene sequence data constituted limitations.

410 **Conclusion**

411 In a retrospective study of the largest yet cohort of well-characterized patients with pediatric-412 onset mastocytosis and at least 8 years of follow-up by the same clinicians, we showed that 413 congenital mastocytosis and the KIT D816V mutation were associated with spontaneous CM 414 regression. Furthermore, even though the worsening of MCAs was correlated with the 415 worsening of CM, we found that MCAs continued to worsen into adulthood in a subset of 416 patients with CM regression. The latter observation suggests that several different 417 mechanisms are involved in the accumulation and the activation of MCs and justifies long-418 term follow-up in these pediatric patients with CM regression.

419

420

422 Acknowledgements

423 The authors acknowledge the contribution and participation of the pediatric mastocytosis 424 study group: Professor Sébastien Barbarot, Dr. Nathalie Beneton, Dr. Benoît Catteau, Dr. 425 Karine Despontin, Dr. Jean Donadieu, Dr. Catherine Eschard, Dr. Marion Gillibert-Yvert, Dr. 426 Anne Hoppe, Dr. Catherine Humiliere, Professor Gisele Kanny, Dr. Laurence Lion, Professor 427 Ludovic Martin, Professor Annabel Maruani, Dr. Guy Molitor, Dr. Alice Phan, Dr. Jean-428 Louis Salomon, Dr. Beatrice Sans, Dr. Francois Skowron, Dr. Jean-Francois Stadler, Dr. 429 Christine Labreze, Dr. Stéphane Barete, Dr. Benoit Catteau, Dr. François Skowron and Dr. 430 Yves Turc (data collection). Dr. Laura Polivka was funded by the Bettencourt Schueller 431 Foundation.

433 **References**

Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Beau MML, et al. The 2016
 revision to the World Health Organization classification of myeloid neoplasms and acute
 leukemia. Blood. 2016;127:2391–405.

437 2. Valent P, Akin C, Metcalfe DD. Mastocytosis: 2016 updated WHO classification and
438 novel emerging treatment concepts. Blood. 2017;129:1420–7.

Hartmann K, Escribano L, Grattan C, Brockow K, Carter MC, Alvarez-Twose I, et al.
Cutaneous manifestations in patients with mastocytosis: Consensus report of the European
Competence Network on Mastocytosis; the American Academy of Allergy, Asthma &
Immunology; and the European Academy of Allergology and Clinical Immunology. J Allergy
Clin Immunol. 2016;137:35–45.

444 4. Méni C, Bruneau J, Georgin-Lavialle S, Le Saché de Peufeilhoux L, Damaj G, Hadj445 Rabia S, et al. Paediatric mastocytosis: a systematic review of 1747 cases. Br J Dermatol.
446 2015;172:642–51.

447 5. Lange M, Niedoszytko M, Renke J, Gleń J, Nedoszytko B. Clinical aspects of
448 paediatric mastocytosis: a review of 101 cases. J Eur Acad Dermatol Venereol JEADV.
449 2013;27:97–102.

450 6. Ben-Amitai D, Metzker A, Cohen HA. Pediatric cutaneous mastocytosis: a review of
451 180 patients. Isr Med Assoc J IMAJ. 2005;7:320–2.

452 7. Güneş Bilgili S, Karadağ AS, Takci Z, Çalka Ö, Kösem M. Comparison of cutaneous
453 mastocytosis with onset in children and adults. Turk J Med Sci. 2014;44:504–10.

454 8. Kiszewski AE, Durán-Mckinster C, Orozco-Covarrubias L, Gutiérrez-Castrellón P,

455 Ruiz-Maldonado R. Cutaneous mastocytosis in children: a clinical analysis of 71 cases. J Eur

456 Acad Dermatol Venereol JEADV. 2004;18:285–90.

457 9. Hannaford R, Rogers M. Presentation of cutaneous mastocytosis in 173 children.

- 458 Australas J Dermatol. 2001;42:15–21.
- 459 10. Azaña JM, Torrelo A, Mediero IG, Zambrano A. Urticaria pigmentosa: a review of 67
 460 pediatric cases. Pediatr Dermatol. 1994;11:102–6.
- 461 11. Mukai K, Tsai M, Saito H, Galli SJ. Mast cells as sources of cytokines, chemokines,
 462 and growth factors. Immunol Rev. 2018;282:121–50.
- Valent P, Akin C, Arock M, Brockow K, Butterfield JH, Carter MC, et al. Definitions,
 criteria and global classification of mast cell disorders with special reference to mast cell
 activation syndromes: a consensus proposal. Int Arch Allergy Immunol. 2012;157:215–25.
- 466 13. Valent P, Akin C, Bonadonna P, Hartmann K, Brockow K, Niedoszytko M, et al.
- 467 Proposed Diagnostic Algorithm for Patients with Suspected Mast Cell Activation Syndrome. J
- 468 Allergy Clin Immunol Pract. 2019;7:1125-1133.e1.
- 469 14. Longley BJ, Metcalfe DD, Tharp M, Wang X, Tyrrell L, Lu SZ, et al. Activating and
 470 dominant inactivating c-KIT catalytic domain mutations in distinct clinical forms of human
 471 mastocytosis. Proc Natl Acad Sci U S A. 1999;96:1609–14.
- 472 15. Arock M, Wedeh G, Hoermann G, Bibi S, Akin C, Peter B, et al. Preclinical human
 473 models and emerging therapeutics for advanced systemic mastocytosis. Haematologica.
 474 2018;103:1760–71.
- 475 16. Bodemer C, Hermine O, Palmérini F, Yang Y, Grandpeix-Guyodo C, Leventhal PS, et
 476 al. Pediatric mastocytosis is a clonal disease associated with D816V and other activating c477 KIT mutations. J Invest Dermatol. 2010;130:804–15.
- 478 17. Meni C, Georgin-Lavialle S, Le Saché de Peufeilhoux L, Jais JP, Hadj-Rabia S,
 479 Bruneau J, et al. Paediatric mastocytosis: long-term follow-up of 53 patients with whole
 480 sequencing of KIT. A prospective study. Br J Dermatol. 2018;179:925–32.
- 481 18. Yanagihori H, Oyama N, Nakamura K, Kaneko F. c-kit Mutations in patients with
 482 childhood-onset mastocytosis and genotype-phenotype correlation. J Mol Diagn JMD.

483 2005;7:252–7.

484 19. Lanternier F, Cohen-Akenine A, Palmerini F, Feger F, Yang Y, Zermati Y, et al.
485 Phenotypic and genotypic characteristics of mastocytosis according to the age of onset. PloS
486 One. 2008;3:e1906.

Wiechers T, Rabenhorst A, Schick T, Preussner LM, Förster A, Valent P, et al. Large
maculopapular cutaneous lesions are associated with favorable outcome in childhood-onset
mastocytosis. J Allergy Clin Immunol. 2015;136:1581-1590.e1-3.

490 21. Garcelon N, Neuraz A, Salomon R, Faour H, Benoit V, Delapalme A, et al. A clinician
491 friendly data warehouse oriented toward narrative reports: Dr. Warehouse. J Biomed Inform.
492 2018;80:52–63.

493 22. Arock M, Sotlar K, Akin C, Broesby-Olsen S, Hoermann G, Escribano L, et al. KIT
494 mutation analysis in mast cell neoplasms: recommendations of the European Competence
495 Network on Mastocytosis. Leukemia. 2015;29:1223–32.

496 23. Molderings GJ. The genetic basis of mast cell activation disease - looking through a
497 glass darkly. Crit Rev Oncol Hematol. 2015;93:75–89.

498 24. Gentek R, Ghigo C, Hoeffel G, Bulle MJ, Msallam R, Gautier G, et al. Hemogenic
499 Endothelial Fate Mapping Reveals Dual Developmental Origin of Mast Cells. Immunity.
500 2018;48:1160-1171.e5.

501 25. Zappulla JP, Dubreuil P, Desbois S, Létard S, Hamouda NB, Daëron M, et al.
502 Mastocytosis in mice expressing human Kit receptor with the activating Asp816Val mutation.
503 J Exp Med. 2005;202:1635–41.

Mayerhofer M, Gleixner KV, Hoelbl A, Florian S, Hoermann G, Aichberger KJ, et al.
Unique effects of KIT D816V in BaF3 cells: induction of cluster formation, histamine
synthesis, and early mast cell differentiation antigens. J Immunol Baltim Md 1950.
2008;180:5466–76.

- 508 27. Gross AR, Theoharides TC. Chondroitin sulfate inhibits secretion of TNF and CXCL8
- from human mast cells stimulated by IL-33. BioFactors Oxf Engl. 2019;45:49–61.

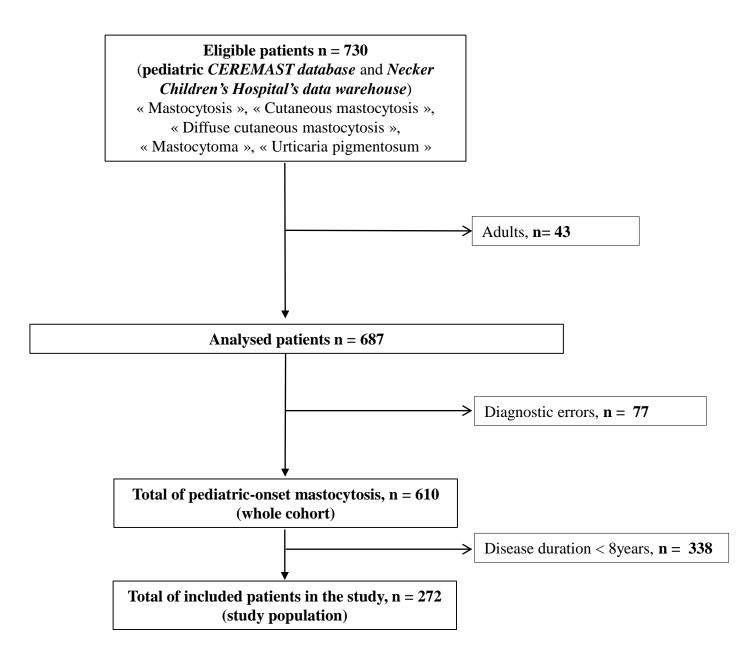
510

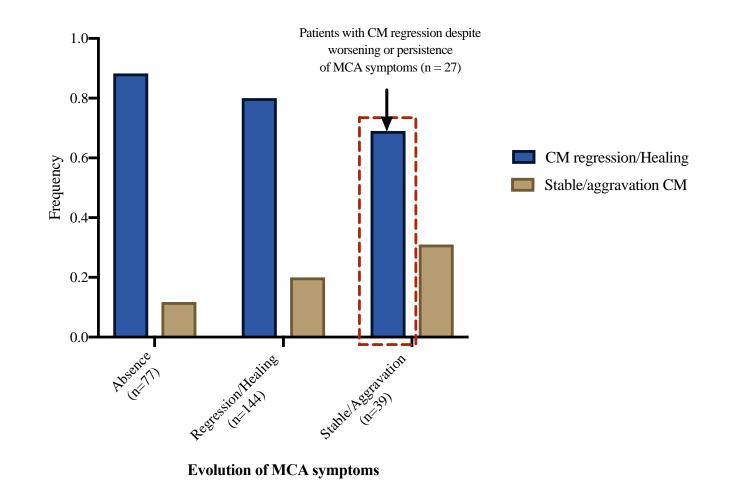
- 512 **Table and figure legends**
- 513 Table 1. Main characteristics of the study population and the different cutaneous514 mastocytosis subtype groups.
- 515 DCM: diffuse cutaneous mastocytosis, F: female, M: male, MCAs: mast cell activation
- 516 symptoms, MPCM: maculopapular cutaneous mastocytosis, NC: non-classified.
- 517 *p<.05
- 518 Table 2. Main mast cell activation symptoms at baseline, as a function of the type of519 cutaneous mastocytosis
- 520 DCM: diffuse cutaneous mastocytosis, MPCM: maculopapular cutaneous mastocytosis.

521 *p<.05

- 522 **Table 3**. Congenital vs. non-congenital mastocytosis
- 523 DCM: diffuse cutaneous mastocytosis, F: female, M: male, MPCM: maculopapular cutaneous
- 524 mastocytosis, NC: non-classified.
- 525 *p<.05
- 526 **Table 4**. Univariate analysis clinical, biological and molecular variables associated with the
- 527 progression of cutaneous mastocytosis
- 528 MCAs: mast cell activation symptoms, MPCM: maculopapular cutaneous mastocytosis.
- 529 *p<.05, **p<.01, ***p<.001
- 530 ¹ Other types of MCAs (digestive symptoms, pain, a drained feeling etc.) were not
- 531 significantly associated with changes in CM (data not shown)
- Table 5. Univariate analysis clinical, biological and molecular variables associated with the
 progression of MCAs.
- 534 CM: cutaneous mastocytosis, MCAs: mast cell activation symptoms, MPCM: maculopapular
 535 cutaneous mastocytosis.
- 536 *p<.05, ***p<.001

- ¹ Other types of MCAs (digestive symptoms, pain, a drained feeling etc.) were not
 significantly associated with the overall progression of MCAs (data not shown)
- 539 **Table 6.** A comparison of clinical, biological and molecular variables in patient group 1
- 540 (correlated regression of both CM and MCAs) and patient group 2 (no correlation).
- 541 CM: cutaneous mastocytosis, DCM: diffuse cutaneous mastocytosis, F: female, M: male,
- 542 MCAs: mast cell activation symptoms, NC: non-classified.
- 543 *p<.05, ***p<.001
- 544
- 545 **Figure 1.** Study flow chart
- 546 **Figure 2**. Correlation between CM regression and changes over time in MCAs.
- 547 CM: cutaneous mastocytosis, MCAs: mast cell activation symptoms, MPCM: maculopapular
 548 cutaneous mastocytosis.
- 549
- 550





For 12 patients, cutaneous and MCA symptoms evolutions were unknown (patients lost to follow-up)

Table 1. Main characteristics of the study population and the different cutaneous mastocytosis subtype groups

Characteristics	Population n = 272 (n/total%)	MPCM n = 211 (n/total%)	MPCM- large n = 85	MPCM- small n = 78	p.value (MPCM- large vs MPCM- small)	MPCM- NC n = 48	Mastocytoma n = 46 (n/total%)	DCM n = 15 (n/total%)
Clinical								
Females	122	93	37	36	-	20	22	7
Males	150	118	48	42	-	28	24	8
Sex ratio (M/F)	1.23	1.27	1.43	1.17	0.75	1.4	1.09	1.14
Mean age at diagnosis (days)	1308.5	1319	1369.3	1348.6	-	1184.5	1263,2	1302
Mean age at onset (days)	301.23	302.22	305.93	302.22	-	305.05	300,87	329,82
Congenital	115 (42.3)	82 (38.9)	41 (48.2)	23 (29.5)	0.016*	18 (37.5)	25 (54.3)	8 (53.3)
Familial mastocytosis	23 (8.5)	22 (10.4)	5 (5.9)	14 (17.9)	0.026*	3 (6.25)	1 (2.2)	0 (0)
Number of MCAs at diagnosis (mean)	2.04	2.09	2.09	2.19	0.75	1.52	1.85	1.93
Absence of MCAs	77 (28.3)	59 (28)	23 (27.1)	15 (19.2)	0,27	21 (43.8)	16 (34.8)	2 ((13.3)
Molecular – Sequencing of <i>KIT</i> gene	66	58	23	26		9	4	4
<i>KIT</i> point mutation at codon 816	28 (42.4)	28 (48.3)	11 (47.8)	12 (46.2)	1	5 (55.6)	0 (0)	0 (0)
Other <i>KIT</i> mutations	28 (42.4)	22 (37.9)	9 (39.2)	10 (38.5)	1	3 (33.3)	3 (75)	3 (75)
<i>KIT</i> WT	10 (15.2)	8 (13.8)	3 (13)	4 (15.4)	1	1 (11.1)	1 (25)	1 (25)
Biological								
Level of baseline tryptase [mean (µg,L ⁻¹), range, number of patients]	7.52 [1 – 60] (n=121)	7.36 [1- 45.2] (n=101)	7.23 [1.5- 23.1] (n=48)	7.55 [1- 45.2] (n=42)	0.82	7.60 [2.9- 18.3] (n=11)	4.32 [1.3 – 11.5] (n=16)	23.99 [2- 60] (n=4)

Table 2. Main mast cell activation symptoms at baseline, as a function of the type of cutaneous mastocytosis

	MPCM (n/total %) n = 211	Mastocytoma (n/total %) n = 46	DCM (n/total %) n = 15	p.value (MPCM vs Mastocytoma)	p.value (MPCM vs DCM)
Cutaneous signs:					
Pruritus/Flushes	86 (40.8)	21 (45.7)	11 (73.3)	0.62	0.016*
Urticaria	5 (2.4)	1 (2.2)	1 (6.7)	1	0.34
Digestive symptoms: anorexia,					
abdominal pain, diarrhea, nausea and /or vomiting	72 (34.1)	11 (23.9)	4 (26.7)	0.22	0.78
Food reactions	32 (15.2)	7 (15.2)	1 (6.7)	1	0.70
Loss of consciousness, drained feeling and/or episodes of pallor	44 (20.9)	7 (15.2)	2 (13.3)	0.54	0.74
Sleep disorders and/or nightmares	17 (8.1)	3 (6.5)	1 (6.7)	1	1
Muscle/joint/bone pain(s)	18 (8.5)	3 (6.5)	2 (13.3)	1	0.63
Emotional disturbance or behavioral disorders	2 (0.9)	1 (2.2)	0 (0)	0.45	1
Pollakiuria	5 (2.4)	1 (2.2)	0 (0)	1	1

 Table 3. Congenital vs. non-congenital mastocytosis

Characteristics	Congenital group n = 127 (n/total %)	Non-congenital group n = 145 (n/total %)	p.value
Clinical			
Females	52 (40.9)	70 (48.3)	
Males	75 (59.1)	75 (51.7)	0.271
Sex ratio (M/F)	1.4	1.07	
Mean age at diagnosis (days)	1237.3	1371.3	0.544 (t test)
Familial mastocytosis	8 (6.3)	15 (10.3)	0.28
МРСМ	90 (70.9)	121 (83.4)	0.019 *
-Large (polymorphic lesions)	44 (48.9)	41 (33.9)	0.033*
-Small (<0.5cm)	27 (30)	51 (42.1)	0.084
-NC	19 (21.1)	29 (24)	0.34
Mastocytoma	27 (21.3)	19 (12.9)	0.26
DCM	10 (7.9)	5 (3.4)	0.12
Molecular – Sequencing of <i>KIT</i> gene	29 (22.8)	37 (25.5)	
<i>KIT</i> point mutation at codon 816	12 (41.4)	16 (43.2)	0.93
Other KIT mutations	13 (44.8)	15 (40.5)	0.804
<i>KIT</i> WT	4 (13.8)	6 (16.2)	1
Biological			
Level of baseline tryptase [mean (µg.L ⁻¹), range, number of patients]	8.92 [1-60] (n=52)	6.53 [1.1-28.2] (n=69)	0.1

Table 4. Univariate analysis – clinical, biological and molecular variables associated with the progression of cutaneous mastocytosis

Variables	Odds Ratio	95%CI	p.value
Clinical			
Sexe	1.46	[0.786-2.72]	0.230
Age at onset	1.18	[0.374-3.14]	0.833
Congenital mastocytosis	0.476	[0.236-0.917]	0.031*
Familial mastocytosis	1.18	[0.374-3.14]	0.757
Mastocytoma	0.594	[0.215-1.40]	0.268
MPCM	1.03	[0.502-2.25]	0.941
Number of MCAs at diagnosis	1.12	[0.997-12.6]	0.051
Presence of MCAs at diagnosis	2.20	[1.05-5.06]	0.047*
¹ Pruritus	0.19	[0.072-0.43]	0.000***
Flush	0.3	[0.128-0.663]	0.004**
Aggravation/Stability of MCAs	3.36	[1.28-9.13]	0.015*
Regression/healing of MCAs	1.92	$[8.88 \times 10^{-1} - 4.53]$	0.112
Biological			
Tryptase	1.01	[0.958-1.07]	0.566
Molecular – Sequencing of <i>KIT</i> gene			
<i>KIT</i> point mutation at codon 816	0.173	[0.025-0.719]	0.031*
Other KIT mutation	2.07	[0.627-7.15]	0.235
<i>KIT</i> WT	3.00	[0.666-12.6]	0.135

Table 5. Univariate analysis – clinical, biological and molecular variables associated with the progression of MCAs

Variables	Odds Ratio	95%CI	p.value
Clinical			
Sexe	1.03	[0.601-1.76]	0.923
Age at onset	1.00	[1.00-1.00]	0.142
Congenital mastocytosis	0.815	[0.476-1.400]	0.454
Familial mastocytosis	3.03	[0.998-1.31]	0.081
Mastocytoma	0.718	[0.368-1.44]	0.339
MPCM	1.09	[0.569-2.03]	0.788
Number of MCAs at diagnosis	653	[1.18.10 ² -1.24.10 ⁴]	0.000***
¹ Urticaria	4.97	[1.05-26.2]	0.041*
Aggravation/Stability of CM	2.20	[1.05-5.06]	0.047*
Biological			
Tryptase	1.05	[0.973-1.17]	0.341
Molecular – Sequencing of <i>KIT</i> gene			
<i>KIT</i> point mutation at codon 816	1.53	[0.427-6.33]	0.525
Other KIT mutation	0.469	[0.124-1.66]	0.243
<i>KIT</i> WT	2.25	[0.363-4.37]	0.464

¹Other types of MCAs (digestive symptoms, pain, drained feeling etc.) were not significantly associated with the overall progression of MCAs (data not shown)

Table 6. A comparison of clinical, biological and molecular markers in patient group 1 (correlated regression of both CM and MCAs) and patient group 2 (no correlation)

N = 142 patients with CM regression and changes in MCAs	Groupe 1 n= 115 (n/total %)	Groupe 2 n = 27 (n/total %)	p.value	
Clinical		· · · · · · · · · · · · · · · · · · ·		
Sex ratio (M/F)	1.25	1.45	0.83	
Familial mastocytosis	14 (12.2)	1 (3.7)	0.0097	
Congenital mastocytosis	48 (41.7)	12 (44)	0.34	
MPCM	89 (77.4)	23 (85.2)	0.44	
Large	38	6	0.16	
Small	37	10	1	
NC	14	7	0.13	
Mastocytoma	20 (17.4)	3 (13)	0.57	
DCM	6 (5.2)	1 (3.7)	1	
Number of MCAs at diagnosis	2.5 [1–11]	1,97 [0 - 4]	0.0002***	
Cutaneous signs: Pruritus/Flushes/Urticaria	81 (70.4)	12 (44.4)	0.014*	
Digestive symptoms: anorexia, abdominal pain, diarrhea, nausea and /or vomiting	58 (50.4)	10 (37.0)	0.28	
Discomfort, loss of consciousness, drained feeling and/or episodes of pallor	35 (30.4)	4 (14.8)	0.15	
Food reaction	28 (24.3)	4 (14.8)	0.44	
Bone, joint and/or muscular pain(s)	15 (13)	0 (0)	0.074	
Biological				
Level of baseline tryptase [mean (µg.L ⁻¹), range, number of patients]	7.54 [1 - 45.2] (n=64)	7.75 [1.5-28.2] (n=11)	0.379	
Molecular – Sequencing of <i>KIT</i> gene	n=36	n=6	-	
Other KIT mutations	13 (36.1)	3 (50)	-	
KIT point mutation at codon 816	18 (50)	3 (50)	-	
<i>KIT</i> WT	5 (13.4)	0 (0)	-	