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A 3D REACTION-DIFFUSION SYSTEM DESCRIBING CALCIUM DYNAMICS IN CARDIAC CELL*

Mostafa Bendahmane¹, Elmahdi Erraji² and Fahd Karami³

Abstract. We are interested in modeling the interaction of calcium dynamics in a medium including sarcolemma and sarcoplasmic reticulum. The governing equations consist of a nonlinear reaction-diffusion system representing the various calcium fluxes and theirs buffers in the two media. We address the question of existence of weak solutions by using a fixed-point approach. We propose a finite element method for this system, we establish the existence of the discrete solution, and we show that the discrete solution generated by the given scheme converges to the corresponding weak solution for the model studied. Finally, we give some 2D and 3D numerical examples to our model.

Mathematics Subject Classification. 92C50, 65M60, 65N12, 92C40.

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Introduction

The study of calcium dynamics in cardiac cell is a priority to reveal the mysterious causes of cell failure. One of the principal causes of heart failure is the perturbation of calcium induced-calcium release (CICR) cycle of a cardiac cell (see for e.g. [20]). The calcium cycle in cardiac cell is a complicated process, since it involves different mechanisms such as swapping between the cytoplasm and sarcoplasmic reticulum through different type of channels and interactions with biochemical species in the cell medium. On the cellular scale, the perturbation of CICR could be explained by pathological behavior of ionic channels especially Ryanodine receptor (RyR). The scientific community is convinced, based on strong evidence (we refer to [33]) that a vast majority of pathological heart cases is due to RyR irregularity. Which induces more questions than answers about understanding the causes and designing an effective therapeutic strategy on the cellular scale. This would require a larger collaboration between various scientific fields including physics, chemistry, biology and mathematics.

In cardiac and other muscle cells, calcium signaling is the main factor to mediate excitation-contraction coupling (ECC). Its cycle lies between the action potential and muscle contraction. The ECC is a consequence of a propagating action potential through muscle cell membranes. At the cellular scale, action potential propagates along the cell membrane including the T-tubules which forms a tubular cavity inside the cell. This action potential triggered voltage opened a channel (VOC) or L-type channels located on the cell membrane surface. As a result, a passive flux of Ca^{2+} crosses the L-type due to the large difference of calcium concentration between the extracellular ($[Ca^{2+}]_e \approx 1mM$) and the intracellular ($[Ca^{2+}]_i \approx 0.1\mu M$) medium. The rising Ca^{2+} concentration triggered the calcium-induced calcium release process (CICR). It begins by triggering a ryanodine receptor located on sarcoplasmic reticulum, this allows the Ca^{2+} influx into intracellular medium from sarcoplasmic reticulum, where Ca^{2+} is stored at elevated concentration ($[Ca^{2+}]_{sr} \approx 1.3mM$, see [2] for this detail), not to mention the calcium concentration maintained by the heavy buffering. This leads to the simultaneous opening of the other ionic channel. Then, the influx of Ca^{2+} ion diffuses through myofibrils to cause mechanical activity.

In literature, many authors attempt to build a primal model of calcium dynamics. We mention, for example the works of Peskoff et al. in [17,25], their first work establishes a primal model of calcium distribution evolution in diad neighbor. A more complete calcium model is established by extending the previous model to the whole cell and considering interaction with local buffers in the medium (see [26] for more details). In passing, we want to mention the work of Sobie et al. in [31], where the authors studied the dynamics of Ca^{2+} under new consideration on RyR Stochastic behavior, period of release and there distribution over space. Note that these studies of spatial calcium dynamics doesn't take in consideration the complexity of cell medium (for example the presence of mitochondria in the cell and the heterogenerous of the media). This will affect the calcium diffusion and produce some serious needs in the modeling part. Recently, some new spatial calcium models are based on a transmission defined by interaction in a local position of ionic channels (see for e.g. [5, 17]). These models can be interpreted as a transmission boundary value problem and can be studied by using domain decomposition techniques. From the analytical and the numerical perspectives, there are many results that has been established in this context (see for e.g. [9,16,21]).

Note that the previous spatial calcium models are governed by a linear diffusion. We know that the solution of a linear diffusion equation admits an infinite speed of propagation whereas the solution of the nonlinear diffusion equation shows a finite speed of propagation. Moreover, from the application point of view it is not realistic to have the property of infinite speed of propagation because the calcium can not diffuse infinitely. In this paper,

we establish the mathematical model of species dynamics in cardio-myocyte with nonlinear diffusion. Note that strong experimental evidence proves the complexity of calcium diffusion (see for e.g. [8]). Theoretically, the ionic nature of Ca^{2+} as well as the complexity of the cytosolic and sacroplasmic medium imposes some modifications on the previous models (we refer here to the works [27,28]). Moreover, advanced microscopical imaging technique shows that the cytoplasm of mammalian cardiac myocytes seems to be highly heterogeneous and anisotropic due to the dense presence of some organelles. We mention, for example the presence of mitochondria and Golgi complex on the micron-scale. A zoom into a smaller scale, clarify the presence of a network of elements such as microtubule and actin filaments. Hence, based on strong experimental evidence shown in [10], the authors highly recommend to consider an anomalous diffusion in the cell medium. Based on the same arguments between others [29], a recent work [12] has considered a porous medium model describing cardiac-electro-physiology.

Our main contribution in this context is to generalize the previous models to a nonlinear model with non constant diffusion coefficients. This paper is devoted to a rigorous study of existence and uniqueness of L^{∞} bounded solutions of the proposed model. Moreover, we establish a fully discrete scheme in finite element framework for spatial discretization, whereas the first order backward Euler scheme is used for time discretization. Then, we move forward to prove the convergence result of the proposed scheme. We want to mention here (in the context of finite element method) that we cannot obtain the maximum principle as in the continuous case because the positive and negative cuts test functions may not be admissible. For the convergence of the scheme, we will assume that the initial conditions are only bounded in L^2 .

The plan of this paper is as follows: In Section 1 we propose the derivation of the calcium model based on conservation law and interaction between species. Section 2 is devoted to the existence and uniqueness of weak solution (defined in Definition 2.1 below) by using the Schauder fixed point theorem and the passage to the limit that transforms the regularized system into the original problem. We introduce in Section 3 some notations for the finite element method and we present our scheme. The proof of the convergence result (of the scheme) is divided into a priori estimates, compactness for discrete solution and convergence to a weak solution. Finally, we give in Section 4 some numerical examples to our model.

1. CALCIUM TRANSMISSION MODEL

We devote this section to a brief derivation of the calcium model of cardiac tissue. As principal references for this model with constant diffusion we found [5]. Before starting our calcium dynamics modeling, we refer to Figure 1 for a good description of the cytosolic and sarcoplasmic reticulum domains.

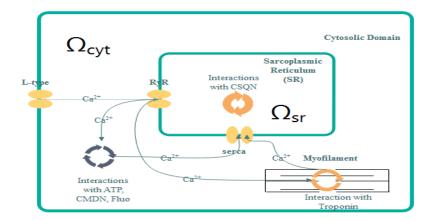


Figure 1. Description of calcium cycle in Cardiac cell

1.1. Description of calcium transmission dynamics

In both sub-domains, calcium behaves in a similar manner. To derive the partial differential equation controlling calcium behavior, we consider an arbitrary domain Ω . By the conservation law, we have

$$\frac{d}{dt} \int_{\Omega} c \, dV = \int_{\Omega} R \, dV - \int_{\partial \Omega} J \cdot \eta \, dA,\tag{1}$$

where c is the calcium concentration, J its flux trough $\partial\Omega$, R its source inside Ω due to interaction of Ca^{2+} with other species and η is the normal vector on $\partial\Omega$.

Now to conclude this equation, we use Stokes Ostradski formula

$$\int_{\partial\Omega} J \cdot \eta \, dA = \int_{\Omega} \operatorname{div} J \, dV. \tag{2}$$

Combining (1) and (2), we arrive

$$\frac{d}{dt} \int_{\Omega} c \, dV = \int_{\Omega} R \, dV - \int_{\Omega} \operatorname{div} J \cdot \eta \, dV. \tag{3}$$

This implies

$$\partial_t c = R - \operatorname{div} J \text{ in } (0, T) \times \Omega.$$
 (4)

Thanks to Fick law, the calcium moves from high concentration to low concentration in cell medium. So the flux J is directed in the opposite direction as the concentration gradient. Moreover, the diffusion coefficient depend on solvent concentration. In summary, calcium flux can be written as

$$J = -D_c(c)\nabla c,\tag{5}$$

where D_c is a diffusion-dependent function of c. Hence, we deduce :

$$\partial_t c = \operatorname{div} \left(D_c(c) \nabla c \right) + R_c \text{ in } (0, T) \times \Omega, \tag{6}$$

and similarly the equation of Ca^{2+} in SR is:

$$\partial_t s = \operatorname{div} \left(D_s(s) \nabla s \right) + R_s \text{ in } (0, T) \times \Omega,$$
 (7)

where D_s is a diffusion-dependent function of s. We will define the sources R_c and R_s explicitly in the following sub-sections.

1.2. Description of buffers dynamics

In this subsection, we establish the equations governing by the buffers (recall that the buffers are proteins that interact with calcium to produce another molecule). We assume that the total concentration of the free buffer [buffer] and the new buffer [buffer- Ca^{2+}] (resulting from the interaction of the free buffer with Ca^{2+}) is a constant in time. The chemical equation describing this phenomenon is:

Buffer
$$+ Ca^{2+} \underset{k_{off}}{\overset{k_{on}}{\rightleftharpoons}} \text{Buffer-} Ca^{2+},$$
 (8)

where k_{off} and k_{on} are two rates coefficients.

Next we let b the concentration of buffer- Ca^{2+} , B_{tot} the total concentration of the buffer and B the concentration of buffer only. An application of the law of mass action (see for e.g [14]), we obtain

$$\partial_t b = R(c, b) = k_{on} cB - k_{off} b. \tag{9}$$

We notice that $B = B_{tot} - b$ so the equation becomes:

$$\partial_t b = R(c, b) = k_{on} c(B_{tot} - b) - k_{off} b. \tag{10}$$

Each buffer (for more information, check [23]) is described by a conservation law similar to (1). So we conclude a similar PDE. The buffer list living in the cytosol is:

• ATP or nucleoside triphosphate used in the cells as a coenzyme often called the molecular currency of intracellular energy transfer.

$$\partial_t b_1 = D_{b_1} \Delta b_1 + R_1(c, b_1), \tag{11}$$

where D_{b_1} is a diffusion coefficient and the source term $R_1(c,b_1) := k_1^{on} c(B_1 - b_1) - k_1^{off} b_1$.

• CMDN or calmodulin is a multi-functional intermediate calcium-binding messenger protein.

$$\partial_t b_2 = D_{b_2} \Delta b_2 + R_2(c, b_2), \tag{12}$$

where D_{b_2} is a diffusion coefficient and the source term $R_2(c,b_2) := k_2^{on} c(B_2 - b_2) - k_2^{off} b_2$.

• Fluo-4 is used in the life sciences generally as a non destructive way of tracking or analyzing biological molecules such as Ca^{2+} .

$$\partial_t b_3 = D_{b_3} \Delta b_3 + R_3(c, b_3), \tag{13}$$

where D_{b_3} is a diffusion coefficient and the source term $R_3(c,b_3) = k_3^{on}c(B_3 - b_3) - k^{off}b_3$.

• TRPN or troponin is a protein of muscle that together with tropomyosin forms a regulatory protein complex controlling the interaction of actin and myosin and when combined with calcium ions permits muscular contraction. Similarly to Fluo-4, the protein TRPN satisfies the following equation

$$\partial_t b_4 = D_{b_4} \Delta b_4 + R_4(c, b_4), \tag{14}$$

where D_{b_4} is a diffusion coefficient and the source term $R_4(c, b_4) = k_4^{on} c(B_4 - b_4) - k^{off} b_4$. We consider only one buffer living in SR:

• CSQN or calsequestrin is a calcium binding protein. It helps hold calcium in the citerna of the sarcoplasmic reticulum after a muscle contraction. Moreover, the protein CSQN is governed by the equation

$$\partial_t b_5 = D_{b_5} \Delta b_5 + R_5(s, b_5), \tag{15}$$

where D_{b_5} is a diffusion coefficient and the source term $R_5(s, b_5) = k_5^{on} s(B_5 - b_5) - k^{off} b_5$. The interaction between buffers and calcium define the source terms R_c and R_s mentioned in (6) and (7).

$$R_c = -\sum_{i=1}^4 R_i, \qquad R_s = -R_5.$$

1.3. Boundary condition

In cardiac cell's, the calcium concentration in SR is considerably high. The activation of ionic channels and Ca^{2+} low concentration in cytosol induces a calcium flow throw RyR proportional to its difference between the two sub-domains:

$$J \cdot \eta = \pm q(s-c).$$

Due to Ficks law, we have

$$D_s(s)\nabla s \cdot \eta_s = g(s-c)$$
, on $\Gamma_{rur,T} := \Gamma_{rur} \times (0,T)$,

and

$$D_c(c)\nabla c \cdot \eta_c = -g(s-c)$$
 on $\Gamma_{rur,T} := \Gamma_{rur} \times (0,T)$,

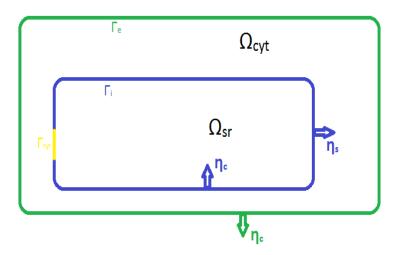


FIGURE 2. A Graphical representations of the boundary of the calcium model. We note the sarcoplasmic membrane $\Gamma^{sr} = \Gamma_i \cup \Gamma_{ryr}$ and cytosolic membrane $\Gamma = \Gamma_e \cup \Gamma_i \cup \Gamma_{ryr}$, while we consider $\Gamma_{iso}^{cyt} = \Gamma_e \cup \Gamma_i$ the impermeable part of the cytosolic membrane, $\Gamma_{iso}^{sr} = \Gamma_i$ is the impermeable part of the sarcoplasmic membrane.

where η_c (Resp. η_s) is the outward normal vector with respect to Ω_{cyt} (Resp. Ω_{sr}), Γ_{ryr} is subset of $\partial\Omega_{sr}\cup\partial\Omega_{cyt}$ and $\Gamma_{ryr,T} := \Gamma_{ryr} \times (0,T)$.

Since the SR and cell membrane are impermeable. Then, there is no flux through it. This can be implemented as an homogeneous Neumann boundary condition

$$D_s(s)\nabla s \cdot \eta_s = 0$$
 on $\Gamma_{iso,T}^{cyt} := \Gamma_{iso}^{cyt} \times (0,T),$

and

$$D_c(c)\nabla c \cdot \eta_c = 0 \text{ on } \Gamma^{sr}_{iso,T} := \Gamma^{sr}_{iso} \times (0,T),$$

where $\Gamma_{iso}^i := \partial \Omega_i - \Gamma_{ryr}$ for i = cyt, sr. The flux of buffers is null through cell membrane

$$\nabla b_i \cdot \eta_c = 0$$
, on $\Gamma_T := \Gamma \times (0, T)$, for $i = 1, 2, 3, 4$,

and

$$\nabla b_5 \cdot \eta_s = 0 \text{ on } \Gamma_T^{sr} := \Gamma^{sr} \times (0, T).$$

Finally, combining the previous equations, we get the following nonlinear reaction-diffusion system

$$\begin{cases} \partial_t c = \operatorname{div} \left(D_c(c) \nabla c \right) - \sum_{i=1}^4 R_i(c, b_i) & \text{in } \Omega_{cyt,T} := \Omega_{cyt} \times (0, T), \\ \partial_t b_i = D_{b_i} \Delta b_i + R_i(c, b_i) & \text{in } \Omega_{cyt,T} \text{ for } i = 1, \dots, 4, \\ \partial_t s = \operatorname{div} \left(D_s(s) \nabla s \right) - R_s(s, b_5) & \text{in } \Omega_{sr,T} := \Omega_{sr} \times (0, T), \\ \partial_t b_5 = D_{b_5} \Delta b_5 + R_5(s, b_5) & \text{in } \Omega_{sr,T}, \\ D_c(c) \nabla c \cdot \eta_c = 0 & \text{on } \Gamma_{iso,T}^{cyt} := \Gamma_{iso}^{cyt} \times (0, T), \\ D_s(s) \nabla s \cdot \eta_s = 0 & \text{on } \Gamma_{iso,T}^{sr} := \Gamma_{iso}^{sr} \times (0, T), \\ \nabla b_i \cdot \eta_c = 0 & \text{on } \Gamma_T := \Gamma \times (0, T), \quad i = 1, \dots, 4, \\ \nabla b_5 \cdot \eta_c = 0 & \text{on } \Gamma_{Tr}^{sr} := (\Gamma_i \cup \Gamma_{ryr}) \times (0, T), \\ D_s(s) \nabla s \cdot \eta_s = -g(s - c) & \text{on } \Gamma_{ryr,T} := \Gamma_{ryr} \times (0, T), \\ D_c(c) \nabla c \cdot \eta_c = g(s - c) & \text{on } \Gamma_{ryr,T} := \Gamma_{ryr} \times (0, T), \\ (c, b_i)(\cdot, 0) = (c_0, b_{i,0})(\cdot) \text{ in } \Omega_{cyt} \text{ and } (s, b_5)(\cdot, 0) = (s_0, b_{5,0})(\cdot) \text{ in } \Omega_{sr}. \end{cases}$$

In this system, the nonlinear diffusivities D_c, D_s are assumed continuous, positive, coercive, and there exist constants $0 < D^{\min} < D^{\max}$:

$$D_s, D_c$$
 continuous, $0 < D^{\min} \le D_s(m), D_c(m) \le D^{\max} < \infty$ for $m \in \mathbb{R}$,
and $0 < D^{\min} \le D_{b_i}$ where D_{b_i} is a constant diffusion for $i = 1, \dots, 4$. (17)

Remark 1. Because of the membrane impermeability, we considered in our model the Neumann boundary condition on buffers equations. For calcium dynamics in both sub-domain, the cell membrane contains the ionic channel RyR that allows only the transmission of Ca^{2+} . Hence, we have restricted the transmission boundary condition only for calcium equations on Γ_{ryr} . Moreover, we want to mention that the initial data $b_{i,0}$ of the buffer should be less than its total concentration B_i (Total concentration with or without calcium) for i = 1, ..., 5. In the paper [4], the author has studied the diffusivity of some buffers (In our case b_4 and b_5). Without loss of generality, in this work we consider the small diffusion coefficient in troponin (b_4) and calsequestrin (b_5) equations.

2. Well-posedness (existence and uniqueness) of the weak solution: Continuous case.

In this section, we introduce the variational formulation of the problem (16). Then, we look after existence results.

Definition 2.1 (L^2 bound solution). A weak L^2 bound solution of (16) is a seven component $(c, b_1, b_2, b_3, b_4, s, b_5)$ such that $c, b_i \in L^{\infty}(0, T; L^2(\Omega_{cyt})) \cap L^2(0, T; H^1(\Omega_{cyt}))$ and $s, b_5 \in L^{\infty}(0, T; L^2(\Omega_{sr})) \cap L^2(0, T; H^1(\Omega_{sr}))$,

 $\partial_t c, \partial_t b_i \in L^2(0,T;H^1(\Omega_{cut})'), \ \partial_t s, \partial_t b_5 \in L^2(0,T;H^1(\Omega_{sr})'), \ \text{and satisfying}$

$$\int_{0}^{T} \langle \partial_{t}c, \varphi_{c} \rangle_{cyt} dt + \iint_{\Omega_{cyt,T}} D_{c}(c) \nabla c \cdot \nabla \varphi_{c} dx dt - \iint_{\Gamma_{ryr,T}} g(s-c) \varphi_{c} d\sigma dt
= -\iint_{\Omega_{cyt,T}} \sum_{i=1}^{4} R_{i}(c, b_{i}) \varphi_{c} dx dt,
\int_{0}^{T} \langle \partial_{t}s, \varphi_{s} \rangle_{sr} dt + \iint_{\Omega_{sr,T}} D_{s}(s) \nabla s \cdot \nabla \varphi_{s} dx dt + \iint_{\Gamma_{ryr,T}} g(s-c) \varphi_{s} d\sigma dt
= -\iint_{\Omega_{sr,T}} R_{5}(s, b_{5}) \varphi_{s} dx dt,
\int_{0}^{T} \langle \partial_{t}b_{i}, \varphi_{i} \rangle_{cyt} dt + D_{b_{i}} \iint_{\Omega_{cyt,T}} \nabla b_{i} \cdot \nabla \varphi_{i} dx dt = \iint_{\Omega_{cyt,T}} R_{i}(c, b_{i}) \varphi_{i} dx dt,
\int_{0}^{T} \langle \partial_{t}b_{5}, \varphi_{5} \rangle_{sr} dt + D_{b_{5}} \iint_{\Omega_{cyt,T}} \nabla b_{5} \cdot \nabla \varphi_{5} dx dt = \iint_{\Omega_{sr,T}} R_{5}(c, b_{5}) \varphi_{5} dx dt,$$
(18)

for all $\varphi_c, \varphi_i \in L^2(0,T; H^1(\Omega_{cyt})), \varphi_s, \varphi_5 \in L^2(0,T; H^1(\Omega_{sr}))$ for $i = 1, \ldots, 4$. Here, $\langle \cdot, \cdot \rangle$ denotes the duality pairing between H^1 and $(H^1)'$

Definition 2.2 (L^{∞} bound solution). A weak L^{∞} bound solution of (16) is a nonnegative and a L^{∞} function with seven components $(c, b_1, b_2, b_3, b_4, s, b_5)$ satisfying Definition 2.1.

Our first main result is the following existence theorem for weak solutions.

Theorem 2.3. Assume that condition (17) holds. If $c_0, b_{1,0}, b_{2,0}, b_{3,0}, b_{4,0} \in L^{\infty}(\Omega_{cyt})$ and $s_0, b_{5,0} \in L^{\infty}(\Omega_{sr})$, then the system (16) possesses a unique weak solution in sense of Definition 2.2.

To prove theorem 2.3 we first prove existence of solutions to the approximate problem (19) below by applying the Schauder fixed-point theorem (in an appropriate functional setting). Then we send the regularization parameter ϵ to zero to produce a weak solution of the original system (16) as the limit of a sequence of such approximate solutions. Convergence is achieved by means of a priori estimates and compactness arguments.

2.1. Existence result to the regularized problem: the fixed-point method

In this subsection we prove, for each fixed $\varepsilon > 0$, the existence of solutions to the approximate problem (19) below, by applying the Schauder fixed-point theorem. For this purpose, we introduce the following closed subset of the Banach space

$$\mathcal{A} = \{(c,s) \in H: 0 \le c(t,x_c), s(t,x_s) \le R \text{ for a.e. } (t,x_c) \in \Omega_{cyt,T} \text{ and } (t,x_s) \in \Omega_{sr,T}\},$$

for some constant R > 0. Herein, $H := L^2(\Omega_{cyt,T}) \times L^2(\Omega_{sr,T})$. For a given nonnegative function $\hat{U} = (\hat{c}, \hat{s})$ such that $\hat{c} \in L^2(\Omega_{cyt,T})$ and $\hat{s} \in L^2(\Omega_{sr,T})$, we solve the approximate system

$$\begin{cases} \partial_t c = \operatorname{div} \left(D_c(\hat{c}) \nabla c \right) - \sum_{i=1}^4 R_{i,\varepsilon}(c^+, b_i^+) & \text{in } \Omega_{cyt,T}, \\ \partial_t b_i = D_{b_i} \Delta b_i + R_{i,\varepsilon}(\hat{c}^+, b_i^+) & \text{in } \Omega_{cyt,T}, \text{ for } i = 1, \dots, 4, \\ \partial_t s = \operatorname{div} \left(D_s(\hat{s}) \nabla s \right) - R_{s,\varepsilon}(s^+, b_5^+) & \text{in } \Omega_{sr,T}, \\ \partial_t b_5 = D_{b_5} \Delta b_5 + R_{5,\varepsilon}(\hat{s}^+, b_5^+) & \text{in } \Omega_{sr,T}, \\ D_c(c) \nabla c \cdot \eta_c = 0 & \text{on } \Gamma_{iso,T}^{cyt}, \\ D_s(s) \nabla s \cdot \eta_s = 0 & \text{on } \Gamma_{iso,T}^{sr}, \\ \nabla b_i \cdot \eta_c = 0 & \text{on } \Gamma_T^{sr}, \text{ for } i = 1, \dots, 4 \\ \nabla b_5 \cdot \eta_s = 0 & \text{on } \Gamma_T^{sr}, \\ D_s(s) \nabla s \cdot \eta_s = -g(s - c) & \text{on } \Gamma_{ryr,T}, \\ D_c(c) \nabla c \cdot \eta_c = g(s - c) & \text{on } \Gamma_{ryr,T}, \\ D_c(c) \nabla c \cdot \eta_c = g(s - c) & \text{on } \Gamma_{ryr,T}, \\ (c, b_i)(\cdot, 0) = (c_0, b_{i,0})(\cdot) \text{ in } \Omega_{cyt} \text{ and } & (s, b_5)(\cdot, 0) = (s_0, b_{5,0})(\cdot) \text{ in } \Omega_{sr}, \end{cases}$$

defined for each $\varepsilon > 0$. Herein,

$$R_{\kappa,\varepsilon} = \frac{R_{\kappa}}{1 + \varepsilon |R_{\kappa}|} \text{ for } \kappa = 1,\dots,5.$$
 (20)

Next, we introduce the map \mathcal{F} defined by

$$\begin{array}{cccc} \mathcal{F}: & \mathcal{A} & \longmapsto & \mathcal{A} \\ & \hat{U} & \longrightarrow & U. \end{array}$$

where U = (c, s) solves the approximate system (19). The following lemma shows some estimates that will guarantee the continuity and compactness of \mathcal{F} below.

Lemma 2.4. Let $c_0, b_{1,0}, b_{2,0}, b_{3,0}, b_{4,0} \in L^{\infty}(\Omega_{cyt})$ and $s_0, b_{5,0} \in L^{\infty}(\Omega_{sr})$, then for any $(\hat{c}, \hat{s}) \in L^2(\Omega_{cyt,T}) \times L^2(\Omega_{sr,T})$, the system (19) has a unique solution. Moreover, there exists a constant C > 0 depending on the initial conditions, such that

$$||c||_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))} + ||c||_{L^{2}(0,T;H^{1}(\Omega_{cyt}))} \leq C,$$

$$||b_{i}||_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))} + ||b_{i}||_{L^{2}(0,T;H^{1}(\Omega_{cyt}))} \leq C \text{ for } i = 1,\dots,4,$$

$$||s||_{L^{\infty}(0,T;L^{2}(\Omega_{sr}))} + ||s||_{L^{2}(0,T;H^{1}(\Omega_{sr}))} \leq C,$$

$$||b_{5}||_{L^{\infty}(0,T;L^{2}(\Omega_{sr}))} + ||b_{5}||_{L^{2}(0,T;H^{1}(\Omega_{sr}))} \leq C.$$

$$(21)$$

Proof. For the proof it suffices to use assumptions on the initial conditions and the definition of functions R_i (for i = 1, ..., 5) combined with the general result for parabolic equations (see for e.g. for more details [15]). \square

To prove the existence and uniqueness of (16), we apply now the Schauder fixed point theorem to the map \mathcal{F} . The goal is to prove that such map has a fixed-point. First, we notice that \mathcal{F} is well defined. To satisfy the hypothesis of the fixed point theorem, we show that \mathcal{F} is continuous and compact mapping. Firstly, we prove the continuity of \mathcal{F} . Let (\hat{c}_n, \hat{s}_n) be a sequence in \mathcal{A} and $(\hat{c}, \hat{s}) \in \mathcal{A}$ be such that

$$(\hat{c}_n, \hat{s}_n) \to (\hat{c}, \hat{s})$$
 in $L^2(\Omega_{cyt,T}) \times L^2(\Omega_{sr,T})$,

as $n \to \infty$. Let us then define

$$(c_n, s_n) = \mathcal{F}(\hat{c}_n, \hat{s}_n),$$

i.e., (c_n, s_n) is the solution of (19) associated with the sequence (\hat{c}_n, \hat{s}_n) . The objective is to show that:

$$(c_n, s_n)$$
 converges to $\mathcal{F}(\hat{c}, \hat{s})$,

in $L^2(\Omega_{cut,T}) \times L^2(\Omega_{sr,T})$. We start with the following lemma:

Lemma 2.5. Let $(c_n, b_{1,n}, b_{2,n}, b_{3,n}, b_{4,n}, s_n, b_{5,n})_n$ be the solution to problem (19). Then

(i) There exists a constant C > 0 such that for i = 1, ..., 4

$$0 < c_n(x_c, t), b_{i,n}(x_c, t), s_n(x_s, t), b_{5,n}(x_s, t) < C$$

for a.e. $(x_c, t) \in \Omega_{cyt,T}$ and $(x_s, t) \in \Omega_{sr,T}$.

 $\begin{array}{ll} \mbox{(ii)} & The \ sequence \ (c_n,b_{1,n},b_{2,n},b_{3,n},b_{4,n},b_{5,n},s_n)_n \ is \ bounded \ in \\ & \left(L^2(0,T;H^1(\Omega_{cyt}))\cap L^\infty(0,T;L^2(\Omega_{cyt}))\right)^5 \times \left(L^2(0,T;H^1(\Omega_{sr}))\cap L^\infty(0,T;L^2(\Omega_{sr}))\right)^2. \end{array}$

(iii) The sequence $(c_n, b_{1,n}, b_{2,n}, b_{3,n}, b_{4,n}, b_{5,n}s_n)_n$ is relatively compact in $(L^2(\Omega_{cyt,T}))^5 \times (L^2(\Omega_{sr,T}))^2$.

Proof. To prove (i), we multiply the first and the third equations in (19) by $-c_n^-$ and $-s_n^-$ (where $\alpha^- := (\alpha - |\alpha|)/2$) and we integrate over Ω_{cyt} and Ω_{sr} , respectively. The result is

$$\frac{1}{2} \frac{d}{dt} \int_{\Omega_{cyt}} |c_n^-|^2 dx - \int_{\Omega_{cyt}} D_c(\hat{c}_n) \nabla c_n \cdot \nabla c_n^- dx + \int_{\Gamma_{ryr}} g(s_n - c_n) c_n^- d\sigma dt
= \sum_{i=1}^4 \int_{\Omega_{cyt}} R_{i,\varepsilon}(c_n^+, b_{i,n}^+) c_n^- dx \le 0,$$

and

$$\frac{1}{2} \frac{d}{dt} \int_{\Omega_{sr}} \left| s_n^- \right|^2 dx - \int_{\Omega_{sr}} D_s(\hat{s}_n) \nabla s_n \cdot \nabla s_n^- dx - \int_{\Gamma_{ryr}} g(s_n - c_n) s_n^- d\sigma dt
= \int_{\Omega_{sr}} R_{s,\varepsilon}(c_n^+, b_{i,n}^+) s_n^- dx \le 0.$$

Using this and (17), we deduce

$$\frac{1}{2}\frac{d}{dt}||c_n^-||_{L^2(\Omega_{cyt})}^2 + \frac{1}{2}\frac{d}{dt}||s_n^-||_{L^2(\Omega_{sr})}^2 + \iint_{\Gamma_{rur}} g(s-c)(c_n^- - s_n^-)d\sigma\,dt \leq 0.$$

Observe that the function $(\cdot)^-$ is a decreasing function. This implies

$$\iint_{\Gamma_{rur,T}} g(s_n - c_n)(c_n^- - s_n^-) d\sigma \, dt \ge 0.$$

Therefore we obtain

$$\frac{1}{2}\frac{d}{dt}||c_n^-||^2_{L^2(\Omega_{cyt})} + \frac{1}{2}\frac{d}{dt}||s_n^-||^2_{L^2(\Omega_{sr})} \leq 0.$$

Since the initial condition (c_0, s_0) is nonnegative we conclude

$$c_n(t, x_c), s_n(t, x_s) \ge 0$$
 for a.e. $(t, x_c) \in \Omega_{cyt,T}$ and $(t, x_s) \in \Omega_{sr,T}$. (22)

Reasoning similarly we get the nonnegativity of $b_{i,n}$ for $i=1,\ldots,5$.

In the next step we prove first (recall that B_i is the total concentration of the buffer)

$$b_{i,n}(t,x_c) \le B_i$$
 and $b_{5,n}(t,x_s) \le B_5$ for a.e. $(t,x_c) \in \Omega_{cyt,T}$ and $(t,x_s) \in \Omega_{sr,T}$, (23)

for $i=1,\ldots,4$. For this, we multiply $(b_{i,n}-B_i)^+$ (where $\alpha^+:=(\alpha+|\alpha|)/2$) by the corresponding equation of $b_{i,n}$ in (19) and we integrate over Ω_{cyt} for $i=1,\ldots,4$ and Ω_{sr} for i=5, respectively, to deduce from the definition of $R_{i,\varepsilon}$

$$\frac{1}{2} \sum_{i=1}^{4} \frac{d}{dt} ||((b_{i,n} - B_i)^+||_{L^2(\Omega_{cyt})}^2 + \frac{1}{2} \frac{d}{dt} ||((b_{5;n} - B_5)^+||_{L^2(\Omega_{sr})}^2 + \sum_{i=1}^{3} ||\nabla (b_{i,n} - B_i)^+||_{L^2(\Omega_{sr},T)}^2 \\
= \sum_{i=1}^{4} \iint_{\Omega_{cyt,T}} R_{i,\varepsilon} (\hat{c}_n, b_{i,n}) (b_{i,n} - B_i)^+ dx dt + \iint_{\Omega_{sr}} R_{5,\varepsilon} (\hat{s}_n, b_{5,n}) (b_{5,n} - B_5)^+ dx dt \le 0.$$

Using this and the Gronwall lemma we get (23). To complete the prove, let us define a super solution to the equations of calcium:

$$\begin{cases}
\frac{d}{dt}M(t) = -\sum_{i=1}^{5} R_i(M(t), 0) = -\sum_{i=1}^{5} k_i^{on}M(t)B_i, \\
M(0) = ||c_0||_{L^{\infty}(\Omega_{cyt})} + ||s_0||_{L^{\infty}(\Omega_{sr})}.
\end{cases}$$
(24)

Now, we multiply the first and the third equations in (19) by $(c_n - M(t))^+$ and $(s_n - M(t))^+$, respectively, and we integrate over Ω_{cyt} and Ω_{sr} , respectively, and we sum the resulting equations to obtain

$$\frac{1}{2} \frac{d}{dt} || (c_n - M(t))^+ ||_{L^2(\Omega_{cyt})}^2 + \frac{1}{2} \frac{d}{dt} || (s_n - M(t))^+ ||_{L^2(\Omega_{sr})}^2 + || \nabla (c_n - M(t))^+ ||_{L^2(\Omega_{cyt,T})}^2
+ || \nabla (s_n - M(t))^+ ||_{L^2(\Omega_{sr,T})}^2 + \iint_{\Gamma_{ryr}} g(s_n - c_n) ((s_n - M(t))^+ - (c_n - M(t))^+) d\sigma dt
= \sum_{i=1}^4 \iint_{\Omega_{cyt,T}} (R_{i,\varepsilon}(M(t), 0) - R_{i,\varepsilon}(c_n, b_{i,n})) (c_n - M(t))^+ dx dt
+ \iint_{\Omega_{sr}} (R_{s,\varepsilon}(M(t), 0) - R_{s,\varepsilon}(s_n, b_{5,n})) (s_n - M(t))^+ dx dt
= \sum_{i=1}^4 \iint_{\Omega_{cyt,T}} (R_{i,\varepsilon}(M(t), 0) - R_{i,\varepsilon}(c_n, 0) + R_{i,\varepsilon}(c_n, 0) - R_{i,\varepsilon}(c_n, b_{i,n})) (c_n - M(t))^+ dx dt
+ \iint_{\Omega_{sr}} (R_{s,\varepsilon}(M(t), 0) - R_{s,\varepsilon}(s_n, 0) + R_{s,\varepsilon}(s_n, 0) - R_{s,\varepsilon}(s_n, b_{5,n})) (s_n - M(t))^+ dx dt.$$
(25)

By monotonicity of $(\cdot - M(t))^+$ we have $\iint_{\Gamma_{ryr,T}} g(s_n - c_n)((s_n - M(t))^+ - (c_n - M(t))^+)d\sigma dt \ge 0$. For the right hand terms, we assume that $c_n \ge M(t)$ and $s_n \ge M(t)$. Then, by monotonicity of $R_{i,\varepsilon}$ we conclude that :

$$\frac{1}{2}\frac{d}{dt}||(c_n - M(t))^+||_{L^2(\Omega_{cyt})}^2 + \frac{1}{2}\frac{d}{dt}||(s_n - M(t))^+||_{L^2(\Omega_{sr})}^2 \le 0.$$
 (26)

Therefore, we have $c_n(t, x_c)$, $s_n(t, x_s) \leq M(t)$ for all $(t, x_c) \in \Omega_{cyt,T}$ and $(t, x_s) \in \Omega_{sr,T}$. Moreover, an application of the Gronwall lemma on (24) we obtain

$$M(t) \le M(0)e^{-tC} \le ||c_0||_{L^{\infty}(\Omega_{cyt})} + ||s_0||_{L^{\infty}(\Omega_{sr})} \text{ with } C = \sum_{i=1}^5 B_i k_i^{on}.$$

To prove (ii), we multiply the first, the second, the third, the fourth and the fifth equation in (19) by c_n , $b_{i,n}$ (for $i=1,\ldots,4$), s_n and $b_{5,n}$ respectively. Herein, we replace the solution $(c,b_1,b_2,b_3,b_4,s,b_5)$ by $(c_n,b_{1,n},b_{2,n},b_{3,n},b_{4,n},s_n,b_{5,n})$. Next, we sum and integrate the resulting equation on (0,T), on Ω_{cyt} for $(c_n,b_{1,n},b_{2,n},b_{3,n},b_{4,n})$ and on Ω_{sr} for $(s_n,b_{5,n})$. Hence the Gronwall lemma and Poincare inequality yields

$$\|(c_n,b_{1,n},b_{2,n},b_{3,n},b_{4,n})\|_{(L^2(0,T;H^1(\Omega_{cyt}))\cap L^\infty(0,T;L^2(\Omega_{cyt})))^5} \le C,$$

and

$$||(s_n, b_{5,n})||_{(L^2(0,T;H^1(\Omega_{sr}))\cap L^{\infty}(0,T;L^2(\Omega_{sr})))^2} \le C,$$

for some constant C > 0 not depending on n. Finally, note that from (i) and (ii) we get easily for $i = 1, \ldots, 4$

$$\|\partial_t c_n\|_{L^2(0,T;(H^1(\Omega_{cyt}))')} + \|\partial_t s_n\|_{L^2(0,T;(H^1(\Omega_{sr}))')} + \|\partial_t b_{i,n}\|_{L^2(0,T;(H^1(\Omega_{cyt}))')} + \|\partial_t b_{5,n}\|_{L^2(0,T;(H^1(\Omega_{sr}))')} \le C,$$

for some constant C > 0 not depending on n. Therefore, (iii) is a consequence of this, (ii) and the compactness argument (see for e.g. [30] for more details). This concludes the proof of Lemma 2.5.

In summary, Lemma 2.5 and the theory of compact sets [30], imply that there exists $(c, b_1, b_2, b_3, b_4, s, b_5) \in (L^2(0, T; H^1(\Omega_{cyt})))^5 \times (L^2(0, T; H^1(\Omega_{sr})))^2$ such that, up to extracting subsequences if necessary,

$$\begin{cases} & (c_n, b_{i,n}, b_{2,n}, b_{3,n},, b_{4,n}, b_{5,n}, s_n) \rightarrow (c, b_1, b_2, b_3, b_4, b_5, s) \text{ in } (L^2(\Omega_{cyt,T}))^5 \times (L^2(\Omega_{sr,T}))^2 \\ & \text{strongly and a.e., and in } (L^2(0, T; H^1(\Omega_{cyt})))^5 \times (L^2(0, T; H^1(\Omega_{sr})))^2 \text{ weakly.} \end{cases}$$

Consequently, the continuity of \mathcal{F} on \mathcal{A} holds.

Moreover, using again Lemma 2.5 and the theory of compact sets [30], we get the compactness of the map $\mathcal{F}(\mathcal{A}) \hookrightarrow L^2(\Omega_{cyt,T}) \times L^2(\Omega_{sr,T})$. Then, thanks to Schauder fixed-point theorem, the operator \mathcal{F} has a fixed point $(c_{\varepsilon}, s_{\varepsilon})$. That is, there exists a solution to the approximate problem (19). That is there exists a solution to

$$\int_{0}^{T} \langle \partial_{t} c_{\varepsilon}, \varphi_{c} \rangle dt + \iint_{\Omega_{cyt,T}} D_{c}(c_{\varepsilon}) \nabla c_{\varepsilon} \cdot \nabla \varphi_{c} dx dt - \int_{0}^{T} \int_{\Gamma_{ryr}} g(s_{\varepsilon} - c_{\varepsilon}) \varphi_{c} \sigma dt \\
= -\iint_{\Omega_{cyt,T}} \sum_{i=1}^{4} R_{i,\varepsilon}(c_{\varepsilon}, b_{i,\varepsilon}) \varphi_{c} dx dt, \\
\int_{0}^{T} \langle \partial_{t} s_{\varepsilon}, \varphi_{s} \rangle dt + \iint_{\Omega_{sr,T}} D_{s}(s_{\varepsilon}) \nabla s_{\varepsilon} \cdot \nabla \varphi_{s} dx dt + \int_{0}^{T} \int_{\Gamma_{ryr}} g(s_{\varepsilon} - c_{\varepsilon}) \varphi_{s} \sigma dt \\
= -\iint_{\Omega_{sr,T}} R_{s,\varepsilon}(s_{\varepsilon}, b_{5,\varepsilon}) \varphi_{s} dx dt, \\
\int_{0}^{T} \langle \partial_{t} b_{i,\varepsilon}, \varphi_{i} \rangle dt + D_{b_{i}} \iint_{\Omega_{cyt,T}} \nabla b_{i,\varepsilon} \cdot \nabla \varphi_{i} dx dt = \iint_{\Omega_{cyt,T}} R_{i,\varepsilon}(c_{\varepsilon}, b_{i,\varepsilon}) \varphi_{i} dx dt, \\
\int_{0}^{T} \langle \partial_{t} b_{5,\varepsilon}, \varphi_{5} \rangle dt + D_{b_{5}} \iint_{\Omega_{sr,T}} \nabla b_{5,\varepsilon} \cdot \nabla \varphi_{5} dx dt = \iint_{\Omega_{sr,T}} R_{5,\varepsilon}(c_{\varepsilon}, b_{5,\varepsilon}) \varphi_{5} dx dt,$$

for all $\varphi_c, \varphi_i \in L^2(0,T; H^1(\Omega_{cyt}))$ and $\varphi_s, \varphi_5 \in L^2(0,T; H^1(\Omega_{sr}))$ for $i = 1, \dots, 4$.

2.2. Existence of weak solutions

We have shown in Section 2.1 that problem (27) admits a solution $(c_{\varepsilon}, b_{1,\varepsilon}, b_{2,\varepsilon}, b_{3,\varepsilon}, b_{4,\varepsilon}, s_{\varepsilon}, b_{5,\varepsilon})$. The goal in this subsection is to send the regularization parameter ε to zero in sequences of such solutions to obtain weak solutions of the original system (16). Observe that, for each fixed $\varepsilon > 0$, we have shown the existence of a solution. $(c_{\varepsilon}, b_{i,\varepsilon}, b_{2,\varepsilon}, b_{3,\varepsilon}, b_{4,\varepsilon}, s_{\varepsilon}, b_{5,\varepsilon})$ to (27) such that for $i = 1, \ldots, 4$

$$0 \le c_{\varepsilon}(x_c, t), b_{i,\varepsilon}(x_c, t), s_{\varepsilon}(x_s, t), b_{5,\varepsilon}(x_s, t) \le M,$$

for a.e. $(x_c,t) \in \Omega_{cyt,T}$ and $(x_s,t) \in \Omega_{sr,T}$. Herein, the constant M > 0 is not depending on ε . Now, we substitute $\varphi_c = c_{\varepsilon}$, $\varphi_s = s_{\varepsilon}$, $\varphi_i = b_{i,\varepsilon}$ in (27) for $i = 1, \ldots, 5$, and we work exactly as in the proof of (ii) in

Lemma 2.5, to obtain for i = 1, ..., 4

$$\sup_{0 \le t \le T} \int_{\Omega_{cyt}} |c_{\varepsilon}(x,t)|^{2} dx + \sup_{0 \le t \le T} \int_{\Omega_{sr}} |s_{\varepsilon}(x,t)|^{2} dx + \sup_{0 \le t \le T} \int_{\Omega_{cyt}} |b_{i,\varepsilon}(x,t)|^{2} dx + \sup_{0 \le t \le T} \int_{\Omega_{sr}} |b_{5,\varepsilon}(x,t)|^{2} dx \le C,$$
(28)

and

$$\iint_{\Omega_{cyt,T}} |\nabla c_{\varepsilon}(x,t)|^{2} dx dt + \iint_{\Omega_{sr,T}} |\nabla s_{\varepsilon}(x,t)|^{2} dx dt + \iint_{\Omega_{cyt,T}} |\nabla b_{i,\varepsilon}(x,t)|^{2} dx dt + \iint_{\Omega_{sr,T}} |\nabla b_{5,\varepsilon}(x,t)|^{2} dx dt \leq C,$$
(29)

for some constant C > 0 independent of ε . Repeating the steps of the proof of (iii) in Lemma 2.5, we derive the bound for i = 1, ..., 4,

$$\|\partial_{t}c_{\varepsilon}\|_{L^{2}(0,T;(H^{1}(\Omega_{cyt}))')} + \|\partial_{t}s_{\varepsilon}\|_{L^{2}(0,T;(H^{1}(\Omega_{sr}))')} + \|\partial_{t}b_{i,\varepsilon}\|_{L^{2}(0,T;(H^{1}(\Omega_{cyt}))')} + \|\partial_{t}b_{5,\varepsilon}\|_{L^{2}(0,T;(H^{1}(\Omega_{sr}))')} \leq C,$$
(30)

for some constant C > 0. Then, combining (28), (29) and (30) with standard compactness results (cf. [30]) we can extract subsequences, which we do not relabel, such that, as ε goes to 0,

$$\begin{cases}
(c_{\varepsilon}, b_{i,\varepsilon}, s_{\varepsilon}, b_{5,\varepsilon}) \to (c, b_{i}, s, b_{5}) \text{ in } (L^{2}(\Omega_{cyt,T})^{2}) \times (L^{2}(\Omega_{sr,T}))^{2}, \\
\text{strongly and a.e., and in } (L^{2}(0, T; H^{1}(\Omega_{cyt})))^{2} \times (L^{2}(0, T; H^{1}(\Omega_{sr})))^{2} \text{ weakly,} \\
(\partial_{t} c_{\varepsilon}, \partial_{t} b_{i,\varepsilon}) \to (\partial_{t} c, \partial_{t} b_{i}) \text{ weakly in } (L^{2}(0, T; (H^{1}(\Omega_{cyt}))'))^{2}, \\
(\partial_{t} s_{\varepsilon}, \partial_{t} b_{5,\varepsilon}) \to (\partial_{t} s, \partial_{t} b_{5}) \text{ weakly in } (L^{2}(0, T; (H^{1}(\Omega_{sr}))'))^{2},
\end{cases}$$
(31)

for i = 1..., 4. This result, together with the weak-* convergences in L^{∞} of $(c_{\varepsilon}, b_{1,\varepsilon}, b_{2,\varepsilon}, b_{3,\varepsilon}, s_{\varepsilon})$ to (c, b_1, b_2, b_3, s) , yields

$$\begin{cases} (c_{\varepsilon}, b_{1,\varepsilon}, b_{2,\varepsilon}, b_{3,\varepsilon}, b_{4,\varepsilon}) \to (c, b_1, b_2, b_3, b_4), (s_{\varepsilon}, b_{5,\varepsilon}) \to (s, b_5) \text{ strongly in } (L^p(\Omega_{cyt,T}))^5, \\ \text{and in } (L^p(\Omega_{sr,T}))^2, \text{ respectively, for } 1 \le p < \infty, \end{cases}$$
(32)

which in turn implies that for i = 1, ..., 5

$$R_{i,\varepsilon}(c_{\varepsilon}, b_{i,\varepsilon}) \to R_i(c, b_i), \quad R_{s,\varepsilon}(s_{\varepsilon}, b_{5,\varepsilon}) \to R_s(s, b_5),$$
 (33)

a.e. and strongly in L^p for $1 \le p < \infty$. As a consequence of (31), (32) and (33), it is clear that as $\varepsilon \to 0$ in (27), we can identify the limit as the weak solution from Definition 2.2.

Remark 2. Note that, since the interaction terms are Lipschitz (recall that our solutions are uniformly bounded in L^{∞}) and the positivity of diffusion function, the uniqueness of weak solution can be obtained easily (for a similar proof see [1]).

3. Numerical approximation

In this section we construct our finite element scheme for the system (16). We establish several a priori (discrete energy) estimates for our scheme and compactness of discrete solutions which eventually will imply the desired convergence results to the unique weak solution.

3.1. The semi-discrete scheme

We consider a quasi-uniform family $\mathcal{T}_{i,h}$ of triangulations of $\bar{\Omega}_i$ by tetrahedra K, for i=cyt,sr. The parameter h has the sense of an upper bound for the maximum diameter of the control volumes in $\mathcal{T}_{i,h}$ for i=cyt,sr. For

a given an integer $k \geq 0$ and $S \subset \mathbb{R}^3$, we denote by $\mathcal{P}_k(S)$ the space of polynomial functions defined in S of total degree up to k. Next, we define the finite element subspaces $\mathcal{V}_{i,h}$ by

$$\mathcal{V}_{i,h} = \left\{ m_h \in H^1(\Omega_i) : m_h|_K \in \mathcal{P}_1(K) \, \forall K \in \mathcal{T}_{i,h} \right\},\,$$

where for any $K \in \mathcal{T}_{i,h}(\Omega_i)$ for i = cyt, sr. respectively.

We consider the appropriate L^2 -projections of the initial condition $(c_h^0, b_{i,h}^0) = \mathbb{P}_{\mathcal{V}_{cyt,h}}(c_0, b_{i,0})$ (for $i = 1, \ldots, 4$) and $(s_h^0, b_{5,h}^0) = \mathbb{P}_{\mathcal{V}_h}(s_0, b_{5,0})$. To advance the numerical solution from t_{n-1} to $t_n = t_{n-1} + \delta t$ ($\delta t = T/N$), we use the following implicit finite element scheme:

For $n \in \{1, \dots, N\}$, find $(c_h^n, b_{1,h}^n, b_{2,h}^n, b_{3,h}^n, b_{4,h}^n) \in (\mathcal{V}_{cyt,h})^5$ and $(s_h^n, b_{5,h}^n) \in (\mathcal{V}_{sr,h})^2$ such that

$$\left(\frac{c_h^n - c_h^{n-1}}{\delta t}, \varphi_c\right)_{\Omega_{cyt}} + \int_{\Omega_{cyt}} D_c(c_h^n) \nabla c_h^n \cdot \nabla \varphi_c \, dx - \int_{\Gamma_{ryr}} g(s_h^n - c_h^n) \varphi_c d\sigma
= -\sum_{i=1}^4 \left(R_i(c_h^n, b_{i,h}^n), \varphi_c\right)_{\Omega_{cyt}},
\left(\frac{s_h^n - s_h^{n-1}}{\delta t}, \varphi_s\right)_{\Omega_{sr}} + \int_{\Omega_{sr}} D_s(s_h^n) \nabla s_h^n \cdot \nabla \varphi_s \, dx + \int_{\Gamma_{ryr}} g(s_h^n - c_h^n) \varphi_s d\sigma
= -\left(R_5(s_h^n, b_{5,h}^n), \varphi_s\right)_{\Omega_{sr}},
\left(\frac{b_{i,h}^n - b_{i,h}^{n-1}}{\delta t}, \varphi_i\right)_{\Omega_{cyt}} + \int_{\Omega_{cyt}} D_{b_i} \nabla b_{i,h}^n \cdot \nabla \varphi_i \, dx = \left(R_i(c_h^n, b_{i,h}^n), \varphi_i\right)_{\Omega_{cyt}},
\left(\frac{b_{5,h}^n - b_{5,h}^{n-1}}{\delta t}, \varphi_5\right)_{\Omega_{sr}} + \int_{\Omega_{sr}} D_{b_5} \nabla b_{5,h}^n \cdot \nabla \varphi_5 \, dx = \left(R_5(s_h^n, b_{5,h}^n), \varphi_5\right)_{\Omega_{sr}},$$

$$(34)$$

for all $\varphi_c, \varphi_i \in \mathcal{V}_{cyt,h}$ and $\varphi_s, \varphi_5 \in \mathcal{V}_{sr,h}$ for $i = 1, \dots, 4$.

3.2. Numerical convergence analysis

Note that the problem of proving uniform boundedness of the discrete finite element solutions is still an open problem (recall that the positive and negative cuts test functions are not admissible in finite element context). So, comparing to the continuous case, we assume that the initial conditions $(c_0, b_{1,0}, b_{2,0}, b_{3,0}, b_{4,0})$ and $(s_0, b_{5,0})$ are only bounded in L^2 .

The convergence of solutions generated by the scheme (34) is next established following two main lemmas. The first one states $L^{\infty}(L^2) \cap L^2(H^1)$ boundedness of the discrete concentrations and buffers $(c_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}, b_{5,h})$. We establish several a priori (discrete energy) estimates, which eventually will imply the desired convergence results.

Remark 3. In the proof of Lemmas 3.1 and 3.2 below, we will use the following Gagliardo-Nierenberg inequality: For $\Omega \subset \mathbb{R}^3$ there exists a constant G such that for $u \in H^1(\Omega)$

$$||u||_{L^{q}(\Omega)} \le G ||u||_{H^{1}(\Omega)}^{\alpha} ||u||_{L^{2}(\Omega)}^{1-\alpha} \text{ for } \frac{1}{q} = \frac{3-2\alpha}{6} \text{ and } \alpha \in (0,1).$$
 (35)

Lemma 3.1. Let $(c_h^n, b_{1,h}^n, b_{2,h}^n, b_{3,h}^n, b_{4,h}^n, s_h^n, b_{5,h}^n)$ be part of the solution of (34). Then there exists a constant C > 0 depending on $\|c_0\|_{L^2(\Omega_{cyt})}, \|b_{i,0}\|_{L^2(\Omega_{cyt})}$, and $\|s_0\|_{L^2(\Omega_{sr})}, \|b_{5,0}\|_{L^2(\Omega_{sr})}$ such that for $i = 1, \ldots, 4$

$$\|(c_{h}, b_{i,h})\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt},\mathbb{R}^{2}))} + \|(s_{h}, b_{5,h})\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt},\mathbb{R}^{2}))} \leq C,$$

$$\|\nabla c_{h}\|_{L^{2}(\Omega_{T,cyt})} + \sum_{i=1}^{3} \|\nabla b_{i,h}\|_{L^{2}(\Omega_{T,cyt})} + \|\nabla s_{h}\|_{L^{2}(\Omega_{T,sr})} \leq C.$$

$$(36)$$

Proof. We use (34) with $\phi_c = c_h^n$, $\phi_s = -s_h^n$ and $\phi_i = b_{i,h}^n$ (for i = 1, ..., 5), and we sum over n = 1, ..., k for all $1 < k \le N$. The result is

$$\begin{split} &\frac{1}{2}\Bigg((c_h^k,c_h^k)_{\Omega_{cyt}} + (s_h^k,s_h^k)_{\Omega_{sr}} + \sum_{i=1}^4 (b_{i,h}^k,b_{i,h}^k)_{\Omega_{cyt}} + (b_{5,h}^k,b_{5,h}^k)_{\Omega_{sr}}\Bigg) \\ &+ D^{\min}\Bigg(\sum_{n=1}^k \int_{\Omega_{cyt}} \Big|\nabla c_h^n\Big|^2 + \sum_{n=1}^k \int_{\Omega_{sr}} \Big|\nabla s_h^n\Big|^2\Bigg) \\ &+ \int_{\Gamma_{ryr}} g \left|s_h^n - c_h^n\right|^2 d\sigma \\ &\leq \frac{1}{2}\Bigg((c_{0,h},c_{0,h})_{\Omega_{cyt}} + (s_{0,h},s_{0,h})_{\Omega_{sr}} + \sum_{i=1}^4 (b_{i,0,h},b_{i,0,h})_{\Omega_{cyt}} + (b_{5,0,h},b_{5,0,h})_{\Omega_{sr}}\Bigg) \\ &+ C_1\Bigg(\sum_{n=1}^k \int_{\Omega_{cyt}} \Big(|c_h^n|^2 + \sum_{i=1}^4 \Big|b_{i,h}^n\Big|^2\Big) + \sum_{n=1}^k \int_{\Omega_{sr}} \Big(|s_h^n|^2 + \Big|b_{5,h}^n\Big|^2\Big)\Bigg) \\ &+ C_2\Bigg(\sum_{i=1}^4 \sum_{n=1}^k \int_{\Omega_{cyt}} \Big(\Big|(c_h^n)^2 b_{i,h}^n\Big| + \Big|c_h^n(b_{i,h}^n)^2\Big|\Big) + \sum_{n=1}^k \int_{\Omega_{sr}} \Big(\Big|(s_h^n)^2 b_{5,h}^n\Big| + \Big|s_h^n(b_{5,h}^n)^2\Big|\Big)\Bigg) \\ &:= \frac{1}{2}I_1 + C_1I_2 + C_3I_3, \end{split}$$

for some constant $C_1, C_2 > 0$. Herein, we have used the positivity of D_c and D_s , and the convexity inequality

$$a^{n}(a^{n}-a^{n-1}) \ge \frac{|a^{n}|^{2}}{2} - \frac{|a^{n-1}|^{2}}{2}.$$

Observe that since the initial conditions are bounded in L^2 , we get

$$I_1 \leq C_3$$
,

for some constant $C_3 > 0$. Regarding the term I_3 , we use Gagliardo-Nieremberg inequality (35) for q = 4, Poincaré inequality and Young inequality to deduce for i = 1, ..., 4

$$\begin{split} \sum_{n=1}^{k} \int_{\Omega_{cyt}} \left| (c_{h}^{n})^{2} b_{i,h}^{n} \right| &\leq \sum_{n=1}^{k} \left(\| c_{h}^{n} \|_{L^{2}(\Omega_{cyt})} \| c_{h}^{n} \|_{L^{4}(\Omega_{cyt})} \left\| b_{i,h}^{n} \right\|_{L^{4}(\Omega_{cyt})} \right) \\ &\leq G \sum_{n=1}^{k} \left(\| c_{h}^{n} \|_{L^{2}(\Omega_{cyt})}^{5/4} \| c_{h}^{n} \|_{H^{1}(\Omega_{cyt})}^{3/4} \left\| b_{i,h}^{n} \right\|_{L^{2}(\Omega_{cyt})}^{1/4} \left\| b_{i,h}^{n} \right\|_{H^{1}(\Omega_{cyt})}^{3/4} \right) \\ &\leq \frac{D^{\min}}{2} \sum_{n=1}^{k} \left\| c_{h}^{n} \right\|_{H^{1}(\Omega_{cyt})} \left\| b_{i,h}^{n} \right\|_{H^{1}(\Omega_{cyt})} + C_{4} \sum_{n=1}^{k} \left\| c_{h}^{n} \right\|_{L^{2}(\Omega_{cyt})}^{5/4} \left\| b_{i,h}^{n} \right\|_{L^{2}(\Omega_{cyt})} \\ &\leq \frac{D^{\min}}{4} \sum_{n=1}^{k} \left(\left\| c_{h}^{n} \right\|_{H^{1}(\Omega_{cyt})}^{2} + \left\| b_{i,h}^{n} \right\|_{H^{1}(\Omega_{cyt})}^{2} \right) \\ &+ C_{5} \sum_{n=1}^{k} \left(\left\| c_{h}^{n} \right\|_{L^{2}(\Omega_{cyt})}^{6} + \left\| b_{i,h}^{n} \right\|_{L^{2}(\Omega_{cyt})}^{6} \right), \end{split}$$

for some constants $C_4, C_5 > 0$. Similarly, we get for for i = 1, ..., 4

$$\begin{split} \sum_{n=1}^{k} \int_{\Omega_{cyt}} \left| c_h^n (b_{i,h}^n)^2 \right| &\leq \frac{D^{\min}}{4} \sum_{n=1}^{k} \left(\| c_h^n \|_{H^1(\Omega_{cyt})}^2 + \left\| b_{i,h}^n \right\|_{H^1(\Omega_{cyt})}^2 \right) \\ &+ C_6 \sum_{n=1}^{k} \left(\| c_h^n \|_{L^2(\Omega_{cyt})}^6 + \left\| b_{i,h}^n \right\|_{L^2(\Omega_{cyt})}^6 \right), \end{split}$$

$$\sum_{n=1}^{k} \int_{\Omega_{sr}} \left| (s_{h}^{n})^{2} b_{5,h}^{n} \right| \leq \frac{D^{\min}}{4} \sum_{n=1}^{k} \left(\left\| s_{h}^{n} \right\|_{H^{1}(\Omega_{sr})}^{2} + \left\| b_{5,h}^{n} \right\|_{H^{1}(\Omega_{sr})}^{2} \right) + C_{7} \sum_{n=1}^{k} \left(\left\| s_{h}^{n} \right\|_{L^{2}(\Omega_{sr})}^{6} + \left\| b_{5,h}^{n} \right\|_{L^{2}(\Omega_{sr})}^{6} \right),$$

and

$$\sum_{n=1}^{k} \int_{\Omega_{cyt}} \left| s_h^n (b_{5,h}^n)^2 \right| \leq \frac{D^{\min}}{4} \sum_{n=1}^{k} \left(\left\| s_h^n \right\|_{H^1(\Omega_{sr})}^2 + \left\| b_{5,h}^n \right\|_{H^1(\Omega_{sr})}^2 \right) + C_8 \sum_{n=1}^{k} \left(\left\| s_h^n \right\|_{L^2(\Omega_{sr})}^6 + \left\| b_{5,h}^n \right\|_{L^2(\Omega_{sr})}^6 \right),$$

for some constants $C_6, C_7, C_8 > 0$. Collecting the previous inequalities, we obtain

$$\left(\int_{\Omega_{cyt}} \left(\left|c_{h}^{k}\right|^{2} + \sum_{i=1}^{4} \left|b_{i,h}^{k}\right|^{2}\right) + \int_{\Omega_{sr}} \left(\left|s_{h}^{k}\right|^{2} + \left|b_{5,h}^{k}\right|^{2}\right)\right) + \frac{D^{\min}}{2} \left(\sum_{n=1}^{k} \int_{\Omega_{cyt}} \left|\nabla c_{h}^{n}\right|^{2} + \sum_{n=1}^{k} \int_{\Omega_{sr}} \left|\nabla s_{h}^{n}\right|^{2}\right) \\
\leq 2C_{3} + 2C_{1} \sum_{n=1}^{k} \left(\int_{\Omega_{cyt}} \left(\left|c_{h}^{n}\right|^{2} + \sum_{i=1}^{4} \left|b_{i,h}^{n}\right|^{2}\right) + \int_{\Omega_{sr}} \left(\left|s_{h}^{n}\right|^{2} + \left|b_{5,h}^{n}\right|^{2}\right)\right) \\
+ C_{9} \left(\sum_{n=1}^{k} \left(\left\|c_{h}^{n}\right\|_{L^{2}(\Omega_{cyt})}^{6} + \sum_{i=1}^{4} \left\|b_{i,h}^{n}\right\|_{L^{2}(\Omega_{cyt})}^{6}\right) + \sum_{n=1}^{4} \left(\left\|s_{h}^{n}\right\|_{L^{2}(\Omega_{sr})}^{6}\right) + \sum_{i=1}^{4} \left\|b_{5,h}^{n}\right\|_{L^{2}(\Omega_{sr})}^{6}\right)\right) \\
+ \sum_{n=1}^{k} \left(\left\|s_{h}^{n}\right\|_{L^{2}(\Omega_{sr})}^{6} + \sum_{i=1}^{4} \left|b_{5,h}^{n}\right|^{2}\right) + \int_{\Omega_{sr}} \left(\left|s_{h}^{n}\right|^{2} + \left|b_{5,h}^{n}\right|^{2}\right)\right) \\
\times \left[1 + \int_{\Omega_{cyt}} \left(\left|c_{h}^{n}\right|^{2} + \sum_{i=1}^{4} \left|b_{i,h}^{n}\right|^{2}\right) + \int_{\Omega_{sr}} \left(\left|s_{h}^{n}\right|^{2} + \left|b_{5,h}^{n}\right|^{2}\right)\right]^{2},$$

for some constants $C_9, C_{10} > 0$. Now we set

$$U := 1 + \int_{\Omega_{cut}} \left(|c_h|^2 + \sum_{i=1}^4 |b_{i,h}|^2 \right) + \int_{\Omega_{sr}} \left(|s_h|^2 + |b_{5,h}|^2 \right).$$

Observe that (37) implies

$$U^k \le C_{11} + C_{10} \sum_{n=1}^k (U^n)^3$$
 for every $1 < k \le N$,

and for some constant $C_{11} > 0$. Therefore by the discrete Gronwall inequality [32] to U, yields

$$\int_{\Omega_{cyt}} \left(|c_h|^2 + \sum_{i=1}^4 |b_{i,h}|^2 \right) + \int_{\Omega_{sr}} \left(|s_h|^2 + |b_{5,h}|^2 \right) \\
\leq U(t) \leq Const(\|(c_0, b_{i,0}, s_0, b_{5,0})\|)_{L^2(\Omega_{cyt}, \mathbb{R}^2) \times L^2(\Omega_{sr}, \mathbb{R}^2)},$$

for i = 1, ..., 4. Therefore, we get the first estimate of (36). Using this and (37) we get the second part of estimate (36). This concludes the proof of Lemma 3.1.

Now, we consider $(\overline{c}_h, \overline{b}_{i,h})$ and $(\overline{s}_h, \overline{b}_{5,h})$ the piecewise affine in t functions in $(W^{1,\infty}([0;T]; V_{cyt,h}))^2$ and in $(W^{1,\infty}([0;T]; V_{sr,h}))^2$, respectively, and interpolating $(c_h^n, b_{i,h}^n)_{n=0,\dots,N} \subset (V_{cyt,h})^2$ and $(s_h^n, b_{5,h}^n)_{n=0,\dots,N} \subset (V_{sr,h})^2$ at the points $(n\Delta t)_{n=0,\dots,N}$ (recall the interpolation scheme $\overline{n}_h = n_h^{n-1} + \frac{t - t_{n-1}}{\Delta t}(n_h^n - n_h^{n-1})$ for $n = c, s, b_i$) for $i = 1, \dots, 4$. Then we have

$$\begin{cases}
\partial_{t}\overline{c}_{h} = \operatorname{div}\left(D_{c}(c_{h})\nabla c_{h}\right) - \sum_{i=1}^{4} R_{i}(c_{h}, b_{i,h}), & \text{in } \Omega_{cyt,T}, \\
\partial_{t}\overline{b}_{i,h} = \operatorname{div}\left(D_{b_{i}}\nabla b_{i,h}\right) + R_{i}(c_{h}, b_{i,h}), & i \in \{1, \dots, 4\}, & \text{in } \Omega_{cyt,T}, \\
\partial_{t}\overline{s}_{h} = \operatorname{div}\left(D_{s}(s_{h})\nabla s_{h}\right) - R_{5}(s_{h}, b_{5,h}), & \text{in } \Omega_{sr,T}, \\
\partial_{t}\overline{b}_{5,h} = \operatorname{div}\left(D_{b_{5}}\nabla b_{5,h}\right) + R_{5}(s_{h}, b_{5,h}), & \text{in } \Omega_{sr,T}.
\end{cases}$$
(38)

In the next lemma, we establish the relative compactness in L^2 of the sequences $(c_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}, s_h, b_{5,h})$, which is achieved by constructing space and time translates and using the a priori estimates given above.

Lemma 3.2. There exist a constant C > 0 depending on initial conditions, Ω_{sr} , Ω_{cyt} and T such that for $i = 1, \ldots, 4$

$$\|(\overline{c}_h, \overline{b}_{i,h})\|_{(L^2(0,T;H^1(\Omega_{cyt})))^2} + \|(\overline{s}_h, \overline{b}_{5,h})\|_{(L^2(0,T;H^1(\Omega_{sr})))^2} \le C, \tag{39}$$

and

$$\left| \int_{0}^{T} \langle \partial_{t} \overline{c}_{h}, \varphi_{c} \rangle_{cyt} dt \right| + \sum_{i=1}^{4} \left| \int_{0}^{T} \langle \partial_{t} \overline{b}_{i,h}, \varphi_{i} \rangle_{cyt} dt \right|$$

$$+ \left| \int_{0}^{T} \langle \partial_{t} \overline{s}_{h}, \varphi_{s} \rangle_{sr} dt \right| + \left| \int_{0}^{T} \langle \partial_{t} \overline{b}_{5,h}, \varphi_{5} \rangle_{sr} dt \right|$$

$$\leq C \left(\|\varphi_{c}\|_{L^{2}(0,T;H^{1}(\Omega_{cyt}))} + \sum_{i=1}^{4} \|\varphi_{i}\|_{L^{2}(0,T;H^{1}(\Omega_{cyt}))}$$

$$+ \|\varphi_{s}\|_{L^{2}(0,T;H^{1}(\Omega_{sr}))} + \|\varphi_{5}\|_{L^{2}(0,T;H^{1}(\Omega_{sr}))} \right),$$

$$(40)$$

for any $\varphi_c, \varphi_i \in L^2(0,T; H^1(\Omega_{cyt}))$ and $\varphi_s, \varphi_5 \in L^2(0,T; H^1(\Omega_{sr}))$.

Proof. First, we begin by proving (39) but this is the consequence of the definition of $(\bar{c}_h, \bar{s}_h, \bar{b}_{i,h})$ (for i = 1, ..., 5) and Lemma 3.1.

The next part of the prove is devoted to (40). Let $\varphi_c \in L^2(0,T;H^1(\Omega))$. Using (38), Lemma 3.1 Gagliardo-Nieremberg inequality, Poincaré inequality and Young inequality, we obtain

$$\left| \int_{0}^{T} \langle \partial_{t} \bar{c}_{h}, \varphi_{c} \rangle_{cyt} dt \right|
= \left| \int_{0}^{T} (D_{c}(c_{h}) \nabla c_{h}, \varphi_{c})_{cyt} dt - \sum_{i=1}^{4} \int_{0}^{T} (R_{i}(c_{h}, b_{i,h}), \varphi_{c})_{cyt} dt \right|
\leq D^{\max} \| \nabla c_{h} \|_{L^{2}(\Omega_{cyt,T})} \| \nabla \varphi_{c} \|_{L^{2}(\Omega_{cyt,T})} + C_{12} \left(\| c_{h} \|_{L^{2}(\Omega_{cyt,T})} \right)
+ \sum_{i=1}^{4} \| b_{i,h} \|_{L^{2}(\Omega_{cyt,T})} \right) \| \varphi_{c} \|_{L^{2}(\Omega_{cyt,T})} + C_{13} \sum_{i=1}^{4} \int_{0}^{T} (|c_{h}, b_{i,h}|, |\varphi_{c}|)_{cyt} dt
\leq C_{14} \| \varphi_{c} \|_{L^{2}(0,T;H^{1}(\Omega_{cyt}))} + C_{13} \sum_{i=1}^{4} I_{i},$$
(41)

for some constants C_{12} , C_{13} , $C_{14} > 0$. Regarding the integral I_i , we use Gagliardo-Nieremberg inequality (35) (for q = 3) and Young inequality to obtain for i = 1, ..., 4

$$I_{i} \leq \int_{0}^{T} \|c_{h}\|_{L^{2}(\Omega_{cyt})} \|b_{i,h}\|_{L^{3}(\Omega_{cyt})} \|\varphi_{c}\|_{L^{6}(\Omega_{cyt})} dt$$

$$\leq G \|c_{h}\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))} \sqrt{\|b_{i,h}\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))}} \int_{0}^{T} \sqrt{\|b_{i,h}\|_{H^{1}(\Omega_{cyt})}} \|\varphi_{c}\|_{L^{6}(\Omega_{cyt})} dt$$

$$\leq G \|c_{h}\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))} \sqrt{\|b_{i,h}\|_{L^{\infty}(0,T;L^{2}(\Omega_{cyt}))}}$$

$$\times \sqrt{\|b_{i,h}\|_{L^{1}(0,T;H^{1}(\Omega_{cyt})}} \|\varphi_{c}\|_{L^{2}(0,T;L^{6}(\Omega_{cyt}))} dt.$$

$$(42)$$

Now we use Lemma 3.1 and the continuous embedding of $L^2(0,T;H^1(\Omega_{cyt}))$ into $L^2(0,T;L^6(\Omega_{cyt}))$ to get from (42)

$$I_i \le C \|\varphi_c\|_{L^2(0,T;H^1(\Omega_{cyt}))},$$

for some constant C > 0, for i = 1, ..., 4. From this and (41) we deduce the bound

$$\|\partial_t \overline{c}_h\|_{L^2(0,T;(H^1(\Omega))')} \le C. \tag{43}$$

Reasoning similarly, we get (40) for $(\bar{b}_{i,h}, \bar{s}_h)$ for i = 1, ..., 5, therefore finishing our proof.

Observe that from the definition of $(\bar{c}_h, \bar{s}_h, \bar{b}_{i,h})$ together with (34) eventually implies that

$$\begin{aligned} \|\bar{c}_h - c_h\|_{L^2(\Omega_{cyt,T})}^2 + \sum_{i=1}^4 \|\bar{b}_{i,h} - b_{i,h}\|_{L^2(\Omega_{cyt,T})}^2 \\ &\leq \sum_{n=1}^N \Delta t \|c_h^n - c_h^{n-1}\|_{L^2(\Omega_{cyt})}^2 + \sum_{i=1}^4 \|b_{i,h}^n - b_{i,h}^{n-1}\|_{L^2(\Omega_{cyt})}^2 \leq \mathcal{C}(\Delta t) & \to 0 \text{ as } \Delta t \to 0, \end{aligned}$$

and

$$\begin{split} \|\overline{s}_h - s_h\|_{L^2(\Omega_{sr,T})}^2 + \|\overline{b}_{5,h} - \overline{b}_{5,h}\|_{L^2(\Omega_{sr,T})}^2 \\ &\leq \sum_{n=1}^N \Delta t \|s_h^n - s_h^{n-1}\|_{L^2(\Omega_{sr})}^2 + \|b_{5,h}^n - b_{5,h}^{n-1}\|_{L^2(\Omega_{sr})}^2 \leq \mathcal{C}(\Delta t) & \to 0 \text{ as } \Delta t \to 0. \end{split}$$

Therefore, from Lemma 3.2 and standard compactness results (see for e.g. [30]), there exists a subsequence of $(c_h, s_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}, b_{5,h})$ not relabeled, such that, as $h \to 0$,

$$\begin{cases}
(c_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}) \to (c, b_1, b_2, b_3, b_4) \text{ strongly and a.e. in } (L^2(\Omega_{cyt,T})^5, \\
(s, b_{5,h}) \to (s, b_5) \text{ strongly and a.e. in } L^2(\Omega_{sr,T})^2, \\
(c_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}) \to (c, b_1, b_2, b_3, b_4) \text{ weakly in } L^2(0, T; H^1(\Omega_{cyt}))^5, \\
(s, b_{5,h}) \to (s, b_5) \text{ weakly in } L^2(0, T; H^1(\Omega_{sr}))^2.
\end{cases} (44)$$

Finally, from the convergence in (44) we can identify the limit $(c, b_1, b_2, b_3, b_4, s, b_5)$ as the weak solution of (16), and the convergence result is summarized as follows.

Theorem 3.3. Assume that condition (17) holds. If $c_0, b_{1,0}, b_{2,0}, b_{3,0}, b_{4,0} \in L^2(\Omega_{cyt})$ and $s_0, b_{5,0} \in L^2(\Omega_{sr})$, then the finite element solution $(c_h, b_{1,h}, b_{2,h}, b_{3,h}, b_{4,h}, s_h, b_{5,h})$, generated by (34), converges along a subsequence to $(c, b_1, b_2, b_3, b_4, s, b_5)$ as $h, \delta t \to 0$, where $(c, b_1, b_2, b_3, b_4, s, b_5)$ is a weak solution of (16) in the sense of Definition 2.1.

4. Numerical simulation

In this section, we present some numerical simulations in several experiments with different parameters. In the first experiment, we investigate the diffusion function to catch the accurate calcium wave front characteristics using imaging observations as reference [7]. In the next experiment, we analyze the influence of the RyR conductance on calcium front wave propagation in cytosolic domain, followed by an investigation on pathological consequences on the cellular scale. Finally, we extend our simulation results in three dimensional framework and we discuss the phenomenological behavior of calcium spark through RyR.

To approximate the solution, we consider the finite element framework defined in Section 3 along with Euler implicit time discretization with time step $\delta t = 10^{-5}s$. The mesh parameters are determined using mesh adaptation to deal with high gradient of the solutions that came from the non uniform distribution of b_4 and b_5 (See Figure 3b below). For calcium initial concentration, we take $c_0 = 0.14$ in Ω_{cyt} and $s_0 = 1.3 \times 10^3$ in Ω_{sr} . For buffers initial concentration in Ω_{cyt} we put $b_{1,0} = 318 \times 10^{-3}$, $b_{2,0} = 471 \times 10^{-3}$, $b_{3,0} = 2.82$, $b_{4,0} = 13.2$ and the initial concentration of CSQN in Ω_{sr} is $b_{5,0} = 24.6 \times 10^3$. The values of previous parameters are provided in μM . We present the diffusion coefficient and interaction parameters (see [11] and the references therein) in Table 1. Almost all parameters have been discovered in cardiac cell medium.

Diffusion coefficient	$\overline{D_c}$	$\overline{D_s}$	D_{b_1}	D_{b_2}	D_{b_3}
Value	220	73.3	140	25	42
Total concentration	B_1	B_2	B_3	B_4	B_5
Value	140	25	42	42	42
Off rates	k_1^{off}	k_2^{off}	k_3^{off}	k_4^{off}	k_5^{off}
Value	45	0.238	0.110	0.0196	0.065
On rates	k_1^{on}	k_2^{on}	k_3^{on}	k_4^{on}	k_5^{on}
Value	255	34	110	32.7	102

Table 1. Total concentration, reaction and diffusion rates in a cardiac cell.

Remark 4. Table 1 shows experimental data related to cardiac cell. The units are presented as follows: The diffusion coefficient (D_l) unit is $10^3 nm^2ms^{-1}$ and the total mass unit (B_i) is μM . The k_i^{on} unit is $10^{-3}ms^{-1}\mu m^{-1}$ and k_i^{off} unit is ms^{-1} . The diffusion of buffers b_4 and b_5 are taken very small.

2D numerical simulation of calcium model

In cell medium, we mention that fixed buffers are not uniformly distributed (see [11] for more details) which would induce a higher gradient of the solution in some regions. Hence, we consider a random distribution of buffers and we adapt the mesh generation (see Figure 3) to achieve a finer approximation in the critical regions. The mesh adaptation is performed using a maximal edge length $h_{\text{max}} = 0.3$ on both computational domain with maximal number of vertexes 120×10^3 (resp. 200×10^3) for cytosolic domain (resp. sarcoplasmic domain). Figure 4 shows the two dimensional Ca^{2+} -distribution for a time instant of t = 1ms after the calcium spark. The concentration dependent diffusion coefficient is given by:

$$D_s(v) = 0.1(1+v) \text{ and } D_c(u) = \overline{D_c}(0.5+u^m)$$

with different values of m (A similar formulation is given in [6]).

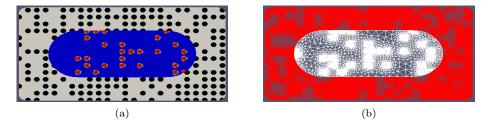


FIGURE 3. The left hand-side figure represents the distribution of b_4 and b_5 . The right hand-side figure shows the Meshing used during the simulation

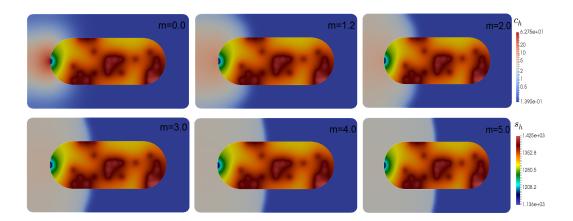


FIGURE 4. The Graphical representation of the approximated solution in Cytosolic domain with non constant diffusion coefficient with m=0,...,5 at $t=50\times 10^{-5}s$.

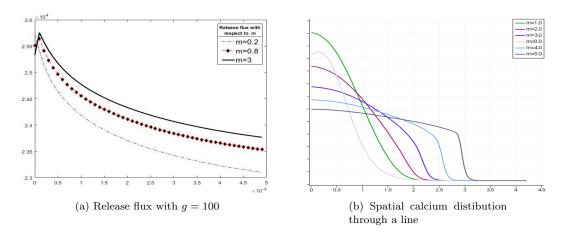


FIGURE 5. Figure 5a shows temporal dynamics of release flux (i.e. $\int_{\Gamma_{ryr}} g(s-c)$) and Figure 5b represents a one dimensional calcium spatial distribution for various values of m=1,...,5

By changing m values from 0 to 5 (see Figure 5b), we observe that Ca^{2+} wavefront displays an ellipsoidal shape with a fast decaying near to the front wave. In imaging results from [7], we observe the same phenomenon due to the heavy fixed buffering in the cell medium that decreases the calcium diffusion in the low concentration areas. We mention also the explosive increasing of release flux at the beginning of calcium spark which has been confirmed experimentally in [34]. Hence, The non constant diffusion coefficient induces the same phenomenon. The larger m is taken more the explosive increasing is important.

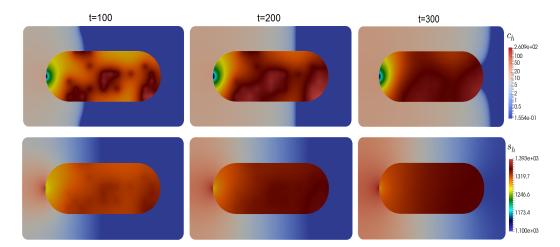


FIGURE 6. The figures on the down side represent the evolution of calcium distribution in the linear case. The figures on the up side devoted to the nonlinear case with m=3

To understand the wavefront differences between linear $(D_c(u) = 1.5\overline{D}_c \text{ and } D_s(v) = \overline{D}_s)$ and nonlinear case (m=3), Figure 6 shows the evolution Ca^{2+} -distribution in both cases. In the second row, we observe a large propagating front wave moving away from the Ca^{2+} highest concentration around RyR (similar observation shown in [11]) because of the linear diffusion with the simple Fickian law. On the upside, the propagating front wave is much smaller and clear in this case. Moreover, this affects the release flux through RyR, see Figure 5a.

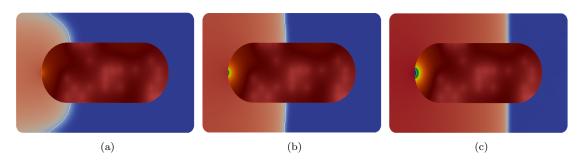


FIGURE 7. The influence of RyR conductance on calcium wave front velocity. The values of RyR conductance g in Figures 7a, 7b and 7c are respectively 50, 150 and 300

Figure 7, shows the influence of RyR conductance on calcium wave front velocity. Figure 7c shows a faster wave front than in Figure 7a case. Pathological behaviors of RyR influences directly the calcium wave velocity and consequently influences the heart beat rate. In other cases some perturbations on heavy buffering protein (CSQN) inside the SR influence the release flux through RyR. In this case, the universal RyR conductance should be corrected to regularize the calcium wave by regularizing the release flux through ionic channels.

3D numerical simulation of calcium model

To fulfill the requirement of the finite element method in three dimensional space we consider a tetrahedral mesh of both domain. Figure 8 shows a section of the external domain mesh (Cytosolic domain) and a meshing of the sarcoplasmic domain. The right figure present the section of Ω_{sr} where the visualization of Ca^{2+} -concentration occur.

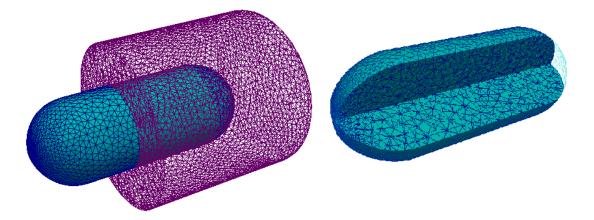


FIGURE 8. In the left hand-side we show complete mesh grid of Ω_{sr} and a section of mesh grid of Ω_{cyt} . In the right hand-side we show the volume section in which 3D simulation will be visualized.

In Figure 9, we present a numerical visualization of calcium solution in both domains. In the sarcoplasmic domain, we show the calcium distribution on the internal sarcoplasmic membrane. We choose to present the calcium dynamics in two different 2D sections with different distance from the ionic channel. To clarify the calcium wave sharpness, we have depict different isosurfaces. Each one describes a specific value of calcium

concentration. The value of isosurfaces equidistantly varying between 0.14 and 0.44 for linear and nonlinear diffusion cases.

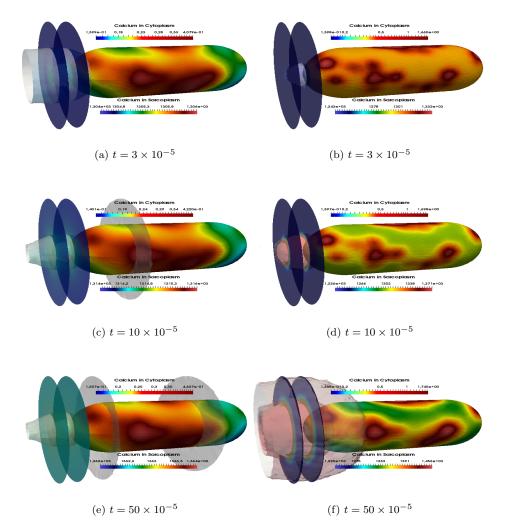


Figure 9. 3D presentation of calcium distribution in both domain.

In Figure 10, we plot the discs shown in Figure 9. The white curves (in Figure 10) present the line curves attached to values of calcium concentrations.

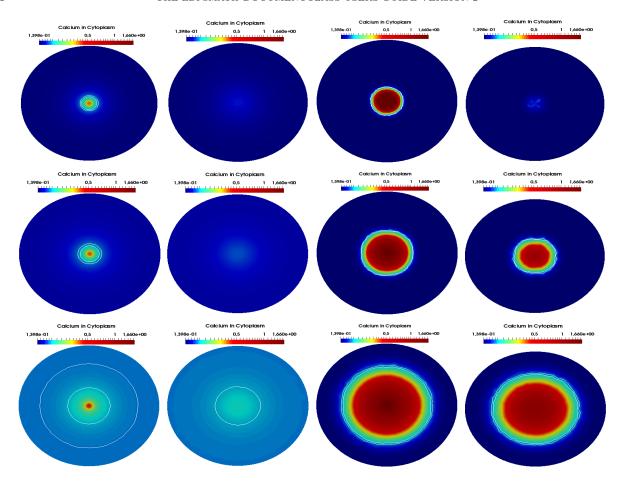


FIGURE 10. The first (resp. second) left column present the linear Ca^{2+} distribution near (resp. far from) of disc to the ionic channel. The third (resp. forth) column presents the nonlinear distribution of Ca^{2+} near (resp. far from) of the ionic channel.

To understand the calcium wavefront differences between the anomalous diffusion model and a linear model, we have computed the calcium distribution in both cases in different time frames. Here, we clearly notice the main difference between the linear diffusion and nonlinear diffusion. In Figure 9, the isosurfaces present a splaying dynamics due to the linear diffusion in the medium comparing with the anomalous diffusion. The spatial dynamics in Figure 10 clarifies the anomalous diffusion between the linear model (see for e.g. [5]) and the proposed model. In the linear model we observe that calcium concentration is much higher near to the ionic channels than further (to the ionic channels) which has been shown in [5]. But this doesn't cover the heterogeneity of the cell medium since the constant diffusion based on the assumption that the calcium flux is constantly proportional to the calcium gradient in the whole domain. In the right two columns of Figure 10, we observe that the calcium front-wave is sharper than the linear case. The choice of an increasing function of calcium concentration may explain the slow penetration of calcium through the cellular heterogeneous media (we refer to [10,29] for more details). The complex dense cell structure prevents normal diffusion in Ca^{2+} low concentration, and the fast diffusion occurs in high concentration region due to high kinetics of Ca^{2+} particles (for more information see [6] and the references therein), which should form a sharp wavefront pattern. This is approved by experimental micro-imaging technics on calcium diffusion in the cardiac myocyte.

Finally, to test the numerical scheme convergence, we consider a simple geometry on $\Omega_{sr} = [-2,0] \times [0,1]$ and $\Omega_{cyt} = [0,4] \times [0,1]$. In space triangularization, we consider a uniform mesh grid of length $h = \frac{1}{N}$ with N varying from 5 to 250, the time step $\delta t = 0.05$ and the final time value T = 2. Table 2 shows errors relative to the exact solutions using P1 and P2 finite element. To simplify, we do not consider the interaction with buffers. We have used as an exact solution (for more information, see [13]) the following function:

$$s_e(t, x, y) = b_1 \exp(-b_2 t) \sin(b_3(x+2)) + b_4 x + b_5 + b_6 \cosh(b_7 x) \cos(b_8 y)$$

$$c_e(t, x, y) = a_1 \exp(-a_2 t) \sin(a_3(x-4)) + a_4 x + a_5 + a_6 \cosh(a_7 x) \cos(a_8 y)$$

The functions $c_e(t, x, y)$ and $s_e(t, x, y)$ satisfies the transmission boundary problem for a specific constant. Let A = 100, C = -10, $k_1 = 2$, $k_2 = 2$, $a_1 = -310$, $a_3 = -0.0072$, $a_4 = -1.97$. The other constants are defined as follows:

$$a_8 = \frac{k_1 \pi}{4}, \ b_8 = \frac{k_1 \pi}{2}, \ a_7 = a_8, \ b_7 = b_8, \ a_6 = \frac{A}{\cosh(4a_7)}, \ a_6 = \frac{C}{\cosh(2b_7)}, \ a_2 = k_s a_3^2, \ b_2 = a_2, \ b_3 = \sqrt{\frac{D_s a_3^2}{D_c}}, \ b_4 = \frac{a_4 D_s}{D_c b_3 \cos(2b_3)} \ \text{and} \ a_5 = s_0 + 4a_4, \ b_5 = c_0 - 2b_4 \ \text{For the conductance value:}$$

$$g(t,y) = \frac{D_s a_1 a_3 \exp(-a_2 t) \cos(4a_3) + a_4}{b_1 \exp(-b_2 t) \sin(-2b_3) + b_5 + b_6 \cos(b_3 y)}$$

After running the test using different parameters. We present in Table 2 different values of the error

$$e_{L^2}^2 = \iint_{\Omega_{cyt,T}} |c_e - c_h|^2 dx dt + \iint_{\Omega_{sr,T}} |s_e - s_h|^2 dx dt$$

and

$$e_{H^1}^2 = \iint_{\Omega_{cyt,T}} |c_e - c_h|^2 \, dx \, dt + \iint_{\Omega_{cyt,T}} |\nabla c_e - \nabla c_h|^2 \, dx \, dt + \iint_{\Omega_{sr,T}} |s_e - s_h|^2 \, dx \, dt + \iint_{\Omega_{sr,T}} |\nabla s_e - \nabla s_h|^2 \, dx \, dt.$$

Hence, we have the following table:

	P_1		P_2	
N	e_{L^2}	e_{H^1}	e_{L^2}	e_{H^1}
20	0.163607	1.20455	0.00106206	0.0910758
40	0.0413642	0.307202	6.92604E-5	0.0121671
60	0.0184233	0.137055	1.3835E-5	0.00368335
100	0.00663965	0.049437	1.80705E-6	0.000809208
160	0.00259459	0.0193244	2.76497E-7	0.000199436
250	0.0010629	0.00791734	5.09595E-8	5.25769E-5

TABLE 2. This table represents an estimate of the error between the exact solution and the approximated solution

Conclusion & Perspective

In this work, we have proposed a nonlinear reaction diffusion model describing the evolution of calcium in cardiac cell. Based on compactness results, we have studied the well posedness of the problem. Then, we have used a finite element scheme along with implicit Euler discretization to approximate the biochemical substance

concentrations. We attached a convergence analysis of our scheme using Kolmogorov compactness results. Moreover, we have produced numerical simulations to clarify the physiological link between the given parameters and calcium behavior under linear and anomalous diffusion. On the experimental scale, B. S. Donahue and R. F. Abercrombie [8] deduced that the calcium diffusion in cytosol depends on calcium concentration. Advanced micro-imaging technology in cardiology provides sophisticated results on calcium diffusion profile. Figure 6 proves encouraging similarities between Ca^{2+} -distribution and imaging results shown in [7]. A precise prediction of spacial distribution (including nonlinear diffusion properties) in cytosol is widely important, it may offer some explanation of spontaneous Ca^{2+} -sparks since passive ionic receptor are very sensitive to local calcium concentration. The linear models [5, 11, 17, 19, 24, 25] has shown some paradoxical results with respect to experiment, our model offers a general framework that may give a step ahead to solve incompatibilities between classical models and experiment. In further work, we intend to simulate the therapeutic strategy by targeting the ionic channel's characteristics and establishing a nonlinear optimal control problem on calcium dynamics.

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