

Genome wide association study of incomplete hippocampal inversion in adolescents

Claire Cury, Marzia Antonella Scelsi, Roberto Toro, Vincent Frouin, Eric Artiges, Andreas Heinz, Herve Lemaitre, Jean-Luc Martinot, Jean-Baptiste Poline, Michael N Smolka, et al.

▶ To cite this version:

Claire Cury, Marzia Antonella Scelsi, Roberto Toro, Vincent Frouin, Eric Artiges, et al.. Genome wide association study of incomplete hippocampal inversion in adolescents. PLoS ONE, 2020, 15 (1), 10.1371/journal.pone.0227355 . hal-02416654v1

HAL Id: hal-02416654 https://inria.hal.science/hal-02416654v1

Submitted on 17 Dec 2019 (v1), last revised 27 Jun 2023 (v2)

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Genome wide association study of incomplete hippocampal inversion in adolescents

4

5 Claire Cury^{1,*}, Marzia Antonella Scelsi¹, Roberto Toro^{2,3}, Vincent Frouin⁴, Eric Artiges⁵, Antoine
6 Grigis⁴, Andreas Heinz⁶, Hervé Lemaître⁷, Jean-Luc Martinot⁸, Jean-Baptiste Poline^{9,10}, Michael
7 N. Smolka¹¹, Henrik Walter⁶, Gunter Schumann¹², Andre Altmann^{1,‡}, Olivier Colliot^{13,14,15,16,17,‡} and
8 the IMAGEN Consortium[^]

- 9
- 10

11 1 Centre for Medical Image Computing, Department of Medical Physics and Biomedical 12 Engineering, University College London, London, UK

13 2 Centre National de la Recherche Scientifique, Genes, Synapses and Cognition, URA 2182,

14 Institut Pasteur, Paris, France

15 3 Human Genetics and Cognitive Functions, Institut Pasteur, Paris, France.

16 4 NeuroSpin, CEA, Université Paris-Saclay, F-91191 Gif-sur-Yvette, France

17 5 Inserm U 1000 "Neuroimaging & Psychiatry", University Paris Sud, University Paris Descartes

18 - Sorbonne Paris Cité; and Psychiatry Department 91G16, Orsay Hospital, France

19 6 Charité – Universitätsmedizin Berlin, Department of Psychiatry and Psychotherapy, Campus

20 Charité Mitte, Charitéplatz 1, Berlin, Germany

21 7 Inserm U 1000 "Neuroimaging & Psychiatry", Faculté de médecine, Université Paris-Sud, Le

22 Kremlin-Bicêtre; and Université Paris Descartes, Sorbonne Paris Cité, Paris, France

23 8 Inserm U 1000 "Neuroimaging & Psychiatry", University Paris Sud, University Paris Descartes

24 - Sorbonne Paris Cité; and Maison de Solenn, Paris, France

25 9 McGill University, Faculty of Medicine, Montreal Neurological Institute and Hospital, McConnell

26 Brain Imaging Center, Ludmer Centre for Neuroinformatics and Mental Health, Canada.

27 10 Henry H. Wheeler Jr. Brain Imaging Center, Helen Wills Neuroscience Institute, University of

28 California, Berkeley, California, USA

29 11 Department of Psychiatry and Neuroimaging Center, Technische Universität Dresden,

30 Dresden, Germany

- 31 12 Medical Research Council Social, Genetic and Developmental Psychiatry Centre, Institute of
- 32 Psychiatry, Psychology & Neuroscience, King's College London, United Kingdom
- 33 13 Institut du Cerveau et de la Moelle épinière, ICM, F-75013, Paris, France
- 34 14 Inserm, U 1127, F-75013, Paris, France
- 35 15 CNRS, UMR 7225, F-75013, Paris, France
- 36 16 Sorbonne Université, F-75013, Paris, France
- 37 17 Inria, Aramis project-team, F-75013, Paris, France
- 38
- 39 [‡]Shared senior authors
- 40
- 41 ^ Whole author list is given in the supporting information file S1 File and in the acknowledgment
- 42 section.
- 43
- 44 * Corresponding author information:
- 45 Email: <u>claire.cury.pro@gmail.com</u> (CC)
- 46 Current affiliation:
- 47 Univ Rennes, Inria, CNRS, Inserm, IRISA UMR 6074, Empenn (ex VISAGES) ERL U 1228, F-
- 48 35000 Rennes, France
- 49

50 Abstract

51 Incomplete hippocampal inversion (IHI), also called hippocampal malrotation, is an atypical 52 presentation of the hippocampus present in about 20% of healthy individuals. Here we conducted 53 the first genome-wide association study (GWAS) in IHI to elucidate the genetic underpinnings that 54 may contribute to the incomplete inversion during brain development. A total of 1381 subjects 55 contributed to the discovery cohort obtained from the IMAGEN database. The incidence rate of 56 IHI was 26.1%. Loci with P<1e-5 were followed up in a validation cohort comprising 161 subjects 57 from the PING study. Summary statistics from the discovery cohort were used to compute IHI 58 heritability as well as genetic correlations with other traits. A locus on 18q11.2 (rs9952569; 59 OR=1.999; Z=5.502; P=3.755e-8) showed a significant association with the presence of IHI. A functional annotation of the locus implicated genes AQP4 and KCTD1. However, neither this locus 60 61 nor the other 16 suggestive loci reached a significant p-value in the validation cohort. The h^2 62 estimate was 0.54 (sd: 0.30) and was significant (Z=1.8; P=0.036). The top three genetic 63 correlations of IHI were with traits representing either intelligence or education attainment and 64 reached nominal P<=0.013.

66 Introduction

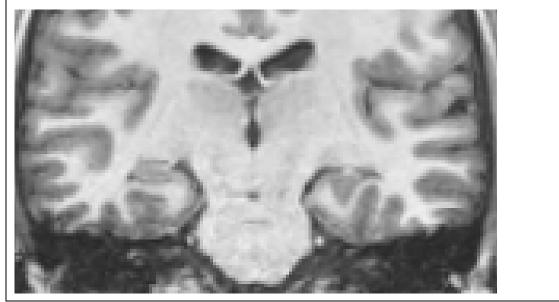
67

68 Human hippocampi are small structures, one in each temporal lobe that belongs to the brain's 69 limbic system and is known to be mainly involved in memory processes such as long term 70 memorisation and spatial navigation [1]. The limbic system and the hippocampus influence the 71 activity of the hypothalamic Pituitary Adrenocortical (HPA) axis, a major neuroendocrine mediator 72 of stress, playing a role in emotional stress responses [2]. Thus, the hippocampus is implicated, 73 with evidence of morphological changes, in a variety of neurological pathologies and psychiatric 74 disorders, such as Alzheimer's disease where hippocampal atrophy increases with the pathology 75 [3]; major depressive disorder where hippocampal volume can predict the response to 76 antidepressants [4,5], is related to suicide attempts [6], and is linked to cortisol disruption 77 (highlighting the implication of the hippocampus in the HPA axis) [7]; Schizophrenia, where 78 patients have smaller hippocampi [8]; or temporal lobe epilepsy, the most frequent form of chronic 79 focal epilepsy in adults, linked to hippocampal sclerosis [9]. Furthermore, during brain 80 development, the growth of the left and the right hippocampi shows distinct responses to postnatal 81 maternal stress [10]. Anatomically, there is a variation to the typical presentation of the 82 hippocampi in normal subjects: the incomplete hippocampal inversion (IHI) also referred to as 83 hippocampal malrotation (Fig 1). This anatomical variant has been initially observed in healthy 84 subjects by [11] and then mostly observed in patients with epilepsy [12,13]. IHIs are mainly left-85 sided and characterized by a rounded or vertical shape, a medial positioning and a deep collateral 86 sulcus [13–15] and are present in around 20% of the normal population [15]. It has been reported 87 that IHI impacts the hippocampal volume: subjects with incomplete inversions appear to have 88 smaller hippocampi [16], and more specifically, the hippocampal subfield CA1 seems to be related 89 to the IHI severity [17]. Also it has been suggested that IHI might interfere with the quality of 90 hippocampal segmentation for volumetric analysis [16,18], which may be clinically relevant, since

91 the hippocampal volume can predict the response to antidepressant in patients without IHI [4.5]. 92 Additionally, a sulcal morphometry analysis suggested that morphological changes associated 93 with IHI are not confined to the hippocampus [15]; significant differences in cortical sulci located 94 along the limbic system are shown between participants with and without complete inversion. 95 Several studies suggest that IHI have their origin in developmental processes [19,20]. For 96 example, [21] observed that during the rotational growth of the hemispheres, the major portion of 97 the hippocampus is carried dorso-laterally and then ventrally to lie in the medial part of the 98 temporal lobe. As the neocortex expands and evolves, the allocortex (the 3 layers cortex) is 99 displaced inferiorly, medially and internally into the temporal horn. This rotational growth of the 100 cortex implies an inversion of the hippocampus during normal development, which in some cases 101 may remain incomplete. Following this hypothesis, [22] conducted a study using foetal MRI and 102 found a correlation between the degree of in-folding and the number of gestational weeks. In a 103 recent study [15] described detailed criteria to evaluate IHI, ultimately making the IHI evaluation 104 more reproducible. In the same study, the introduced criteria had been applied to assess the IHI 105 status of 2000 adolescents without neurological disorders. Results showed a prevalence of about 106 20% of IHI among this normal population. The majority of the IHI cases were left-sided (17% on 107 left side). The lateral preference of left-sided over right-sided IHI may be rooted in the observation 108 of asymmetrical hippocampus development in neonates with the right hippocampus developing faster than the left one [23]. In addition to these developmental observations, IHI has been 109 110 reported to be associated with genetic changes. For instance, IHI was observed at higher 111 prevalence in subjects with chromosome 22q11.2 microdeletion [24], which leads to DiGeorge 112 syndrome.

113

Fig 1: T1 weighted MRI in a coronal view. The left hippocampus (right side in the image) presents an incomplete hippocampal inversion (IHI). The right hippocampus (left side in the image) is an example of a normal or properly inverted hippocampus.



115 116

Given that recent evidence implicates developmental processes in the aetiology of IHI and the observation that the structure and shape of subcortical structures, including the hippocampus, are under genetic control [25], we aimed at elucidating specific genetic variants contributing to IHI. To this end we conducted the first genome-wide association study on the genetics of incomplete hippocampal inversion.

122 Methods

123

124 Subjects

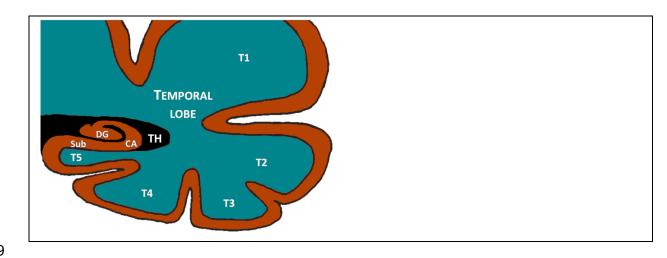
Subjects were investigated from two cohorts: IMAGEN [26] and PING [27]. The IMAGEN cohort comprises >2000 subjects collected at eight sites across Europe [26], and local ethics committee approved the study (see at the end of the paper for details and study [26]). At the time of baseline 128 data collection and study inclusion all participants were 14 years of age. The second cohort was 129 obtained from the Pediatric Imaging Neurocognition and Genetics (PING) Study database 130 (http://ping.chd.ucsd.edu/). PING was launched in 2009 by the National Institute on Drug Abuse 131 (NIDA) and the Eunice Kennedy Shriver National Institute Of Child Health & Human Development 132 (NICHD) as a 2-year project of the American Recovery and Reinvestment Act. The primary goal 133 of PING has been to create a data resource of highly standardized and carefully curated magnetic 134 resonance imaging (MRI) data, comprehensive genotyping data, and developmental and 135 neuropsychological assessments for a large cohort of developing children aged 3 to 20 years. 136 The scientific aim of the project is, by openly sharing these data, to amplify the power and 137 productivity of investigations of healthy and disordered development in children, and to increase 138 understanding of the origins of variation in neurobehavioral phenotypes. Access to the dataset 139 was granted through a Federal Wide Assurance (FWA). For up-to-date information, 140 see http://ping.chd.ucsd.edu/ and [27]. All methods were performed in accordance with relevant 141 guidelines and regulations.

142 Image data processing and IHI scoring

143 The procedure for scoring IHI [15], which has been previously described in detail and shown a 144 good intra- and inter-reproducibility [15], was applied to the subjects used in this study (from 145 IMAGEN and PING). Inter- and intra-rater variability were assessed in a previous publication [15]. 146 This was studied on 42 participants from the discovery cohort using the kappa statistic. In all 147 cases, intra- and inter-rater agreements were beyond substantial ($\kappa \ge 0.64$). Very strong 148 agreements ($\kappa \ge 0.8$) were observed in the majority of comparisons (14/20). Rating on the 149 validation cohort was conducted after by a single rater (CC), thus the rater was not blinded to 150 whether subjects were from the discovery or validation cohort. In brief, the IHI score is composed 151 of four different criteria: (1) assessing the roundness of the hippocampal body; (2) evaluating the verticality of the collateral sulcus which is located between the 4th and the 5th temporal lobe 152

153 convolution (Fig 2); (3) the mediality of the hippocampal body; and (4) the depth of the fusiform 154 gyrus, separating the 4th and the 3rd convolution of the temporal lobe (Fig 2). Each criterion is 155 assessed from a coronal point of view after registering the subjects' T1 weighted MRI into the 156 standard MNI space using the FSL's affine transformation FLIRT [28,29]. Evaluation was carried 157 out using an inhouse Java interface (https://github.com/cclairec/viewerIHI java). During scoring 158 each criterion received a score between 0.0 and 2.0. The first three criteria have a step size of 159 0.5, the fourth criterion is binary (0 or 2), and the 5th criterion, assessed between 0 and 2, has a 160 step size of 1.0. The sum of those criteria forms the overall IHI score ranging from 0.0 to 10.0. 161 This is a semi-continuous score (with a step of 0.5), where an IHI score of 0.0 indicates the total 162 absence of IHI, and a score of 8.0 represents a very pronounced presentation of IHI. In their 163 previous study [15] established an optimal cut-off (at 3.75) of the overall IHI score to indicate 164 presence or absence of IHI, by maximising the accuracy of the classification of a global criterion 165 (blind to individual criteria or IHI scores), indicating if a given hippocampus presents or not an IHI 166 (an intermediate score for partial IHI were present but not used in the estimation of the optimal 167 cut-off): hippocampi without IHI correspond to IHI score < 4.0 and hippocampi with IHI correspond 168 to IHI scores \geq 4.

Fig 2: Hippocampal anatomy in coronal view. The hippocampus comprises the Dentate gyrus (DG), the cornus ammonis (CA) and the subiculum (sub). The temporal lobe is composed of five convolutions: T1, T2, T3, T4 and T5. The collateral sulcus divides T5 from T4 and the sulci of the fusiform gyrus separates T4 from T3. TH indicates the location of the temporal horn of the lateral ventricles.



169

170

For this genetic study, the phenotype was IHI in either left or right hippocampus. To determine IHI, we applied the same cut-off of 4.0 for left and right hippocampi and used, for the IMAGEN cohort, the previously processed data from the IMAGEN study [15].

174

175 SNP genotyping and pre-processing

176 IMAGEN subjects were genotyped from blood samples on 610-Quad SNP and 660-Quad SNP 177 arrays from Illumina. Genetic data was available for 1,841 subjects. In a first round of quality 178 control (QC) we performed subject-level QC by removing subjects with mismatching self-reported 179 sex and genotype inferred sex (N=10) or where more than 10% of SNPs were missing (N=0). 180 Next, we performed ancestry matching based on the HapMap3 data [30]. Population outliers were 181 defined as subjects exhibiting more than five standard deviations distance from the CEU and TSI 182 population in any of the first five principal components. Based on these criteria, 220 subjects were 183 excluded from further analysis (S1 Fig). For the remaining subjects the genetic relationship matrix 184 (GRM) was computed on common SNPs (minor allele frequency [MAF] >5%) after LD pruning 185 using GCTA [31]. Another 18 subjects were removed due to relatedness (i.e., PIHAT > 0.05) 186 leaving a total of 1593 subjects for the analysis. The raw genotyping data were prepared for 187 imputation using a series of scripts (http://www.well.ox.ac.uk/~wrayner/tools/). Haplotype 188 reference consortium (HRC) v1.1 [32] SNPs were imputed on the Sanger imputation server 189 (https://imputation.sanger.ac.uk) using EAGLE2 [33] for pre-phasing and PBWT [34] for 190 imputation. Data from the two different genotyping chips were imputed independently. Genotypes 191 were hard called based on the maximal genotype posterior probability with a threshold of 0.9. 192 That is, if none of the three genotypes reached a posterior probability of at least 0.9, then the SNP 193 was set to missing in the corresponding subject. Finally, an additional round of QC was conducted 194 on SNP level based on imputation quality (INFO score > 0.3), missingness (< 5%), minor allele 195 frequency (MAF>1%) and deviation from Hardy-Weinberg-Equilibrium (p<1e-6) leaving 6,742,645 196 SNPs across the autosomes for the association analysis.

PING subjects were genotyped from saliva samples on Human660W-Quad arrays from Illumina. After QC, genetic data for 1,391 participants was suitable for analysis. Individual SNPs of the PING dataset were accessed through the PING data portal (ping-dataportal.ucsd.edu). Ancestry and admixture proportions in the PING participants were based on the ADMIXTURE software [35] and downloaded through the data portal (for details see [27]). We restricted the validation cohort to participants of at least 12 years of age and of European ancestry (minimum 90% European ancestry as per ADMIXTURE; N=197).

204

205 Genome wide association study

The genome wide association study was carried out with Plink v1.9 [36] assuming an additive genetic model and computing for every SNP a logistic regression while correcting for sex, age at imaging (in days) and five principal components for population structure. Phenotype or covariate information was missing for 212 participants. Thus, the discovery GWAS comprised 1,381 unrelated subjects. The genome-wide statistical significance threshold was set to the standard threshold of p<5e-8 and regional association plots were generated with LocusZoom [37]. SNPs exceeding the threshold for suggestive association with IHI (p<1e-5) were followed up in an
independent cohort of adolescents (PING). In case the top SNP was not genotyped in PING,
LDlink [38](<u>https://analysistools.nci.nih.gov/LDlink/</u>) was used to identify a proxy in LD (r2) within
+/- 50kb of the top SNP's location. Association with single SNPs was tested in R using the glm
function; the logistic model was corrected for age and sex.

217

Functional annotation of GWA summary statistics

The GWA summary statistics were annotated using the web-based version of the FUnctional Mapping and Annotation (FUMA) tool [39] (http://fuma.ctglab.nl/). In order to elucidate the functional consequences of genetic risk loci, FUMA approaches the mapping in two separate steps: first, lead SNPs are identified and mapped to relevant genes on the basis of strand proximity, expression quantitative trait loci (eQTL) and chromatin interaction; second, the reprioritized genes returned by the first step are annotated with respect to expression levels and overrepresentation in differentially expressed gene sets among a wide range of human tissues.

226 For the purposes of this study, SNP-to-gene mapping was performed according to the 227 following parameters: SNPs with p<5e-8 were identified as lead SNPs, and genomic risk loci were 228 constructed by including SNPs in linkage disequilibrium with independent lead SNPs (LD r^2 >0.6 229 in the 1000 Genomes Phase 3 EUR panel) and with a minimum MAF of 1%. Positional mapping 230 was performed by linking lead SNPs to genes in a 50kb window. Mapping based on eQTL was 231 performed by using only SNP-gene pairs significant at FDR<0.05 in all tissues/cell types from 4 232 data repositories (GTEx [40], the Westra blood eQTL dataset [41], the BIOS QTL browser [42] 233 and BRAINEAC [43]); the available data only covers *cis*-eQTLs with up to 1 Mb distance between 234 SNP and gene. Chromatin interaction mapping was also performed to take into account potential 235 long-range interactions between risk loci and genes due to chromatin folding. We based mapping 236 on interactions significant at FDR<1e-6 in 14 tissue types and seven cell lines from [44]. We also based mapping on tissue/cell type specific enhancer or promoter regions annotated in 111
epigenomes from the Roadmap Epigenomics Project [45]. The Major Histocompatibility Complex
(MHC) was excluded from annotations, and mapping to all functional gene classes (proteincoding, non-coding RNA, long intergenic ncRNA, processed transcripts, pseudogenes) was
enabled.

242 After mapping lead SNPs to relevant genes, we performed annotation of the prioritized 243 genes in biological context, mainly with respect to tissue-specific expression levels. Average 244 expression levels (log₂ Read Per Kilobase per Million (RPKM+1)) of protein-coding genes in 53 245 tissues from GTEx v6 were visualized through heat maps, allowing for comparison of expression 246 level across genes and tissue types. Candidate genes were tested for overrepresentation in sets 247 of differentially expressed genes (DEG), as well as sets of genes up- and down-regulated, across 248 53 specific tissue types from GTEx v6 using hypergeometric tests. The same gene-set enrichment 249 analysis strategy was applied to test for overrepresentation of biological functions among the 250 prioritized genes, using gene sets from the Molecular Signatures Database version 5.2 [46], 251 WikiPathways [47] and the GWAS catalog [48], and applying the Benjamini-Hochberg multiple 252 testing correction procedure.

253

254 Heritability analysis and genetic correlation

We used LD score regression [49] in order to estimate IHI heritability from the GWAS summary statistics data. Next, we computed partitioned heritability estimates using the LD score method described in [50] and [51]. Heritability estimates were partitioned into 53 overlapping functional categories, derived from 24 main annotations, from [50]. Stratified LD score regression was also used to test for heritability enrichment in genes specifically expressed in a number of tissues of cell types. For this analysis, we used the specifically expressed gene lists compiled by [51] for the following datasets: expression levels from RNA-seq experiments in the 53 GTEx tissues and cell types, as well as only the 13 GTEx brain regions; the Cahoy dataset, comprising microarray expression data from three cell types (astrocyte, neuron, oligodendrocyte) in the mouse brain [52]; the Franke dataset, comprising microarray expression data in 152 tissues and cell types from human, mouse and rat [53]; and the Immunological Genome Project Consortium dataset, comprising microarray expression data for 292 immune cell types in the mouse [54]. Enrichment p-values were corrected for multiple comparisons using the Benjamini-Hochberg procedure.

Given the reported higher prevalence of IHI in patients with epilepsy, we used LD score regression to compute the genetic correlation [55] between IHI and epilepsy susceptibility based on a recent GWAS [56]. Finally, we conducted an exploratory analysis of genetic correlation between IHI and traits from 832 GWASs using the LD hub [57] (http://ldsc.broadinstitute.org/).

272

Gene expression in the developing human brain

274 In order to explore the transcription pattern of two candidate genes, we downloaded their 275 expression values from BrainSpan through the web interface 276 (http://www.brainspan.org/rnaseg/search/index.html). The data comprises post-mortem gene 277 expression data of 42 subjects at ages spanning from prenatal development (eight post 278 conception weeks) till adulthood (40 years). Brains were sampled across 26 brain structures. 279 Gene expression was measured using RNA-sequencing and expression levels for each gene 280 were provided as reads per kilobase of exon model per million mapped reads (RPKM). We 281 analysed this data using a linear mixed effects model implemented in the Ime4 package in R. In 282 these analyses, the gene expression level was the target variable, subject ID and structure ID 283 were random effects, while an indicator variable for age less than 25 weeks post conception was 284 the fixed effect. We tested for the significant effect of age<25 post conception weeks (pcw) on 285 gene expression. This threshold was selected based on the estimated occurrence of hippocampal 286 inversion between pcw 20 and 30 [20].

287 **Results**

288 Subjects: cohort statistics

In IMAGEN 1381 subjects had genotyping and all phenotype and confounding information available. Incidence rate of IHI was 26.1%. In PING, for the 197 European subjects aged 12 years or older, we could successfully access and score 161 T1 weighted MR images for analysis, and IHI incidence rate was 23.6%; both at the 4.0 cutoff. There was a higher incidence rate of IHI in the left hemisphere in both cohorts. Summary statistics for both cohorts can be found in Table 1.

294	Table 1: Cohort summa	ary statistics of	participants used	for the genetic analyses.
-----	-----------------------	-------------------	-------------------	---------------------------

Cohort	IMAGEN	PING		
Participants (N)	1381	161		
Female (%)	687 (49.7)	78 (48.4%)		
Age (SD)	14.5 (0.41)	16.06 (2.54)		
IHI (%)	360 (26.1)	38 (23.6)		
Left/Right/Bilateral	251/46/63	24/7/7		

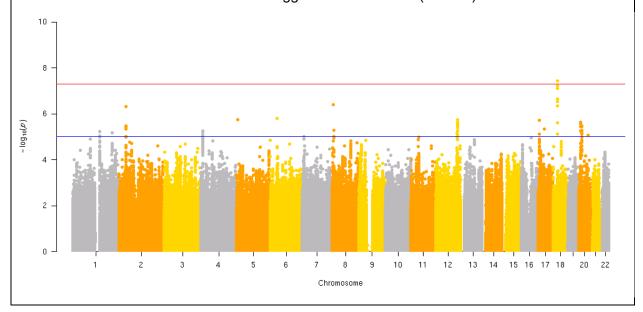
295

Genome wide association study and functional annotation

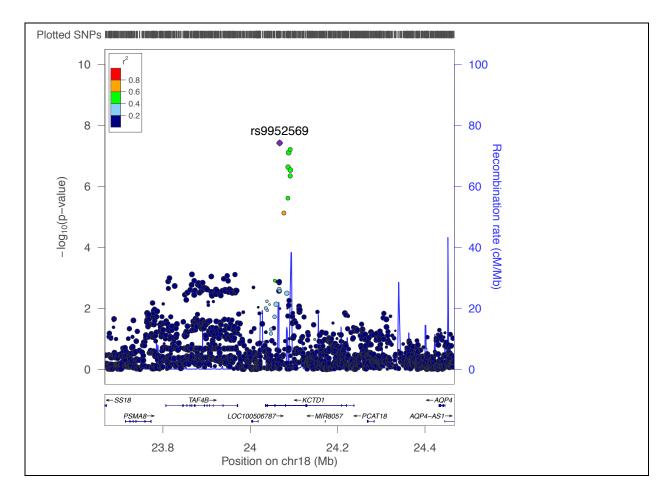
297 We tested each of the 6.7mio SNPs for an association with the presence of IHI. In the discovery 298 dataset comprising subjects from the IMAGEN study, 17 loci passed the threshold for suggestive 299 association (Fig 3; Table 2). One locus on chromosome 18 reached genome-wide significance 300 (top SNP: rs9952569; OR=1.999; Z=5.502; P=3.755e-8; Fig 4). There was no inflation in p-values 301 (λ =1.017; S2 Fig). The top SNP shows a consistently strong association with the continuous IHI 302 score (i.e., not applying the cutoff at 4.0) and the global criterion (C0), but misses the genomewide significance threshold in both cases (S3 Fig: beta=0.5021, Z= 5.129, P=3.332e-07 for the 303 304 continuous score and beta= 0.2542, Z= 5.299, P=1.354e-07 for C0). Functional annotation of the

- 305 GWAS result was carried out using FUMA and linked the significant locus to six genes: AQP4,
- 306 AQP4-AS, CIAPIN1P, KCTD1, RNU6-1289P, and U3 (S4 Fig. left). In fact the top associated SNP
- 307 is located in an intron of *KCTD1*. Brain gene expression based on GTEx shows high and brain-
- 308 specific expression for AQP4 and moderate to high expression levels for KCTD1 (S4 Fig, right).

Fig 3: Manhattan plot. The y-axis depicts the -log10(p-value) of the association between SNP and presence of IHI assuming an additive model in the discovery cohort. The SNPs tested in the study are ordered along their chromosomal position on the x-axis. The red horizontal line donates genome wide significance at the Bonferroni threshold (P=5e-8), while the blue horizontal line marks the threshold for suggestive association (P=1e-5).







310

The gene set enrichment analysis showed that the four protein-coding genes prioritized by FUMA (*AQP4*, *AQP4-AS*, *KCTD1* and *U3*) were statistically significantly enriched (p<0.000314; Bonferroni corrected for 3 * 53 tests) in overexpressed genes in hippocampal and caudate tissue from GTEx v6 (S5 Fig).

We tested the 17 top SNPs within each suggestive loci in the PING cohort for an association with IHI. In PING, there were either genotyped or SNPs in near perfect LD (r2>0.9) for seven loci, intermediate (0.25 < r2 < 0.9) proxies for five loci and weak proxies for the remaining five loci (r2 < 0.25). None of the selected candidate SNPs showed a nominal significant association (uncorrected p < 0.05) with IHI in this cohort. The top SNP from the discovery cohort, rs9952569, was not significant in the validation sample and showed an effect in the opposite direction (OR=0.597; P=0.29; Table 2). Table 2: Suggestive loci from discovery GWAS and results in validation cohort (below). Left side: The table displays the leads SNPs in the discovery cohort (IMAGEN) with p<1e-5 along with their chromosome, base pair (BP), alleles (A1, A2), subjects (N), odds ratio (OR), statistics (STAT) and P-value. Right side: The second table depicts the p-values for the SNPs (VALSNP) in the validation cohort (PING), along with their base pair, correlation to the discovery SNP (R2), alleles (A1, A2), OR and P-value.

326

	Discovery								Validation								
CHR	SNP	BP	A1	A2	NMISS	OR	STAT	Р	CHR	VALSNP	BP	R2	A1	A2	OR	STAT	Р
1	rs79030318	147233364	А	G	1354	1.773	4.534	5.79E-06	1	rs12061877	147264252	0.755	С	Т	1.5584465	1.0434	0.2331419
1	rs3009985	212191991	G	А	1369	1.513	4.501	6.77E-06	1	rs3738449	212240063	0.899	А	G	0.8060804	-0.7252	0.4743359
2	rs10490445	37617484	G	Т	1379	1.812	5.027	4.97E-07	2	rs10490445	37617484	1	G	Т	1.5846445	1.0391	0.1837016
4	rs6852934	7760243	А	G	1379	1.948	4.538	5.68E-06	4	rs2285769	7761286	1	А	G	1.4751031	0.5908	0.3685725
5	rs77126180	6478922	А	G	1337	3.31	4.78	1.75E-06	5	rs272456	6484012	0.042	А	G	0.7735566	-0.2829	0.3407685
6	rs35806781	37081139	А	G	1344	4.889	4.798	1.60E-06	6	rs2395670	37082809	0.011	А	G	1.3437002	1.1122	0.2280719
7	rs11764012	8207615	А	G	1348	0.6388	-4.427	9.56E-06	7	rs2058519	8214574	0.478	G	А	1.2305445	-0.0447	0.4622293
8	rs1870396	9261456	G	С	1333	1.741	5.065	4.08E-07	8	rs6601306	9264711	0.224	Т	С	0.8644291	-1.1714	0.7694562
8	rs2256087	11496193	С	А	1380	0.5757	-4.552	5.32E-06	8	rs2256087	11496193	1	С	А	1.5806599	0.9401	0.116553
12	rs491825	113048587	Т	С	1344	1.522	4.487	7.21E-06	12	rs17824050	113036817	0.553	А	G	0.8728475	0.4514	0.6146593
12	rs471231	118187337	А	G	1379	0.5948	-4.77	1.84E-06	12	rs471231	118187337	1	А	G	1.2764438	1.6962	0.3977656
17	rs75997523	6730523	А	G	1350	2.895	4.756	1.97E-06	17	rs12601392	6699944	0.537	G	А	1.2769892	0.4180	0.6042291
17	rs11656431	32392629	А	G	1327	5.12	4.576	4.74E-06	17	rs11658185	32394763	0.158	G	А	0.8852353	0.1711	0.7424022
18	rs9952569	24067227	С	Т	1374	1.999	5.502	3.76E-08	18	rs9952569	24067227	1	С	Т	0.5970534	-1.0367	0.2872871
20	rs6056644	9467164	С	А	1380	1.816	4.722	2.33E-06	20	rs6056647	9476953	0.976	А	G	1.4052022	0.8625	0.3847928
20	rs6044984	17718017	А	G	1374	0.5503	-4.645	3.40E-06	20	rs13042529	17698510	0.493	А	G	0.8046117	-1.4447	0.5881554
20	rs117452506	50185618	А	G	1336	2.385	4.448	8.68E-06	20	rs8121883	50197080	0.106	G	А	1.4445344	0.8726	0.2055499

328 Heritability analysis and genetic correlation

Heritability of IHI was estimated from the GWAS summary statistics using LD score regression. The h^2 estimate was 0.54 (0.30) and statistically significant using a one-sided test (Z=1.8; P=0.036). We next sought to identify genomics regions or cell type marker genes that show enriched heritability. However, none of the tested genomic regions or gene sets showed statistically significant enrichment after FDR correction (S6 Fig).

Motivated by reported increased prevalence of IHI in persons with epilepsy we computed the genetic correlation (rg) between IHI and epilepsy susceptibility. The estimate of rg was -0.0854 (0.2612) which did not reach statistically significance (Z=-0.3269; P=0.7437).

LDhub was used to compute rg between IHI and 832 GWAS summary statistics; the computation was successful for 749 GWAS (Supporting S1 Table). None of the traits survived the FDR corrected p-value threshold (P_{FDR}<0.05). A total of 20 traits reached nominal significance (p<0.05, Table 3). The top three positively correlated traits were: intelligence [58], College or University degree based on a UK BioBank (UKBB) GWAS and Years of Schooling [59]. Among the 20 nominal significant genetic correlations was also Fluid Intelligence Score (UKBB).

344	Table 3: Genetic correlations between IHI and other GWASs with nominal significance
345	(p<0.05).

Trait	PMID	Category	rg	se	z	р
Intelligence	28530673	cognitive	0.3479	0.139	2.5037	0.0123
Qualifications: College or University degree	0	ukbb	0.3472	0.1392	2.494	0.0126
Years of schooling 2016	27225129	education	0.3227	0.13	2.4828	0.013
HDL cholesterol	20686565	lipids	0.3992	0.1742	2.2923	0.0219
Ulcerative colitis	26192919	autoimmune	0.4354	0.1978	2.2015	0.0277
Started insulin within one year diagnosis of diabetes	0	ukbb	0.7574	0.3488	2.1716	0.0299
Mothers age at death	0	ukbb	0.6082	0.2801	2.1717	0.0299

Medication for cholesterol_ blood pressure or diabetes: Insulin	0	ukbb	1.2001	0.5571	2.154	0.0312
Creatinine	27005778	metabolites	0.7254	0.3513	2.0652	0.0389
Pain type(s) experienced in last month: Back pain	0	ukbb	-0.3682	0.1826	-2.0165	0.0438
Illnesses of father: Chronic bronchitis/emphysema	0	ukbb	-0.4744	0.2357	-2.0132	0.0441
Triglycerides in small VLDL	27005778	metabolites	0.643	0.3196	2.0123	0.0442
Age at Menopause	26414677	reproductive	0.4687	0.2331	2.0103	0.0444
Fluid intelligence score	0	ukbb	0.2584	0.1286	2.0094	0.0445
Smoking/smokers in household	0	ukbb	-0.5759	0.2881	-1.9987	0.0456
Tinnitus: Yes_ now some of the time	0	ukbb	-1.3776	0.6904	-1.9954	0.046
Concentration of small VLDL particles	27005778	metabolites	0.6411	0.3221	1.9904	0.0465
Time spent using computer	0	ukbb	0.2483	0.1249	1.9883	0.0468
Pack years adult smoking as proportion of life span exposed to smoking PREVIEW ONLY	0	ukbb	-0.3127	0.1587	-1.9702	0.0488
Serum total triglycerides	27005778	metabolites	0.6408	0.3258	1.9671	0.0492

346

347

Gene expression in the developing human brain

We extracted the gene expression levels in the developing brain for *AQP4* and *KCTD1*. The summary of the data up to post-conception week (pcw) 100 are depicted in S7 Fig. Expression of *KCTD1* remains rather stable across the entire time frame, while the expression of *AQP4* starts very low and increases with the progression of brain maturation and reaches its peak around the time of term birth (pcw 37-40). Gene expression was significantly different before and after pcw 25 for both *KCTD1* (p=3.115e-04) and *AQP4* (p=2.424e-07) when limited to data acquired before pcw 40.

357 **Discussion**

358

359 Incidence rate of IHI was consistently around 25% in both, the discovery and the validation cohort. 360 This was comparable to previous reports of 18-19% [60,61], especially considering that the IHI 361 score at the 4.0 cutoff includes not only strong IHI (as in the cited studies), but also lighter IHI 362 [15], therefore increasing the IHI rate. In both cohorts there was a higher incidence rate of IHI in 363 the left hippocampus, which agrees well with the observations that the right hippocampus matures 364 faster and thereby inverts correctly. The GWAS highlighted one genome-wide significant locus on 365 chromosome 18, which is linked through chromatin interaction maps (Fig. S4, left) to six genes, 366 two of which show substantial expression in brain tissue: KCTD1 and AQP4. Of note, the locus 367 showed consistently strong association with continuous scales of the IHI phenotype. Furthermore, 368 a genome-wide screen of those continuous scales revealed two genome-wide significant loci 369 (Fig. S3), one of which exceeded the suggestive threshold in the original GWAS (Table 2; 370 rs35806781, OR=4.889, Z=4.798, P=1.603e-06). The other SNP (rs186025034) had a low minor 371 allele frequency (about 1%) and missed the suggestive threshold in the original GWAS 372 (OR=5.159, Z=4.34, P=1.423e-05). Overall, we observed consistency across IHI definitions and 373 their genetic associations.

374 The Potassium Channel Tetramerization Domain Containing 1 (KCTD1) gene negatively 375 regulates the AP-2 family of transcription factors and the Wnt signalling pathway, which controls 376 normal embryonic development, cellular proliferation and growth [62]. Interestingly, mutations in 377 KCTD1 have been linked to Scalp-Ear-Nipple syndrome [63], which is a rare, autosomal-dominant 378 disorder characterized by cutis aplasia of the scalp as well as minor anomalies of the external 379 ears, digits, nails, and malformations of the breast. Clearly, KCTD1 has the ability to influence 380 developmental processes. Thus, it is conceivable that more benign variation in *KCTD1* may play 381 a role in the generation of IHI. Furthermore, KCTD1 is a potassium channel gene and various

members of the potassium channel gene family have been linked as causes of epilepsy [64–66].
In a recent GWAS for epilepsy susceptibility SNPs in the *KCTD1* gene reached p-values as low
as 0.0003758 (S8 Fig) [56].

385 Aguaporin-4 (AQP4) is a bidirectional water channel that is found on astrocytes throughout 386 the central nervous system (S4 Fig). However, while AQP4 expression in brain tissue is in general 387 high in children and adults, its expression is guite low before post-conception week 20 (S7 Fig). 388 MRI studies of IHI during brain development [20,23] show lack of hippocampal inversion during 389 the early phases of development <25 post-conception weeks, which coincides with the time point 390 of increased AQP4 expression. Furthermore, AQP4 has been linked through various lines of 391 evidence to epilepsy, e.g., the lack of aquaporin-4 water channels increased seizure threshold 392 and seizure duration in mice [67,68] and AQP4 expression among chronic temporal lobe epilepsy 393 patients is increased almost twofold in the hippocampus of the affected hemisphere compared to 394 the contralateral hemisphere [69]. Taken together, astroglial AQP4 may modulate neuronal 395 excitability by regulating the extraneuronal and extrasynaptic environments and thereby affect the 396 epileptogenesis. This may explain the observed increased rates of IHI in persons with epilepsy. 397 Interestingly, the four protein-coding genes prioritized by FUMA were also enriched in genes 398 overexpressed in hippocampal and caudate tissue (S5 Fig). However, enrichment results tend to 399 be unstable when only a small gene set is tested for enrichment, thus, this result should be 400 considered with caution regarding its interpretation.

We attempted to validate the genome-wide significant locus in a second independent cohort of adolescents. However, neither the genome-wide significant locus, nor any of the suggestive loci reached nominal significance in the validation cohort. One major contributor to this lack of replication was the limited sample size of the validation cohort. Despite the equally large set of participants in both studies, age and ethnicity restrictions severely limited the available sample size for the validation cohort (N=161) and drastically lowered the statistical power to detect differences (power of 35% for just the top variant). Larger validation cohorts are needed to confirm

408 the association of the identified locus with IHI: e.g., to validate the top variant with 80% at least 409 N=500 subjects are required. Although there are growing imaging and genetic datasets, e.g., the 410 UKBB that aims at 100,000 participants with genetics and brain imaging data, few studies focus 411 on healthy younger subjects (children, adolescents or young adults), which is beneficial for the 412 validation in order to exclude confounding by disease processes or age-related atrophy. One such 413 option is the Philadelphia Neurodevelopmental Cohort (PNC) [70]. However, it is important to 414 keep in mind that the evaluation of IHI is not restricted to adolescents. IHI can be observed in 415 children and adults too, without extra difficulties. The study here focus on adolescents, since the 416 discovery cohort is the dataset used for the reference study [15]. Still, in older patients, even 417 though IHI still exist, their detection may be more difficult and less reliable due to the confounding 418 effect of hippocampal atrophy to ageing, hippocampal sclerosis or neurodegeneration. Also 419 scoring bigger databases (such as UKBB) will be feasible only after automatic methods for IHI 420 scoring have been developed.

421 We estimated heritability of IHI based on the state-of-the-art LD score regression method 422 that operates on the GWAS summary statistics. The inferred heritability was substantial with 423 h^2 =0.54; the estimate was subject to high uncertainty as reflected by the high standard deviation 424 of 0.3, which is likely a direct reflection of the low sample size of the discovery cohort. Analysis 425 on twin data, such as the healthy young adult twins participating in the Queensland Twin IMaging 426 (QTIM) study [71], can be used to confirm this preliminary heritability estimate, but would require 427 a significant effort in manually scoring IHI in these large cohorts. In addition, the magnitude of h^2 428 is comparable to recently published estimates on the heritability of hippocampal volume and 429 shape from more than 3600 subjects [25]; h^2 ranges from 0.08 to 0.337 depending on hemisphere 430 and structural measure, with heritability of volume being generally the lowest. Given the impact 431 of IHI on hippocampal shape and appearance together with the high prevalence of IHI in nearly 432 25% of healthy subjects, it is likely that the observed heritability of hippocampal shape reported 433 by [25] was in part due to IHI.

434 We used the generated genome-wide summary statistics for two additional explorations. 435 First, we sought to investigate if heritability was enriched in any particular region of the genome, 436 characterized by its function or by marker genes for specific cell types. None of the investigated 437 categories achieved statistical significance after FDR correction. Second, we computed genetic 438 correlations with other traits. We hypothesized that there may be genetic link with epilepsy, 439 however the resulting correlation was non-significant and negative, i.e., people with IHI were less 440 likely to be affected by epilepsy, thereby contradicting earlier reports. The exploratory analysis 441 with 832 additional traits highlighted a positive genetic correlation between IHI and intelligence 442 and education attainment. There are various reports highlighting the contribution of the 443 hippocampus and its subregions to various mental aspects that collectively are referred to as 444 intelligence, e.g., spatial processing [72] and working memory [73]. Moreover, one recent study 445 linked hippocampal shape to cognitive performance [74]. In particular, in males the radial distance 446 of the hippocampus correlated with better test scores (e.g., general factor of intelligence, abstract-447 fluid intelligence, and the rotation of solid figures). In females, the effect was reversed. Therefore, 448 a genetic correlation of IHI with intelligence in the broad sense is conceivable.

In conclusion, we presented the first genome-wide association study of IHI, where we identified a genome-wide significant locus. Additional exploration of the resulting summary statistics revealed a high heritability and suggested positive genetic correlation of IHI with traits linked to intelligence and education attainment.

453 Acknowledgements

MAS acknowledges financial support by the EPSRC-funded UCL Centre for Doctoral Training in
Medical Imaging (EP/L016478/1). AA holds an MRC eMedLab Medical Bioinformatics Career
Development Fellowship. This work was supported by the Medical Research Council [grant
number MR/L016311/1]. The research leading to these results has received funding

from the program "Investissements d'avenir" ANR-10-IAIHU-06 (Agence Nationale de la
Recherche-10-IA Institut Hospitalo-Universitaire-6) and from the "Contrat d'Interface Local"
program (to OC) from Assistance Publique-Hôpitaux de Paris (AP-HP).

461

Data and/or research tools used in the preparation of this manuscript were obtained and analyzed from the controlled access datasets distributed from the NIMH-supported research Domain Criteria Database (RDoCdb). RDoCdb is a collaborative informatics system created by the National Institute of Mental Health to store and share data resulting from grants funded through the Research Domain Criteria (RDoC) project. Dataset identifier(s): #2607.

467 Data collection and sharing for this project was funded by the Pediatric Imaging, Neurocognition
468 and Genetics Study (PING) (National Institutes of Health Grant RC2DA029475).

469

470 IMAGEN was supported by the European Union-funded FP6 (LSHM-CT-2007-037286).

471 IMAGEN Consortium author list:

472 Contact: Gunter Schumann (gunter.schumann@kcl.ac.uk)

Tobias Banaschewski M.D., Ph.D.¹⁸; Gareth J. Barker* Ph.D.¹⁹; Arun L.W. Bokde Ph.D.²⁰; Uli
Bromberg Ph.D.²¹; Christian Büchel M.D.²¹; Erin Burke Quinlan, PhD¹⁶; Sylvane Desrivières
Ph.D.¹⁶; Herta Flor Ph.D.^{22,23}; Hugh Garavan Ph.D.²⁴; Penny Gowland Ph.D.²⁵; Bernd Ittermann
Ph.D.²⁶; Marie-Laure Paillère Martinot M.D., Ph.D.²⁷; Eric Artiges M.D., Ph.D.²⁸; Frauke Nees
Ph.D.^{18,22}; Dimitri Papadopoulos Orfanos Ph.D.⁹; Tomáš Paus M.D., Ph.D.²⁹; Luise Poustka
M.D.³⁰; Sarah Hohmann M.D.¹⁸; Sabina Millenet Dipl.-Psych.¹⁸; Juliane H. Fröhner Dipl.-Psych.¹⁵;

479 Robert Whelan Ph.D.³¹

¹⁸Department of Child and Adolescent Psychiatry and Psychotherapy, Central Institute of Mental
 Health, Medical Faculty Mannheim, Heidelberg University, Square J5, 68159 Mannheim,
 Germany; ¹⁹Department of Neuroimaging, Institute of Psychiatry, Psychology & Neuroscience,

King's College London, United Kingdom; ²⁰Discipline of Psychiatry, School of Medicine and Trinity 483 484 College Institute of Neuroscience, Trinity College Dublin; ²¹University Medical Centre Hamburg-485 Eppendorf, House W34, 3.OG, Martinistr. 52, 20246, Hamburg, Germany; ²²Department of 486 Cognitive and Clinical Neuroscience, Central Institute of Mental Health, Medical Faculty 487 Mannheim, Heidelberg University, Square J5, Mannheim, Germany; ²³ Department of Psvchology, School of Social Sciences, University of Mannheim, 68131 Mannheim, Germany; 488 489 ²⁴Departments of Psychiatry and Psychology, University of Vermont, 05405 Burlington, Vermont, USA: ²⁵Sir Peter Mansfield Imaging Centre School of Physics and Astronomy, University of 490 491 Nottingham, University Park, Nottingham, United Kingdom; ²⁶Physikalisch-Technische 492 Bundesanstalt (PTB), Braunschweig and Berlin, Germany [or depending on journal requirements 493 can be: Physikalisch-Technische Bundesanstalt (PTB), Abbestr. 2 – 12, Berlin, Germany; 494 ²⁷Institut National de la Santé et de la Recherche Médicale, INSERM Unit 1000 "Neuroimaging & 495 Psychiatry", University Paris Sud, University Paris Descartes; Sorbonne Université; and AP-HP, Department of Child and Adolescent Psychiatry, Pitié-Salpêtrière Hospital, Paris, France; ²⁸Institut 496 497 National de la Santé et de la Recherche Médicale, INSERM Unit 1000 "Neuroimaging & 498 Psychiatry", University Paris Sud, University Paris Descartes - Sorbonne Paris Cité; and 499 Psychiatry Department 91G16, Orsay Hospital, France: ²⁹Bloorview Research Institute, Holland 500 Bloorview Kids Rehabilitation Hospital and Departments of Psychology and Psychiatry, University 501 of Toronto, Toronto, Ontario, M6A 2E1, Canada; ³⁰Department of Child and Adolescent 502 Psychiatry and Psychotherapy, University Medical Centre Göttingen, von-Siebold-Str. 5, 37075, 503 Göttingen, Germany; ³¹School of Psychology and Global Brain Health Institute, Trinity College 504 Dublin, Ireland;

505

506 Author contributions

507 CC and AA are guarantor of integrity of the entire study. All authors contributed to manuscript 508 drafting or manuscript revision for important intellectual content. All authors gave their approval 509 of the final version of submitted manuscript. CC, MAS, RT, AA and OC did literature research, 510 designed the study, performed experiments and analysed data. CC, MAS, AA and OC wrote the 511 article. VF contributed to genotyping data, JBP contributed to the statistical processing of data. 512 EA, AG, AH, HW, HL, JLM, MS and GS contributed to the clinical study and data acquisition. 513

514 Ethics committee:

515 London: Psychiatry, Nursing and Midwifery (PNM) Research Ethics Subcommittee (RESC), 516 Waterloo Campus, King's College London. Nottingham: University of Nottingham Medical School 517 Ethics Committee. Mannheim: Medizinische Fakultaet Mannheim, Ruprecht Karl Universitaet 518 Heidelberg and Ethik-Kommission II an der Fakultaet fuer Kliniksche Medizin Mannheim. 519 Dresden: Ethikkommission der Medizinischen Fakultaet Carl Gustav Carus, TU Dresden 520 Medizinische Fakultaet. Hamburg: Ethics board, Hamburg Chamber of Phsyicians. Paris: CPP 521 IDF VII (Comité de protection des personnes IIe de France), ID RCB: 2007-A00778-45 September 522 24th 2007. Dublin: TCD School of Psychology REC. Berlin: ethics committee of the Faculty of 523 Psychology. And Mannheim's ethics committee approved the whole study. All informed consents 524 have been obtained from a parent and/or legal guardian. Written informed assent and consent 525 were obtained, respectively, from all adolescents and their parents after complete description of 526 the study. For the PING dataset, written parental informed consent was obtained for all PING 527 subjects below the age of 18, and child assent was also obtained for all participants between the 528 ages of 7 and 17. Written informed consent was obtained directly from all participants aged 18 529 years or older.

530

531 **References**

- 532
- Burgess N, Maguire EA, O'Keefe J. The Human Hippocampus and Spatial and Episodic
 Memory. Neuron. 2002 Aug;35[4]:625–41.
- Jankord R, Herman JP. Limbic Regulation of Hypothalamo-Pituitary-Adrenocortical Function during Acute and Chronic Stress. Annals of the New York Academy of Sciences. 2008 Dec;1148[1]:64–73.
- Barnes J, Bartlett JW, van de Pol LA, Loy CT, Scahill RI, Frost C, et al. A meta-analysis of hippocampal atrophy rates in Alzheimer's disease. Neurobiology of Aging. 2009 Nov;30[11]:1711–23.
- Colle R, Cury C, Chupin M, Deflesselle E, Hardy P, Nasser G, et al. Hippocampal volume predicts antidepressant efficacy in depressed patients without incomplete hippocampal inversion. NeuroImage: Clinical. 2016 Feb 1;12:949–55.
- 544 5. Sämann PG, Höhn D, Chechko N, Kloiber S, Lucae S, Ising M, et al. Prediction of
 antidepressant treatment response from gray matter volume across diagnostic categories.
 European Neuropsychopharmacology. 2013 Nov;23[11]:1503–15.

- Colle R, Chupin M, Cury C, Vandendrie C, Gressier F, Hardy P, et al. Depressed suicide attempters have smaller hippocampus than depressed patients without suicide attempts. Journal of Psychiatric Research. 2015 Feb;61:13–8.
- Travis SG, Coupland NJ, Hegadoren K, Silverstone PH, Huang Y, Carter R, et al. Effects of cortisol on hippocampal subfields volumes and memory performance in healthy control subjects and patients with major depressive disorder. Journal of Affective Disorders. 2016 Sep;201:34–41.
- van Erp TGM, Hibar DP, Rasmussen JM, Glahn DC, Pearlson GD, Andreassen OA, et al.
 Subcortical brain volume abnormalities in 2028 individuals with schizophrenia and 2540 healthy controls via the ENIGMA consortium. Mol Psychiatry. 2016 Apr;21[4]:547–53.
- 557 9. Helmstaedter C, Elger CE. Chronic temporal lobe epilepsy: a neurodevelopmental or 558 progressively dementing disease? Brain. 2009 Oct 1;132[10]:2822–30.
- 10. Qiu A, Rifkin-Graboi A, Chen H, Chong Y-S, Kwek K, Gluckman PD, et al. Maternal anxiety
 and infants' hippocampal development: timing matters. Transl Psychiatry. 2013
 Sep;3[9]:e306–e306.
- 562 11. Bronen RA, Cheung G. MRI of the normal hippocampus. Magnetic Resonance Imaging. 563 1991;9[4]:497–500.
- Lehéricy S, Dormont D, Sémah F, Clémenceau S, Granat O, Marsault C, et al. Developmental
 abnormalities of the medial temporal lobe in patients with temporal lobe epilepsy. AJNR
 American journal of neuroradiology. 1995;16[4]:617–26.
- 567 13. Baulac M, De Grissac N, Hasboun D, Oppenheim C, Adam C, Arzimanoglou A, et al.
 568 Hippocampal developmental changes in patients with partial epilepsy: magnetic resonance
 569 imaging and clinical aspects. Annals of neurology. 1998;44:223–33.
- 570 14. Bernasconi N, Kinay D, Andermann F, Antel S, Bernasconi A. Analysis of shape and
 571 positioning of the hippocampal formation: an MRI study in patients with partial epilepsy and
 572 healthy controls. Brain : a journal of neurology. 2005;128:2442–2452.
- 573 15. Cury C, Toro R, Cohen F, Fischer C, Mhaya A, Samper-González J, et al. Incomplete
 574 Hippocampal Inversion: A Comprehensive MRI Study of Over 2000 Subjects. Frontiers in
 575 neuroanatomy. 2015;9:160.
- 576 16. Cury C. Statistical shape analysis of the anatomical variability of the human hippocampus in
 577 large populations. Sorbonne Université; 2015.
- 578 17. Colenutt J, McCann B, Knight MJ, Coulthard E, Kauppinen RA. Incomplete Hippocampal
 579 Inversion and Its Relationship to Hippocampal Subfield Volumes and Aging: Incomplete
 580 Hippocampal Inversion and Aging. Journal of Neuroimaging. 2018 Jul;28[4]:422–8.
- 18. Kim H, Chupin M, Colliot O, Bernhardt BC, Bernasconi N, Bernasconi A. Automatic
 hippocampal segmentation in temporal lobe epilepsy: Impact of developmental
 abnormalities. NeuroImage. 2012 Feb 15;59[4]:3178–86.

- 19. Okada Y, Kato T, Iwai K, Iwasaki N, Ohto T, Matsui A. Evaluation of hippocampal infolding using magnetic resonance imaging. NeuroReport. 2003;14[10]:1405–9.
- 586 20. Bajic D, Ewald U, Raininko R. Hippocampal development at gestation weeks 23 to 36. An 587 ultrasound study on preterm neonates. Neuroradiology. 2010;52[6]:489–94.
- 588 21. Baker LL, Barkovich AJ. The large temporal horn: MR analysis in developmental brain 589 anomalies versus hydrocephalus. American Journal of Neuroradiology. 1992;13[1]:115–22.
- Sighini A, Zirpoli S, Parazzini C, Bianchini E, Scifo P, Sala C, et al. Hippocampal infolding
 angle changes during brain development assessed by prenatal MR imaging. American
 Journal of Neuroradiology. 2006;27[10]:2093–7.
- 593 23. Bajic D, Canto Moreira N, Wikström J, Raininko R. Asymmetric development of the
 594 hippocampal region is common: A fetal MR imaging study. American Journal of
 595 Neuroradiology. 2012;33[3]:513–8.
- 596 24. Andrade D, Krings T, Chow EWC, Kiehl TR, Bassett AS. Hippocampal malrotation is
 597 associated with chromosome 22q11.2 microdeletion. Canadian Journal of Neurological
 598 Sciences. 2013;40[5]:652–6.
- 599 25. Roshchupkin G V., Gutman BA, Vernooij MW, Jahanshad N, Martin NG, Hofman A, et al.
 600 Heritability of the shape of subcortical brain structures in the general population. Nature
 601 Communications. 2016;7:13738.
- 602 26. Schumann G, Loth E, Banaschewski T, Barbot A, Barker G, Büchel C, et al. The IMAGEN
 603 study: reinforcement-related behaviour in normal brain function and psychopathology.
 604 Molecular psychiatry. 2010;15[12]:1128–39.
- 27. Jernigan TL, Brown TT, Hagler DJ, Akshoomoff N, Bartsch H, Newman E, et al. The Pediatric
 Imaging, Neurocognition, and Genetics (PING) Data Repository. NeuroImage. 2016;124[Pt
 B]:1149–54.
- 28. Jenkinson M, Smith S. A global optimisation method for robust affine registration of brain
 images. Medical Image Analysis. 2001;5[2]:143–56.
- 610 29. Jenkinson M, Bannister P, Brady M, Smith S. Improved optimization for the robust and
 611 accurate linear registration and motion correction of brain images. NeuroImage.
 612 2002;17[2]:825–41.
- 30. The International HapMap 3 Consortium, Altshuler DM, Gibbs RA, Peltonen L, Dermitzakis E,
 Schaffner SF, et al. Integrating common and rare genetic variation in diverse human
 populations. Nature. 2010;467:52–58.
- 616 31. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait 617 analysis. American journal of human genetics. 2011;88[1]:76–82.
- McCarthy S, Das S, Kretzschmar W, Delaneau O, Wood AR, Teumer A, et al. A reference
 panel of 64,976 haplotypes for genotype imputation. Nature Genetics. 2016;48:1279–1283.

- 33. Loh PR, Danecek P, Palamara PF, Fuchsberger C, Reshef YA, Finucane HK, et al.
 Reference-based phasing using the Haplotype Reference Consortium panel. Nature
 Genetics. 2016;48[11]:1443–8.
- 623 34. Durbin R. Efficient haplotype matching and storage using the positional Burrows-Wheeler 624 transform (PBWT). Bioinformatics. 2014;30[9]:1266–72.
- 35. Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated
 individuals. Genome Research. 2009;19[9]:1655–1664.
- 627 36. Chang CC, Chow CC, Tellier LCAM, Vattikuti S, Purcell SM, Lee JJ. Second-generation 628 PLINK: Rising to the challenge of larger and richer datasets. GigaScience. 2015;4[1].
- 37. Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, et al. LocusZoom:
 regional visualization of genome-wide association scan results. Bioinformatics.
 2010;26[18]:2336–7.
- 38. Machiela MJ, Chanock SJ. LDlink: A web-based application for exploring population-specific
 haplotype structure and linking correlated alleles of possible functional variants.
 Bioinformatics. 2015;31[21]:3555–7.
- 39. Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and annotation
 of genetic associations with FUMA. Nature communications. 2017;8[1]:1826.
- 637 40. GTEx Consortium. Human genomics. The Genotype-Tissue Expression (GTEx) pilot
 638 analysis: multitissue gene regulation in humans. Science (New York, NY).
 639 2015;348[6235]:648–60.
- 41. Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, Kettunen J, et al. Systematic
 identification of trans eQTLs as putative drivers of known disease associations. Nature
 Genetics. 2013;45:1238–1243.
- 42. Zhernakova D V., Deelen P, Vermaat M, Van Iterson M, Van Galen M, Arindrarto W, et al.
 Identification of context-dependent expression quantitative trait loci in whole blood. Nature
 Genetics. 2017;49:139–145.
- 43. Ramasamy A, Trabzuni D, Guelfi S, Varghese V, Smith C, Walker R, et al. Genetic variability
 in the regulation of gene expression in ten regions of the human brain. Nature neuroscience.
 2014;17[10]:1418–28.
- 44. Schmitt AD, Hu M, Jung I, Xu Z, Qiu Y, Tan CL, et al. A Compendium of Chromatin Contact
 Maps Reveals Spatially Active Regions in the Human Genome. Cell Reports. 2016;17:2042–
 2059.
- 45. Roadmap Epigenomics Consortium, Kundaje A, Meuleman W, Ernst J, Bilenky M, Yen A, et al. Integrative analysis of 111 reference human epigenomes. Nature. 2015;518:317–330.
- 46. Liberzon A, Subramanian A, Pinchback R, Thorvaldsdóttir H, Tamayo P, Mesirov JP.
 Molecular signatures database (MSigDB) 3.0. Bioinformatics. 2011;27:1739–1740.

- 47. Kutmon M, Riutta A, Nunes N, Hanspers K, Willighagen EL, Bohler A, et al. WikiPathways:
 Capturing the full diversity of pathway knowledge. Nucleic Acids Research. 2016;44:D488–
 D494.
- 48. Welter D, MacArthur J, Morales J, Burdett T, Hall P, Junkins H, et al. The NHGRI GWAS
 Catalog, a curated resource of SNP-trait associations. Nucleic Acids Research.
 2014;42:D1001–D1006.
- 49. Bulik-Sullivan B, Loh P, Finucane H, Ripke S. LD Score regression distinguishes confounding
 from polygenicity in genome-wide association studies. Nature. 2015;
- 50. Finucane HK, Bulik-Sullivan B, Gusev A, Trynka G, Reshef Y, Loh P-R, et al. Partitioning
 heritability by functional annotation using genome-wide association summary statistics.
 Nature genetics. 2015;47[11]:1228–35.
- 51. Finucane HK, Reshef YA, Anttila V, Slowikowski K, Gusev A, Byrnes A, et al. Heritability
 enrichment of specifically expressed genes identifies disease-relevant tissues and cell
 types. Nature genetics. 2018;50[4]:621–9.
- 52. Cahoy JD, Emery B, Kaushal A, Foo LC, Zamanian JL, Christopherson KS, et al. A
 transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for
 understanding brain development and function. The Journal of neuroscience : the official
 journal of the Society for Neuroscience. 2008;28[1]:264–78.
- 53. Pers TH, Karjalainen JM, Chan Y, Westra H-J, Wood AR, Yang J, et al. Biological
 interpretation of genome-wide association studies using predicted gene functions. Nature
 communications. 2015;6:5890.
- 54. Heng TSP, Painter MW, Consortium IGP. The Immunological Genome Project: networks of
 gene expression in immune cells. Nature immunology. 2008;9[10]:1091–4.
- 55. Bulik-Sullivan B, Finucane HK, Anttila V, Gusev A, Day FR, Loh P-R, et al. An atlas of genetic
 correlations across human diseases and traits. Nature genetics. 2015;47[11]:1236–41.
- 56. The International League Against Epilepsy Consortium on Complex Epilepsies. Genetic
 determinants of common epilepsies: a meta-analysis of genome-wide association studies.
 Lancet neurology. 2014;13[9]:893–903.
- 57. Zheng J, Erzurumluoglu AM, Elsworth BL, Kemp JP, Howe L, Haycock PC, et al. LD Hub: a
 centralized database and web interface to perform LD score regression that maximizes the
 potential of summary level GWAS data for SNP heritability and genetic correlation analysis.
 Bioinformatics. 2017;33[2]:272–9.
- 58. Sniekers S, Stringer S, Watanabe K, Jansen PR, Coleman JRI, Krapohl E, et al. Genomewide association meta-analysis of 78,308 individuals identifies new loci and genes
 influencing human intelligence. Nature Genetics. 2017;49[7]:1107–12.
- 691 59. Okbay A, Beauchamp JP, Fontana MA, Lee JJ, Pers TH, Rietveld CA, et al. Genome-wide
 692 association study identifies 74 loci associated with educational attainment. Nature.
 693 2016;533[7604]:539–42.

- 60. Bajic D, Wang C, Kumlien E, Mattsson P, Lundberg S, Eeg-Olofsson O, et al. Incomplete
 inversion of the hippocampus A common developmental anomaly. European Radiology.
 2008;18[1]:138–42.
- 697 61. Bajic D, Kumlien E, Mattsson P, Lundberg S, Wang C, Raininko R. Incomplete hippocampal 698 inversion - Is there a relation to epilepsy? European Radiology. 2009;19[10]:2544–50.
- 699 62. Li X, Chen C, Wang F, Huang W, Liang Z, Xiao Y, et al. KCTD1 Suppresses Canonical Wnt
 700 Signaling Pathway by Enhancing β-catenin Degradation. PLoS ONE. 2014;9[4]:e94343.
- 63. Marneros AG, Beck AE, Turner EH, McMillin MJ, Edwards MJ, Field M, et al. Mutations in
 KCTD1 cause scalp-ear-nipple syndrome. American Journal of Human Genetics.
 2013;92[4]:621–6.
- 64. Singh NA, Charlier C, Stauffer D, DuPont BR, Leach RJ, Melis R, et al. A novel potassium
 channel gene, KCNQ2, is mutated in an inherited epilepsy of newborns. Nature Genetics.
 1998;18:25–29.
- For the second se
- 66. Liu Z, Xiang Y, Sun G. The KCTD family of proteins: structure, function, disease relevance.
 Cell & bioscience. 2013;3[45].
- 67. Binder DK, Yao X, Verkman AS, Manley GT. Increased seizure duration in mice lacking
 aquaporin-4 water channels. Acta Neurochirurgica, Supplementum. 2006;96.
- 68. Binder DK, Oshio K, Ma T, Verkman AS, Manley GT. Increased seizure threshold in mice
 lacking aquaporin-4 water channels. NeuroReport. 2004;15[2]:259–62.
- 69. Das A, Wallace GC, Holmes C, McDowell ML, Smith JA, Marshall JD, et al. Hippocampal tissue of patients with refractory temporal lobe epilepsy is associated with astrocyte activation, inflammation, and altered expression of channels and receptors. Neuroscience.
 2012;220:237–46.
- 719 70. Satterthwaite TD, Elliott MA, Ruparel K, Loughead J, Prabhakaran K, Calkins ME, et al.
 720 Neuroimaging of the Philadelphia neurodevelopmental cohort. NeuroImage. 2014;86:544–
 721 53.
- 722 71. de Zubicaray GI, Chiang M-C, McMahon KL, Shattuck DW, Toga AW, Martin NG, et al.
 723 Meeting the Challenges of Neuroimaging Genetics. 2008;2[4]:258–63.
- 724 72. Maguire EA, Gadian DG, Johnsrude IS, Good CD, Ashburner J, Frackowiak RSJ, et al.
 725 Navigation-related structural change in the hippocampi of taxi drivers. Proceedings of the
 726 National Academy of Sciences. 2000;97[8]:4398–403.
- 727 73. Axmacher N, Henseler MM, Jensen O, Weinreich I, Elger CE, Fell J. Cross-frequency
 728 coupling supports multi-item working memory in the human hippocampus. Proceedings of
 729 the National Academy of Sciences. 2010;107[7]:3228–33.

730 74. Colom R, Stein JL, Rajagopalan P, Martínez K, Hermel D, Wang Y, et al. Hippocampal 731 structure and human cognition: Key role of spatial processing and evidence supporting the 732 efficiency hypothesis in females. Intelligence. 2013;41[2]:129–140.

733

734 Supporting information

S1 Fig. Ancestry matching of the IMAGEN cohort. The panels show pairwise scatter plots for
the first five principal components of the genetic relationship matrix. The HapMap3 subjects are
represented as filled circles and color coded by population (see legend in the top left panel).
IMAGEN participants are represented by black open circles; red open circles indicate subjects
that were excluded based on distance to the CEU+TSI European ancestry.

- 740
- 741 S2 Fig. Quantile-quantile plot. QQ plot for the genome-wide association study. The xaxis
- shows the expected -log10(p-value) while the y-axis shows the study -log10(p-value).
- 743 There was no evidence of p-value inflation λ =1.017.
- 744

745 S3 Fig. Manhattan plots for continuous IHI scores. The y-axis depicts the -log10(p-value) of 746 the association between SNP and presence of IHI assuming an additive model in the discovery 747 cohort. The SNPs tested in the study are ordered along their chromosomal position on the x-axis. 748 The red horizontal line donates genome wide significance at the Bonferroni threshold (P=5e-8), 749 while the blue horizontal line marks the threshold for suggestive association (P=1e-5). Upper plot: 750 Result for the sum of the five criteria and then maxed over left and right hippocampus. Two loci 751 exceed the genome wide significant threshold: on chromosome 6 rs35806781 (beta=1.478, 752 Z=5.766, P=1.006e-08) and on chromosome 9 rs186025034 (beta=1.867, Z=6.202, P=7.408e-753 10). Lower plot: Result for the global criterion presenting 3 classes: 0 = non IHI, 2 = IHI, 1 = partial 754 IHI, and taking the max over left and right hippocampus. One locus exceeds genome-wide 755 significance on chromosome 9: rs186025034 (beta=0.8251, Z=5.575, P=2.98e-08).

756 757

S4 Fig. FUMA results. Left: chromatin interaction plot, mapping the genome-wide significant
locus to six genes on chromosome 18. Right: expression heatmap (average log2(RPKM) in 53
GTEx tissues) for the four mapped protein-coding genes.

761

S5 Fig. Enrichment analysis of prioritized protein-coding genes in GTEx. The four prioritized
 genes are tested for enrichment in tissue-specific gene lists for 53 tissue from the GTEx dataset.
 Each list of differentially expressed genes is split into overexpressed and underexpressed genes.
 P-value threshold for significance was the Bonferroni corrected threshold for 3 * 53 tests
 (p<0.000314). Tissues showing significant enrichment are indicated by red bars.

767

S6 Fig. Heritability partitioning analysis: panels 1-5, among cell/tissue-specific differentially
 expressed genes; panel 6, among genomic functional annotation categories.

- S7 Fig. Gene expression in the developing human brain for AQP4 and KCTD1. The x-axis
 shows the subjects' age in weeks post conception (pcw). The y-axis depicts the mean of the log2
 RPKM (reads per kilobase per million) provided by BrainSpan. AQP4 and KCTD1 are indicated
 by different colors.
- **S8 Fig. Local association plot for KCTD1 in ILAE epilepsy GWAS.**
- 778 S1 Table. Genetic correlation between IHI and 832 GWASs obtained from LD hub.
- **S1 File. Imagen Consortium author list.**