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A STATISTICAL ANALYSIS OF PARTICLE TRAJECTORIES IN LIVING CELLS

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Recent advances in molecular biology and fluorescence microscopy imaging have made possible the inference of the dynamics of single molecules in living cells. Such inference allows to determine the organization and function of the cell. The trajectories of particles in the cells, computed with tracking algorithms, can be modelled with diffusion processes. Three types of diffusion are considered : (i) free diffusion; (ii) subdiffusion or (iii) superdiffusion. The Mean Square Displacement (MSD) is generally used to determine the different types of dynamics of the particles in living cells (Qian, Sheetz and Elson, 1991). We propose here a non-parametric three-decision test as an alternative to the MSD method. The rejection of the null hypothesis – free diffusion – is accompanied by claims of the direction of the alternative (subdiffusion or a superdiffusion). We study the asymptotic behaviour of the test statistic under the null hypothesis, and under parametric alternatives. In addition, we adapt the procedure of Benjamini and Hochberg (2000) to fit with the three-decision test setting, in order to apply the test procedure to a collection of independent trajectories. The performance of our procedure is much better than the MSD method as confirmed by Monte Carlo experiments. The method is demonstrated on real data sets corresponding to protein dynamics observed in fluorescence microscopy.

1. Introduction. A cell is composed of lots of structures in interaction with each other. They continuously exchange biological material, such as proteins, directly via the cytosol or via networks of polymerised filaments namely the microtubules, actin filaments and intermediate filaments. The dynamics of these proteins determine the organization and function of the cell (Bressloff, 2014, chapter 9). The underlying traffic is known to be oriented and it is established that local dynamics of proteins obey to biophysical laws, including subdiffusion (diffusion in a closed domain or in an open but crowded area), free diffusion (or Brownian motion) and superdiffusion (active transport along the microtubules). Then, inference on

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the modes of mobility of molecules is central in cell biology since it reflects the interaction of the structures of the cell. For instance the postsynaptic AMPA-type glutamate receptor (AMPAR) is a protein involved in the fast excitatory synaptic transmission : it plays a crucial part in many aspects of brain functions including learning, memory and cognition. The dynamics of AMPARs determine synaptic transmission : aberrant AMPAR trafficking is implicated in neurodegenerative process, see Henley, Barker and Glebov (2011). Hoze et al. (2012) model their motion with diffusions confined in a potential well. As an other example, Lagache, Dauty and Holcman (2009) modelled the dynamics of a virus invading a cell to infer its mean arrival time to the cell nucleus where it replicates. In their model, the dynamic of the virus alternates between superdiffusion and Brownian motion.

The biophysical literature uses the word diffusion to describe the dynamics of intracellular particles. This word encompasses a great variety of models. First, we have to distinguish diffusion at the macroscopic and microscopic level. The two are closely related in the sense that diffusion at the macroscopic level gives the dynamic of population (or concentration) of particles undergoing microscopic diffusion. In the sequel, we are only interested in microscopic diffusion i-e diffusion of individual trajectories. Secondly, among the variety of mathematical models associated to microscopic diffusion alone, we focus on models that reflects one of the three dynamics : free diffusion, superdiffusion and subdiffusion. Usually, in the biophysical literature, the definition of these dynamics is related to the criterion of the mean square displacement (MSD) (Qian, Sheetz and Elson, 1991). According to the shape of the MSD curve, the literature distinguishes four main types of diffusion (Saxton and Jacobson, 1997; Monnier et al., 2012): superdiffusion, free diffusion, confined diffusion and anomalous diffusion. A particle evolving freely inside the cytosol or along the plasma membrane is modelled by free diffusion. Its motion is due to the constant collisions with smaller particles animated by thermal fluctuations. Then, the particle does not travel along any particular direction and can take a very long time to go to a precise area in the cell. Active intracellular transport can overcome this difficulty so that motion is faster and direct specific. The particles (called in this context cargo) are carried by molecular motors along microtubular filament tracks. Superdiffusions model the motion of molecular motors and their cargo. Subdiffusion, which includes confined diffusion and anomalous diffusion, are the translations of several biological scenarios. Confined or restricted diffusion (Metzler and Klafter, 2000; Hoze et al., 2012) is characteristic of trapped particles : the particle encounters a binding site, then it pauses for a while before dissociating and moving away. Anomalous diffusion includes parti-
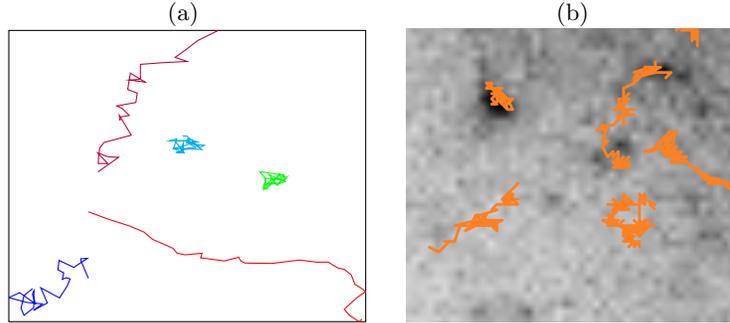
FIG 1. *Representative trajectories from (a) simulated data, (b) a Rab11a protein sequence in a single cell.(Courtesy of UMR 144 CNRS Institut Curie - PICT IBiSA). For the simulated data in Figure (a), the blue trajectory is Brownian, the red one is from a fractional Brownian motion (parameter $\mathfrak{h} > 1/2$), in cyan it is from a Ornstein-Uhlenbeck process and in green from a fractional Brownian motion (parameter $\mathfrak{h} < 1/2$). The parameters of the processes are given in Table (4).*

cles which encounters dynamic or fixed obstacles (Saxton, 1994; Berry and Chaté, 2014), or particles slowed by the contrary current due to the viscoelastic properties of the cytoplasm. In this paper, we will not distinguish confined and anomalous diffusion and consider that both are subdiffusion.

In order to classify the observed trajectories into the three aforementioned types of diffusion *i-e* subdiffusion, superdiffusion or Brownian motion (see Figure 1), a popular statistic is the pathwise Mean Square Displacement (MSD). The simplest rule to classify a trajectory with the MSD is based on the least-squares estimate of the slope β of the log-log plot of the MSD *versus* time (Feder et al., 1996). Didier and Zhang (2015) study the limiting distribution of the pathwise MSD according to the true value of β. Nevertheless, MSD has several limitations :

- The MSD statistic is a summary statistic, and does not suffice to characterize the dynamics of the trajectory. Gal, Lechtman-Goldstein and Weihs (2013) present several other statistics which can be associated to MSD for trajectory analysis. Lund et al. (2014) propose a decision tree for selection motion model combining MSD, Bayesian information criterion and the radius of gyration. Lysy et al. (2016) present a likelihood-based inference as an alternative to MSD for the comparison between two models of subdiffusions : fractional Brownian motion and a generalized Langevin equation. They consider a Bayesian model to estimate the parameter of the diffusion and they use the Bayes factor to compare the models.

- The variance increases with the time lag (see Figure 2) : only the first few points of the MSD may be used to estimate the slope. Moreover the MSD variance is also severely affected at short time lags by dynamic localization error and motion blur. Michalet (2010) details an iterative method, known as the Optimal Least Square Fit (OLSF) for determining the optimal number of points to obtain the best fit to MSD in the presence of localization uncertainty.

- In order to take account of the variance of the MSD estimate, several authors use a set of independent trajectories rather than single trajectories. These trajectories may have different lengths but are assumed to have the same kind of motion. For instance Pisarev et al. (2015) consider weighted-least-square estimate for β by estimating the variance of pathwise MSD. Their motion model selection is then based on the modified Akaike's information criterion. Monnier et al. (2012) propose a Bayesian approach to compute relative probabilities of an arbitrary set of motion models (free, confined, anomalous or directed diffusion). In a general situation, this averaging process can lead to oversimplification and misleading conclusions about the biological process (Gal, Lechtman-Goldstein and Weihs, 2013).

In this paper, we propose a measure that circumvents some limitations of the MSD and which is efficient for classifying single trajectories. In our experiments, the raw data consist in a time series of images obtained by time-lapse fluorescent microscopy. They depict small spots or particles trafficking at the scale of a cell over a fixed time period. Single- and multiple-particle tracking procedures allow us to extract individual molecule and protein trajectories, see Figure 1 (b), which are the data on which we infer. These tracking procedures involve mathematical models and scientific computing to track the single-particle trajectories, see Chenouard et al. (2014); Maroulas et al. (2015); Brunel et al. (2015) and references therein. Our procedure is a three-decision test (Shaffer, 1980). We previously proposed a test with promising results to address this classification problem (Briane, Vimond and Kervrann, 2016). The null-hypothesis is that the observed trajectory is generated from a Brownian motion and the two distinct alternatives are the subdiffusion and superdiffusion. The test statistic T_n is the standardized largest distance covered by the particle from its starting point. We interpret this measure as follows : i/ if the value T_n is low, it means that the process stayed close to its initial position and the particle may be trapped in a small area or hindered by obstacles (subdiffusion); ii/ if the value T_n is high, the particle

went far to its initial position and the particle may be driven by a motor in certain direction (superdiffusion). We propose a theoretical study of the distribution of the test statistic under the null hypothesis, and we assess the power of the test under parametric models illustrating superdiffusion and subdiffusion. At the end, we derive a multiple test procedure in order to apply simultaneously the test on a collection of independent trajectories which are tracked inside the same living cell. This procedure is an adaptation of the procedure of Benjamini and Hochberg (2000).

The present paper is organized as follows. In Section 2, we describe the inference model and provide some examples of subdiffusion and superdiffusion. Our testing procedure is defined in Section 3. In Section 4, we derive a multiple testing procedure for a collection of trajectories. We carry out a simulation study and illustrate the method on real data in Section 5. We focus on the analysis of the Rab11a GTPase protein. This protein is involved in the trafficking of molecules from the endosomes located inside the cell to the cell plasma membrane. The data are TIRF microscopy images depicting the last steps of exocytosis events observed in the region very close the plasma membrane (Schafer et al., 2014). The proofs are postponed to the appendix.

2. Diffusion models for particle trajectories. We observe the successive positions of a single particle in a two-dimensional space at times $t_0, t_1, \ldots, t_n$. We suppose that the lag time between two consecutive observations is a constant Δ. The observed trajectory of the particle is,

$$\mathbb{X}_n = (X_{t_0}, X_{t_1}, \ldots, X_{t_n}),$$

where $X_{t_i} = (X_{t_i}^1, X_{t_i}^2) \in \mathbb{R}^2$ is the position of the particle at time $t_i = t_0 + i\Delta$, $i = 0, \ldots, n$. This discrete trajectory is generated by a stochastic process $(X_t)_{t_0 \leq t \leq t_n}$ with continuous path, solution of the stochastic differential equation (SDE) :

$$(2.1) \qquad dX_t^i = \mu_i(X_t^i)dt + \sigma dB_t^{\mathfrak{h},i}, \quad i = 1, 2.$$

where $B_t^{\mathfrak{h},i}$ are unobserved independent 2D- fractional Brownian motions of unknown Hurst parameter $\mathfrak{h}$, $\sigma > 0$ is the unknown diffusion coefficient and $(\mu_1(x_1), \mu_2(x_2)) : \mathbb{R}^2 \mapsto \mathbb{R}^2$ is the unknown drift term.

ASSUMPTION 1.   *We assume that μ_i fulfils the linear growth hypothesis :*

$$(2.2) \qquad \exists K > 0, \quad \forall x \in \mathbb{R}, \quad |\mu_i(x)| \leq K(1 + |x|),$$

and the Lipschitz condition :

$$(2.3) \qquad \exists M > 0, \quad \forall (x,y) \in \mathbb{R}^2, \quad |\mu_i(x) - \mu_i(y)| \leq M|x - y|.$$

We denote by $\mathcal{L}$ the set of functions verifying Assumption 1. Assumption 1 is sufficient to ensure that SDE (2.1) admits a strong solution (see Nualart and Ouknine (2002) for the case $0 < \mathfrak{h} \leq 1/2$, and Mishura (2008, Chapter 3) for the case $1/2 < \mathfrak{h} < 1$). For a given fractional Brownian motion, we say that (X_t) is a strong solution of the SDE (2.1) if (X_t) verifies (2.1), has continuous paths and that, at time t, X_t depends only on X_{t_0} and on the trajectory of the fractional Brownian motion up to time t. In the following, $P_{\mathfrak{h},\mu,\sigma}$ denotes the measure induced by the stochastic process (X_t) solution of (2.1). This measure comprises all the finite dimensional distributions of the process that is the distribution of the vectors $(X_{t_0}, \ldots, X_{t_n})$, $n \in \mathbb{N}*$ and $t_1 <, \ldots, < t_n$. We also note $\mathcal{P} = \{P_{\mathfrak{h},\mu,\sigma} : 0 < \mathfrak{h} < 1, \mu \in \mathcal{L}, \sigma > 0\}$ the set of solutions of the SDE (2.1).

REMARK 2.1. *In the following, we adopt the large-sample scheme to derive asymptotic properties of our procedure, that is the inter-observation time Δ remains fixed and the number of observations n tends to infinity. In the experimental context of microscopic sequences, Δ is the resolution of the camera while n is the number of frames during which we track the particle. Obviously the resolution of the camera does not change during the experiment. In an ideal situation, we track the particle during an infinite time of observation therefore the number of frames n tends to infinity. Then the large-sample scheme is the most realistic scheme in our context. Other schemes exist (see (Fuchs, 2013, Section 6.1.3)) as the high-frequency scheme for which Δ tends to zero while the duration of observation is fixed. (Fuchs, 2013, Section 6.1.3) emphasizes that the large-sample scheme is the most realistic in real applications while the high-frequency scheme is convenient from a theoretical point of view.*

Heuristically, a SDE models the motion of a particle in a fluid submitted to a deterministic force due to the fluid and a random force due to random collisions with other particles. That is why we model efficiently the motion of intra-cellular particles with these processes. In Equation (2.1), the velocity of the fluid is given by the drift μ while the term $\sigma dB_t^{\mathfrak{h}}$ expresses the random component of the motion due to random collisions.

2.1. *Mean Square Displacement and Modes of Motion.* The mean square displacement is a second order moment defined as :

$$\text{MSD}(t) = \mathbb{E}_{\mathfrak{h},\mu,\sigma}\left(\|X_t - X_{t_0}\|_2^2\right), \tag{2.4}$$

where $\mathbb{E}_{\mathfrak{h},\mu,\sigma}$ is the expectation associated to the measure $P_{\mathfrak{h},\mu,t}$. The empirical mean square displacement of the trajectory $\mathbb{X}_n$ at lag j is given by :

$$\widehat{\text{MSD}}(j\Delta) = \frac{1}{n-j+1}\sum_{k=0}^{n-j}\left\|X_{t_{k+j}} - X_{t_k}\right\|^2. \tag{2.5}$$

According to the shape of the MSD curve, it is common to classify the motions into four classes (rather than into the three groups we presented until now) : Brownian motion (B), anomalous diffusion (A), confined diffusion (C) and directed diffusion (D) (Monnier et al. (2012) Saxton and Jacobson (1997) Pisarev et al. (2015)),

$$\text{MSD}_\text{B}(t) = 2\sigma^2 t, \tag{2.6}$$

$$\text{MSD}_\text{A}(t) = 2\sigma^2 t^\beta, \tag{2.7}$$

$$\text{MSD}_\text{C}(t) = \frac{r_c^2}{a}(1 - be^{-c\sigma^2/(2r_c^2)}), \tag{2.8}$$

$$\text{MSD}_\text{D}(t) = \|v\|^2 t^2 + 2\sigma^2 t, \tag{2.9}$$

where β, a, b, c, r_c are positive constants, and $v \in \mathbb{R}^2$.
In the following subsections, we present diffusion models defined by a SDE (2.1) which have a MSD of the type (A), (B), (C) or (D). We also associate each situation (A), (B), (C) and (D) to one of the three group of diffusions we defined in first place.

Figure 2 illustrates one of the limitation of the MSD : the variance of the estimator (2.5) increases with the lag j. From the MSD model (2.7), Feder et al. (1996) states that the trajectory is subdiffusive if $\beta < 0.9$, superdiffusive if $\beta > 1.1$ and Brownian if $0.9 < \beta < 1.1$. If $\beta < 0.1$ it states that the particle does not move. Figure 2 suggests that the classification of Feder et al. (1996) overdetects subdiffusion and superdiffusion while it is Brownian motion.

2.2. *Free diffusion.* Free diffusion or Brownian motion is the most popular process for describing particle motion suspended in a liquid (Einstein, 1956). It suits particularly well for describing intracellular particle motion as the interior of the cell is mainly made of a fluid called the cytosol. Brownian motion allows dissolved macromolecules to be passively transported without
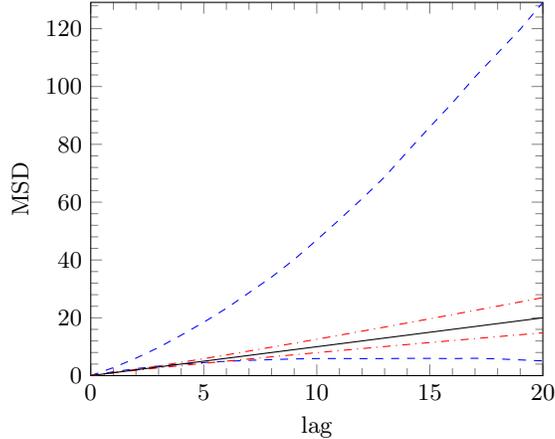
Fig 2. *A classification rule for motion modes from MSD. The dashdotted lines are the bound defined by* Feder et al. *(1996),* $t \to t^\beta$, $\beta = 0.9$ *and* 1.1. *The dashed lines are the pointwise high probability interval of 95% associated to the empirical MSD curve for a standard Brownian motion trajectory of length* $n = 30$. *The bounds of the interval are the 2.5% and 97.5% empirical quantile of* (2.5) *and are computed by Monte Carlo simulation from* 10 001 *Brownian trajectories of size* $n = 30$.

any input of energy. In the SDE (2.1), it matches with the situation where the drift $\mu_i = 0$ and $\mathfrak{h} = 1/2$. In biophysics, it is the process of reference. As a consequence they define the concepts of subdiffusion and superdiffusion from the MSD of Brownian motion (2.6).

2.3. *Subddiffusion.* Anomalous and confined diffusion respectively defined by their MSD (2.7) and (2.8) are both subdiffusions since their MSD is sublinear. There are several ways to model confined diffusion.

On the first hand, we can consider that the domain has non permeable boundaries. Then the motion can be modelled by the SDE (2.1) adding boundary conditions. Equation (2.8) is based on the first two terms of the exact series solution of the MSD of a Brownian particle trapped in a square corral or circular corral defined by the actin cytoskeleton (see Kusumi, Sako and Yamamoto, 1993; Saxton, 1993). The parameter r_c quantifies the area of confinement of the trap while its geometry is described by parameters a, b and c. The diffusion process in this restricted area is called confined diffusion.

On the other hand, we can state that the particle is attracted by an external force modelled by a potential well. We can then use the SDE (2.1) with a specific form for the drift : $\mu_i(x) = -\nabla U_i(x)/\gamma_i$ where $-\nabla U_i$ is the external force of the fluid and γ_i is the frictional coefficient. For instance,

we may consider the Ornstein-Uhlenbeck process :

$$(2.10) \qquad dX_t^i = -\lambda_i(X_t^i - \theta_i)dt + \sigma dB_t^{1/2,i}, \qquad i = 1, 2$$

where $\lambda_i > 0$. Here the particle is assumed to be trapped in a single domain, the potential U is uni-modal and is approximated by a polynomial of order 2 : $U_i(x) = (1/2)k_i(x_i - \theta_i)^2$. The parameter $k_i = \lambda_i\gamma_i$ measures the strength of attraction of the potential (related to the potential depth) while $\theta = (\theta_1, \theta_2)$ is the equilibrium position of the particle. The Ornstein-Uhlenbeck process is a confined diffusion according to the MSD criterion since its MSD is sublinear,

$$(2.11) \qquad MSD(t) = \frac{2\sigma^2(1 - e^{-\lambda t})}{\lambda} \leq 2\sigma^2 t,$$

here it is written in the case $\lambda_i = \lambda$ for simplicity.

Anomalous diffusion can occur for two main reasons. First the particle can bind to an immobile trap that can generate long jump times (Saxton, 1996). In this situation, its motion can be modelled by a continuous time random walk (Metzler and Klafter, 2000). We will not consider this model here as it is not solution of the SDE (2.1). Secondly, the particle can be hindered by mobile or immobile obstacles as the interior environment of cells are crowded with solutes and macromolecules (Bressloff and Newby, 2013). Then, a popular model is the fractional Brownian motion (Jeon et al., 2011). It corresponds to the case $0 < \mathfrak{h} < 1/2$ and $\mu_i = 0$ in (2.1),

$$(2.12) \qquad dX_t^i = \sigma dB_t^{\mathfrak{h},i}, \qquad i = 1, 2.$$

Its MSD is given by (2.7) with $\beta = 2\mathfrak{h} < 1$.

2.4. *Superdiffusion.* At the macroscopic level, the main type of active intracellular transport involves molecular motors which carry particles (cargo) along microtubular filament tracks. The molecular motors and their cargo undergo superdiffusion on a network of microtubules in order to reach a specific area quickly. The molecular motor moves step by step along the microtubules thanks to a mechanicochemical energy transduction process. A single step of the molecular motor is modelled by the so-called Brownian ratchet (Reimann, 2002). When we observe the motion of the molecular motor along a filament on longer time-scales (several steps), its dynamic can be approximated by a Brownian motion with constant drift (also called directed Brownian) (see Peskin and Oster, 1995; Elston, 2000).

The Brownian motion with drift is solution of the SDE :

$$(2.13) \qquad dX_t^i = v_i dt + \sigma dB_t^{1/2,i}, \qquad i = 1, 2,$$

where $v = (v_1, v_2) \in \mathbb{R}^2$ is the constant drift parameter modelling the velocity of the molecular motor. The MSD of the directed Brownian motion is given by (2.9). It is a superdiffusion according to the criterion of MSD.

Superdiffusion can also be modelled by the fractional Brownian motion with Hurst parameter $1/2 < \mathfrak{h} < 1$ since its MSD, as we have already said, is given by (2.7) with $\beta = 2\mathfrak{h} > 1$. However, we note that in the biophysic literature the use of the fractional Brownian motion is mainly related to subdiffusion.

3. A statistical test procedure for a single trajectory. We suppose that the trajectory $\mathbb{X}_n = (X_{t_0}, \ldots, X_{t_n})$ is generated from some unknown diffusion process (X_t) solution of the SDE (2.1). Our procedure allows to test from which type of diffusion the observed trajectory is generated.

We derive two hypothesis test procedures : one for testing H_0 "(X_t) is a free diffusion" versus H_1 "(X_t) is a subdiffusion", the second for testing H_0 "(X_t) is a free diffusion" versus H_2 "(X_t) is a superdiffusion". Then we aggregate the two procedures to build a three-decision procedure.

3.1. *The test statistic.* Let us consider the standardized maximal distance T_n of the process from its starting point :

$$(3.1) \qquad T_n = \frac{D_n}{\sqrt{(t_n - t_0)\hat{\sigma}_n^2}}$$

where D_n is the maximal distance of the process from its starting point,

$$(3.2) \qquad D_n = \max_{i=1,\ldots,n} \|X_{t_i} - X_{t_0}\|_2$$

and $\hat{\sigma}_n$ is a consistent estimator of σ. The choice of $\hat{\sigma}$ is discussed in Section 3.4. If T_n is low, it means the process stays close to its initial position during the period $[t_0, t_n]$: it is likely that it is a subdiffusion. On contrary, if T_n is large, it means the process goes away from its starting point as a superdiffusion does with high probability. This new measure introduces an order in the diffusion processes solution of the SDE (2.1). Then, it allows to classify them into the different classes of diffusion *i-e* free diffusion, superdiffusion and subdiffusion. We want to build a test whose null hypothesis is that the trajectory comes from a Brownian motion, the gold standard process in biophysics. As a consequence T_n must be a pivotal statistic under the hypothesis H_0 that is the trajectory is Brownian.

LEMMA 3.1. *Let $\hat{\sigma}_n$ be a consistent estimator of σ such that the distribution of $\hat{\sigma}_n/\sigma$ does not depend on σ. If (X_t) is a Brownian Motion, the distribution of T_n does not depend on σ.*

Let $q_n(\alpha)$ the quantile of T_n of order $\alpha \in (0,1)$ when (X_t) is a Brownian motion. From Lemma 3.1, $q_n(\alpha)$ does not depend on σ.

3.2. *Two hypothesis test procedures derived from the test statistic.* First we define $\phi_{1,\alpha}$ the hypotheses test associated to H_0 versus H_1 at level $\alpha \in (0,1)$. The procedure $\phi_{1,\alpha}$ is defined through its critical region,

$$(3.3) \qquad \mathcal{R}_{1,\alpha} = \{T_n < q_n(\alpha)\},$$

as the following,

$$\phi_{1,\alpha}(\mathbb{X}_n) = \left\{ \begin{array}{ll} 1 & \text{if } \mathbb{X}_n \in \mathcal{R}_{1,\alpha} \\ 0 & \text{otherwise.} \end{array} \right.$$

Then T_n has probability α to lie in the critical region (3.3). According to Lemma 3.1, the level of the test $\phi_{1,\alpha}$ is α,

$$(3.4) \qquad \sup_{\sigma>0} P_{1/2,0,\sigma}\left(T_n < q_n(\alpha)\right) = \alpha.$$

In a similar way, we can perform the test $\phi_{2,\alpha}$ by replacing subdiffusion by superdiffusion in the alternative hypothesis. The associated critical region is :

$$(3.5) \qquad \mathcal{R}_{2,\alpha} = \{T_n > q_n(1-\alpha)\}.$$

3.3. *A three-decision test procedure.* From the two tests $\phi_{1,\alpha/2}$ and $\phi_{2,\alpha/2}$, we define a new procedure ϕ as follows,

$$(3.6) \qquad \left\{ \begin{array}{l} \text{we decide } H_1 \text{ if } \mathbb{X}_n \in \mathcal{R}_{1,\alpha/2}, \\ \text{we decide } H_2 \text{ if } \mathbb{X}_n \in \mathcal{R}_{2,\alpha/2}, \\ \text{we do not reject } H_0 \text{ otherwise.} \end{array} \right.$$

This procedure is well defined since the intersection of the critical region $\mathcal{R}_{1,\alpha}$ and $\mathcal{R}_{2,\alpha}$ is empty. This procedure is a three-decision test procedure and admits three kinds of errors, see Table 1.

The first kind of errors is to reject the null hypothesis H_0 while H_0 is actually true. The probability that this error occurs is the level of the test which is defined as,

$$(3.7) \qquad \sup_{\sigma>0} \mathbb{E}_{1/2,0,\sigma}\left(\phi_{1,\alpha} + \phi_{2,\alpha}\right) = \alpha.$$

TABLE 1
The three kinds of error in a three-decision test procedure.

		Decision	
Truth	Do not Reject H_0	Decide H_1	Decide H_2
H_0 True	No error	Type I	Type I
H_1 True	Type II	No error	Type III
H_2 True	Type II	Type III	No error

We only control the occurrence of this first kind of error. Then we draw attention that acceptance of H_0 "(X_t) is a free diffusion" does not necessarily demonstrate that H_0 is true. It only means that data does not show any evidence against the null hypothesis. At the end, we reject this assumption in direction to one of the alternatives at level $\alpha/2$.

The second type of errors occurs when we do not reject the null hypothesis while one of the alternatives is true.

The last type of errors is to reject the null hypothesis in favour to a wrong alternative. In the literature of three-decision test such an error is called a Type III error, see for example Rasch (2012) and references therein.

3.4. *Choosing the estimator of σ.* Ideally, we would like to find an estimator of σ which is consistent according to the *large-sample scheme* under the hypotheses H_0, H_1 and H_2, and satisfies the assumption that the distribution of $\hat{\sigma}_n/\sigma$ is free of σ under H_0. However, the *large-sample scheme* is not favourable to get an estimator with such properties. For instance, Florens-Zmirou (1989) shows that the naive maximum likelihood estimator for the drift parameter has an asymptotic bias of the order of lag time Δ. Then, the *high-frequency scheme* and the *rapidly increasing design* turns out to be more convenient to provide consistent estimators. In fact, in the limit, these schemes correspond to the situation in which we have a continuous observation of the process on the time interval of observation. Jiang and Knight (1997) propose non parametric estimators of both the drift and the diffusion coefficient. The consistency of these estimators is proven under the high-frequency scheme only. Therefore, in this section, we discuss about the estimation of the diffusion coefficient under the *large-sample asymptotic*.

The first proposition to estimate σ may be :

$$(3.8) \qquad \hat{\sigma}_{1,n}^2 = \frac{1}{2n\Delta} \sum_{j=1}^{n} \|X_{t_j} - X_{t_{j-1}}\|_2^2$$

Even if the estimator (3.8) is strongly consistent under the *high-frequency scheme* for every process (X_t) solution of (2.1) (Basawa and Prakasa Rao,

1980, Lemma 4.2, p 212), Proposition 1 tells us that it is not the case under the *large-sample scheme*.

PROPOSITION 1.

- *Under H_0, $\hat{\sigma}_{1,n}$ is strongly consistent and the distribution of $\hat{\sigma}_{1,n}/\sigma$ is free of σ.*
- *If (X_t) is an Ornstein-Uhlenbeck process* (2.10), *$\hat{\sigma}_{1,n}^2/\sigma^2$ converges in probability to $(1 - e^{-\lambda\Delta})/(\lambda\Delta)$.*
- *If (X_t) is a Brownian motion with drift* (2.13), *$\hat{\sigma}_{1,n}^2/\sigma^2$ converges almost surely to $\Delta\|v\|_2^2/(2\sigma^2) + 1$.*
- *If (X_t) is a fractional Brownian motion* (2.12), *$\hat{\sigma}_{1,n}^2/\sigma^2$ converges almost surely to $\Delta^{2\mathfrak{h}-1}$.*

A proof of Proposition 1 is given in the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3.1). Proposition 1 states that $\hat{\sigma}_{1,n}$ is adequate to our procedure under the null hypothesis. However $\hat{\sigma}_{1,n}$ is asymptotically biased under some alternatives. Notice that if (X_t) is an Ornstein-Uhlenbeck process (2.10), then $\hat{\sigma}_{1,n}^2$ underestimates σ^2 in average since $(1 - e^{-x})/x < 1$ for $x > 0$. Then T_n might be overvalued with this estimator, increasing Type II or type III error rate in our procedure. If (X_t) is a Brownian motion with drift (2.13), $\hat{\sigma}_1^2$ overestimates σ^2 in average. Then T_n might be overvalued with this estimator, increasing Type II or type III error rate. Similarly, if (X_t) is a fractional Brownian motion (2.12), $\hat{\sigma}_1^2$ underestimates σ^2 if $\mathfrak{h} < 1/2$, and overestimates σ^2 if $\mathfrak{h} < 1/2$.

The second suggestion to estimate σ may be based on the second order differences rather than the first order differences,

$$(3.9) \qquad \hat{\sigma}_{2,n}^2 = \frac{1}{2n\Delta} \sum_{j=1}^{n-1} \|(X_{t_{j+1}} - X_{t_j}) - (X_{t_j} - X_{t_{j-1}})\|_2^2.$$

As $\hat{\sigma}_{1,n}^2$, $\hat{\sigma}_{2,n}^2$ fulfils the assumption of Lemma 3.1 under H_0. This estimator has the advantage of decreasing the bias under some alternatives. For instance it removes the bias in the case of the Brownian motion with drift.

3.5. *Approximation of the distribution of the statistic under the null hypothesis and asymptotic behaviour of our procedure.* Theorem 3.1 gives the asymptotic behaviour of our procedure under the null hypothesis.

THEOREM 3.1.   *Let (X_t) be a Brownian Motion on $\mathbb{R}^2$. Let $\hat{\sigma}_n$ be a consistent estimator of the diffusion parameter σ of (X_t). The test statistic T_n*

converges in distribution to $S_0 = \sup_{0 \le s \le 1} \|W_s\|_2$ as $n \to \infty$. Here (W_t) is a standard 2D Brownian motion that is the Brownian motion of variance $\mathbf{I}_2$ and initialization $W_0 = (0,0)^\top$.

A proof of Proposition 3.1 is given in the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3). The limit distribution of the test statistic under H_0 admits an analytical form (see Borodin and Salminen, 1996, Formulae.1.1.4, p. 280):

$$x \in (0, +\infty) \to \sum_{k=1}^\infty \frac{2e^{-j_{0,k}^2/(2x^2)}}{j_{0,k} J_1(j_{0,k})},$$

where $x \ge 0$, J_ν the Bessel function of order ν and $0 < j_{\nu,1} < j_{\nu,2} < \dots$ the positive zeros of J_ν. Replacing the quantiles $q_n(\alpha)$ by the quantiles of S_0 in our test procedure provides us a test of asymptotic level α.

Furthermore, Proposition 2 gives the asymptotic behaviour of the test statistic under parametric alternatives when the estimator $\hat\sigma_{1,n}$ is considered (see the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3.2) for a proof). More generally, as long as the estimator $\hat\sigma_n$ of the diffusion coefficient is such that $\hat\sigma_n/\sigma$ converges in probability to a positive constant whatever the dynamic of (X_t), then Proposition 2 holds.

PROPOSITION 2. *Assume that we consider the estimator* (3.8) *in our procedure* (3.1).

- *If (X_t) is an Ornstein-Uhlenbeck process* (2.10), *then T_n converges in probability to 0.*
- *If (X_t) is a fractional Brownian motion* (2.12) *with $0 < \mathfrak{h} < 1/2$, then T_n converges in probability to 0.*
- *If (X_t) is a fractional Brownian motion* (2.12) *with $1/2 < \mathfrak{h} < 1$, then T_n converges in probability to $+\infty$.*
- *If (X_t) is a Brownian motion with drift* (2.13), *then T_n converges in probability to $+\infty$.*

Note that Theorem 3.1 and Proposition 2 allow us to control the error rates of type II and type III under parametric alternatives : the associated error rates converges to 0 with n.

However, as in practice n may be small, the asymptotic approximation of the quantiles of T_n may not be accurate. Then the level of the test is no longer α. Since we are able to draw a sample from the distribution of T_n under H_0 (see Algorithm 1), we propose a Monte Carlo estimate of the quantile $q_n(x)$, $0 < x < 1$. This estimate is defined as the $[xN]^{\text{th}}$ order

statistic, $q_n^{(N)}(x)$, of the sample $(T_n^{(1)}, \ldots, T_n^{(N)})$. Table 2 shows that there is a significant difference between asymptotic and non asymptotic quantiles. As expected, as $n \to \infty$, $q_n(\alpha)$ converges to $q(\alpha)$.

Input: n, α, L
`// the length n of the trajectory`
`// the probability` $\alpha \in (0, 1)$
`// the number N of Monte Carlo experiments`
Result: $q_n^{(N)}(\alpha)$.
for $i=1$ **to** N **do**

    `// Simulation of a Brownian trajectory of size` n, `of variance` $\sigma = 1$
    `and with resolution time` $\Delta = 1$.
    initialization $Y_0^{(i)} = (0,0)^\top$;
    for $j=1$ **to** n **do**
        Draw $\epsilon \sim \mathcal{N}(0, \mathbf{I_2})$;
        $Y_j^{(i)} = Y_{j-1}^{(i)} + \epsilon$;
    end
    `// Computation of the test statistic`
    Compute the ratio $T_n^{(i)} = D_n^{(i)}/\hat{\sigma}_n^{(i)}$ from $(Y_0^{(i)}, \ldots, Y_n^{(i)})$;
end

Algorithm 1: Simulation of a N-sample $(T_n^{(1)}, \ldots, T_n^{(N)})$ of the distribution of the statistic T_n under H_0.

In dealing with a test, we can also be interested in computing the p-value. The p-value of the test H_0 *vs* H_1 (subdiffusion as the alternative) is defined as :

$$(3.10) \qquad\qquad p_{1,n} = F_n(T_n),$$

where F_n denotes the cumulative distribution function (cdf) of T_n under H_0. The p-value of the test H_0 *vs* H_2 (superdiffusion as the alternative) is defined as :

$$(3.11) \qquad\qquad p_{2,n} = 1 - F_n(T_n).$$

Testing the hypothesis H_0 *vs* the hypotheses H_1 or H_2 is more tricky as we use a two-sided test with a non-symmetric distribution. In this case we can define the p-value as :

$$(3.12) \qquad\qquad p_n = 2 \min \{p_{1,n}, p_{2,n}\} .$$

Doubling the lowest one-tailed p-value can be seen as a correction for carrying out two one-tailed tests.

TABLE 2

Estimation of the quantiles of order $\alpha/2$ and $1 - \alpha/2$ ($\alpha = 5\%$) for different trajectory lengths n, using algorithm 1 with $N = 1\,000\,001$.

Estimated quantiles	Trajectory size			
quantile order	10	30	100	asymp
2.5%	0.725	0.754	0.785	0.834
97.5%	2.626	2.794	2.873	2.940

We estimate F_n with the standard empirical distribution function estimated by Monte Carlo simulations using Algorithm 1 :

$$(3.13) \qquad \hat{F}_n(x) = N^{-1} \sum_{i=1}^{N} \mathbf{1}(T_n^{(i)} \le x).$$

Then we estimate the p-value (3.12) substituting $\hat{F}_n$ to F_n.

4. Multiple test procedure for a collection of trajectories. Trackers compute a collection of particle trajectories from a sequence of images. Then, it is desirable to decide the modes of mobility for a collection of particle trajectories. From now, we consider a collection $\mathcal{X}_m$ of m trajectories which are simultaneously observed. We denote by $\mathbb{X}_{n_k}^{(k)}$ the observations associated to the kth particle :

$$\mathbb{X}_{n_k}^{(k)} = \left(X_{t_0}^{(k)}, \ldots, X_{t_{n_k}}^{(k)} \right), \qquad k = 1, \ldots, m$$

$$\mathcal{X}_m = \left\{ \mathbb{X}_{n_k}^{(k)}, \ k = 1, \ldots, m. \right\}$$

In this section, we denote by $\mathbb{P}$ the probability distribution of the m-uplet stochastic processes $\left((X_t^{(k)}), \ k = 1 \ldots m \right)$ and by $\mathbb{E}$ its associated expectation. We assume that the observed trajectories are independent, that means $\mathbb{P}$ belongs to the tensorial product of probabilities $\mathcal{P}$, (defined in Section 2) $\mathbb{P} \in \mathcal{P}^{\otimes m}$. For all trajectories $k = 1 \ldots m$, we derive our trichotomy hypothesis test procedure : $H_0^{(k)}$ "$(X_t^{(k)})$ is a free diffusion" versus $H_1^{(k)}$ "$(X_t^{(k)})$ is a subdiffusion" or $H_2^{(k)}$ "$(X_t^{(k)})$ is a superdiffusion". We are faced with the problem of simultaneous tests when the rejections of null hypotheses $H_0^{(k)}$ are accompanied by claims of the direction of the alternative ($H_1^{(k)}$ or $H_2^{(k)}$). In this setup, multiple test procedures are preferable than single test procedures. Indeed, applying the procedure at level α for each trajectory produces in average a number of $m\alpha$ type I errors. A multiple testing procedure aims to control the number of false discoveries. We refer the reader to Shaffer (1995); Roquain (2011); Grandhi (2015) for a review.

TABLE 3

Outcomes in testing m null hypotheses against two-alternatives. For $i = 1, 2$, R_i is the cardinal of $\mathcal{R}_i(\mathcal{X}_m)$. The variables $(S_i)_{i=1\ldots4}, (T_i)_{i=1,2}, U, (V_i)_{i=1,2}$ are not observed and depend on $\mathcal{X}_m$ and P.

True situation	Decision			Total
	Accept H_0	Accept H_1	Accept H_2	
H_0	U	V_1	V_2	$m_0(\mathbb{P})$
H_1	T_1	S_1	S_3	$m_1(\mathbb{P})$
H_2	T_2	S_4	S_2	$m_2(\mathbb{P})$
Total	$m - R_1 - R_2$	R_1	R_2	m

A multiple testing procedure of m null hypotheses against two alternative hypotheses is a rule $\mathcal{R}_1(\mathcal{X}_m) \times \mathcal{R}_2(\mathcal{X}_m)$, where $\mathcal{R}_1(\mathcal{X}_m)$ and $\mathcal{R}_2(\mathcal{X}_m)$ are disjoint subsets of $\{H_0^{(1)}, \ldots H_0^{(m)}\}$. For $i = 1, 2$, $\mathcal{R}_i(\mathcal{X}_m)$ is the set of the rejected hypotheses $H_0^{(k)}$ to the benefit of the alternative $H_i^{(k)}$.

We may commit three kinds of errors in such a multiple testing procedure. Let us introduce the following notations before listing these errors. For a given $\mathbb{P} \in \mathcal{P}^{\otimes m}$, we denote by $\mathcal{I}(\mathbb{P})$ the subset of indexes $\{1, \ldots m\}$ for which the hypothesis $(H_0^{(k)})$ is actually true and by $m_0(\mathbb{P})$ the unknown cardinal of the set $\mathcal{I}(\mathbb{P})$. We denote by $R = R_1 + R_2$ the observed number of null hypotheses which are rejected by the multiple testing procedure. Table 3 summaries the number of errors which may occur following a multiple testing procedure.

- We make a type I error on $H_0^{(k)}$ when we reject $H_0^{(k)}$ while it is a true null hypothesis. In this case, k belongs to the set $\mathcal{I}(\mathbb{P}) \cap (\mathcal{R}_1(\mathcal{X}_m) \cup \mathcal{R}_2(\mathcal{X}_m))$. The number of errors of first kind is $V = V_1 + V_2$.
- Type II error occurs when we do not reject a null hypothesis $H_{0,k}$ while $H_{0,k}$ is false ($k \notin \mathcal{I}(\mathbb{P})$). The number of errors of second kind is $T = T_1 + T_2$.
- The type III errors are directional errors : the index $k \notin \mathcal{I}(\mathbb{P})$ is correctly rejected ($k \in \mathcal{R}_1(\mathcal{X}_m) \cup \mathcal{R}_2(\mathcal{X}_m)$), but for the wrong alternative. We mix up the alternatives deciding one while it is the other. The number of errors of third kind is $S = S_3 + S_4$.

To measure the type I error rate, it is common to consider the k-family-wise error rate (k-FWER) or the false discovery rate (FDR), see Roquain (2011) and references therein. In our settings, controlling the type I error rate is a first step, but it would be necessary to control type III errors as well. In the literature, the sum of the number of errors of first and third kind is controlled using the mixed-directional-family-wise error rate (md-FWER) or the mixed-directional-false discovery rate (mdFDR), see Grandhi

(2015). To our knowledge, the mdFWER and mdFDR are only controlled
for the problem of testing null hypotheses against two-sided alternatives for
finite-dimensional parameters, see for example Guo and Romano (2015) and
references therein.

Biologists are interested in the proportions of each dynamic (subdiffusion,
superdiffusion and Brownian motion) and their geographic location in the
cell. In this context, controlling the FWER, that is the probability to make
a single false discovery, is not relevant. That is why we focus on a proce-
dure which enables to control the FDR. (Guo and Romano, 2015, Section
5) also present several multiple test procedures associated to three-decision
problems which aim to control the FDR. Their approach is different since
the problem is rewritten as a problem which carries out $3m$ null hypothe-
ses. Their proposed procedures control strongly the FDR only on $2m$ null
hypotheses among the $3m$ under the dependence or independence of the
test statistics. In this section, we propose to adapt the multiple testing pro-
cedures of Benjamini and Hochberg (1995) and Benjamini and Hochberg
(2000) controlling the FDR that is the average proportion of false discover-
ies among the discoveries. We stress that our model is non-parametric. Then
we will consider the control of the mdFDR or mdFWER for a next issue.

Let $p^{(k)}, p_1^{(k)}$, and $p_2^{(k)}$ be respectively the p-value (3.12), (3.10) and (3.11)
associated to the kth trajectory, $k = 1 \ldots m$. Let $p^{(1:m)} \leq p^{(2:m)} \leq \ldots \leq$
$p^{(m:m)}$ be the ordered p-values, and $H_0^{(1:m)}, \ldots H_0^{(m:m)}$ the associated null
hypotheses.

The adaptation of the Benjamini-Hochberg (BH) procedure is described
in Procedure 1.

PROCEDURE 1 (Adaptation of the Benjamini-Hochberg (BH) procedure).

1. *Use the Benjamini-Hochberg procedure on the p-values $(p^{(k)})_{k=1\ldots m}$:*
    Let $k^\star$ be the largest k for which $p^{(k:m)} \leq \frac{k}{m}\alpha$.
    $\mathcal{R}_\alpha(\mathcal{X}_m)$ is the set of all hypotheses $H^{(k:m)}$ for $k = 1, \ldots, k^\star$.

2. *Let $\mathcal{R}_{1,\alpha}(\mathcal{X}_m)$ be the subset $\mathcal{R}_\alpha(\mathcal{X}_m)$ such that $p_1^{(k)} < p_2^{(k)}$.*

3. *Let $\mathcal{R}_{2,\alpha}(\mathcal{X}_m)$ be the subset $\mathcal{R}_\alpha(\mathcal{X}_m)$ such that $p_1^{(k)} > p_2^{(k)}$.*

The set $\mathcal{R}_\alpha(\mathcal{X}_m)$ is the set of all rejected null hypotheses for our tri-

chotomy test. According to Finner and Roters (2001), we have,

$$\forall \mathbb{P} \in \mathcal{P}^{\otimes m}, \quad \text{FDR}(\mathcal{R}_\alpha(\mathcal{X}_m), \mathbb{P}) = \mathbb{E}\left(\frac{V}{\max(R, 1)}\right)$$
$$= \frac{m_0(\mathbb{P})}{m}\alpha.$$

Then the FDR of Procedure 1 is controlled by α. Moreover the p-values $p_1^{(k)}$ and $p_2^{(k)}$ give the information to which side of the distribution F_{n_k} the associated test statistic $T_{n_k}^{(k)}$ is. The case of equality $(p_1^{(k)} = p_2^{(k)} = 1/2)$ never occurs since such null hypothesis will not be rejected at the step 1 of the Procedure 1.

Actually, we may also use the adaptive BH procedure of Benjamini and Hochberg (2000) as the first step of Procedure 1. Then the Procedure 1 will be referred to as the adaptive (respectively standard) Procedure 1 when we use the adaptive (respectively standard) BH procedure as the first step. The adaptive BH procedure is more powerful than the standard BH procedure. It uses an estimation of the number of true null hypotheses $m_0(\mathbb{P})$ to increase the power of the BH procedure. Benjamini and Hochberg (2000) simply define the adaptive BH procedure by replacing m by an estimator $\hat{m}_0$ of m_0 in the BH procedure. The associated FDR is $(m_0/\hat{m}_0)\alpha$ and is less than α if $\hat{m}_0 \leq m_0$ almost surely. The estimator $\hat{m}_0$ is based on the plot of the ordered p-values $p^{(i:m)}$ *versus* i. The ordered p-values corresponding to the true null hypothesis are on the right side of the plot. There exists an index i such that the points $(i, p^{(i:m)}), (i + 1, p^{(i+1:m)}), \ldots, (m, p^{(m:m)})$ can be fit by a straight line with slope $\kappa = 1/(m_0 + 1)$. Then they use a method to determine i and estimate m_0 from the estimate of κ. The procedure to estimate m_0 presented in Benjamini and Hochberg (2000) is made for $\hat{m}_0$ to be upward biased. This bias favours the control of the FDR at level α. Due to the fact that $\hat{m}_0$ does not fulfil the condition $\hat{m}_0 \leq m_0$ almost surely, we can not say that the adaptive BH procedure controls the FDR at level α theoretically. However simulations from Benjamini and Hochberg (2000) suggest that the adaptive BH procedure controls the FDR at level α.

5. Simulation study and real data applications. We assess the power of our single test procedure (on a single trajectory) and our multiple test procedure (on a collection of trajectories) by Monte Carlo simulations. We consider parametric alternatives : the Ornstein-Uhlenbeck (2.10) and the fractional Brownian motion $0 < \mathfrak{h} < 1/2$ with Hurst index for subdiffusion processes (H_1); the Brownian motion with drift (2.13) and the fractional Brownian motion with Hurst index $1/2 < \mathfrak{h} < 1$ for superdiffusion processes

(H_2). At the end, we apply our procedure on real data comparing our results with those obtained thanks to a method based on the mean square displacement (MSD).

5.1. *Power of the test procedure for a single trajectory.* In Section 3, we study the asymptotic distribution of the test statistic under the null hypothesis and parametric alternative hypotheses. More precisely Proposition 2 states that the power of the test under parametric alternatives converges to 1 with n. Figure 3 shows the Monte Carlo estimates of the power under the parametric alternatives aforementioned in Proposition 2. For a fixed step of time Δ and a fixed diffusion coefficient σ, we vary the values of the other parameters and the length n of the trajectories. For each parametric alternatives of Proposition 2, we can use exact simulation schemes.

If (X_t) is an Ornstein-Uhlenbeck process (2.10) which is entered in its stationary regime, then the distribution of the test statistic does not depend on θ (see the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3.3)). Figure 3(b) shows the plot of the power regarding the values of λ which models the strength of the restoring force toward the equilibrium position θ. Stronger is the force, more powerful is the test.

Furthermore if (X_t) is a Brownian motion with drift with parameters (v, σ) such that $\|v\| \sqrt{\Delta} > \sigma$, then the particle goes toward the direction of v while the Brownian random part of the SDE (2.13) does not affect much its trajectory (see the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3.3)). The bigger is the norm of the drift parameter v, more powerful is the test, see Figure 3(a).

Finally if (X_t) is a fractional Brownian motion, then the distribution of T_n depends only on the Hurst index $\mathfrak{h}$ (see the supplementary material (Briane, Kervrann and Vimond, 2017, Section 3.3)). Then the test procedure is equivalent to test the null hypothesis "$\mathfrak{h} = 1/2$" versus "$\mathfrak{h} \neq 1/2$", see Figure 3(c).

5.2. *The Average Power and the mdFDR of the multiple test procedure for a collection of trajectories.* The simulation settings are described as follows. According to experience, we choose the number of trajectories to be $m = 100$ or $m = 200$. All trajectories are assumed to have the same size $n = 30$, since this size is reasonable regarding real data. The diffusion coefficient σ and the lag-time Δ are set to 1. The collection of trajectories $\mathcal{X}_m$ is composed of :

- $m_0 < m$ Brownian trajectories (H_0);
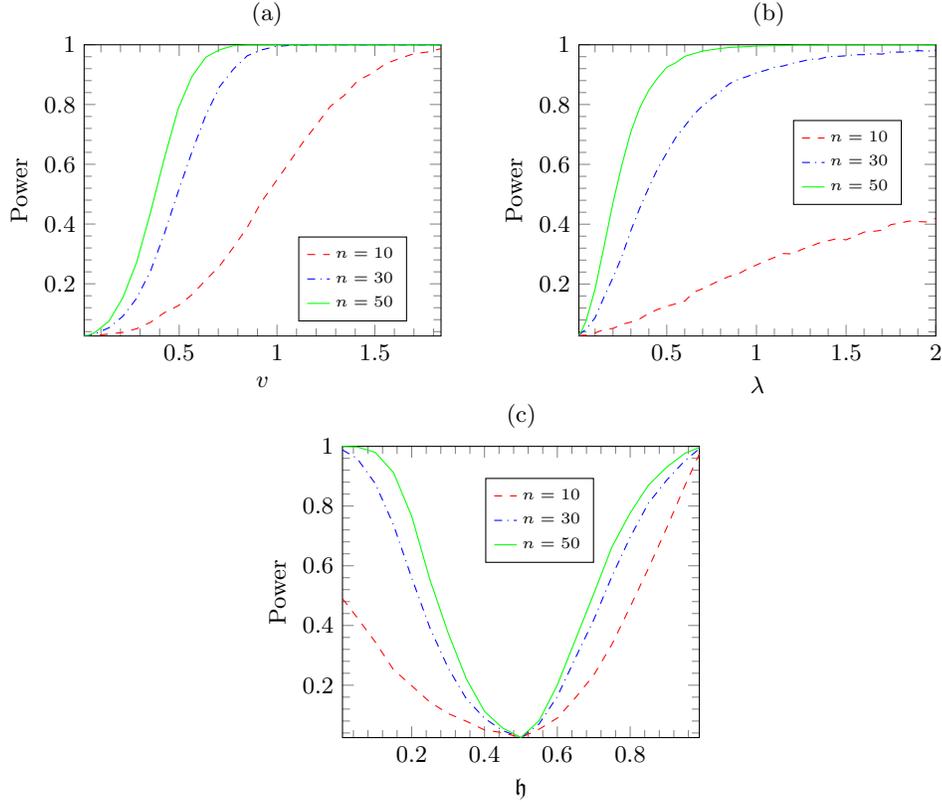- $(m - m_0)/2$ subdiffusive trajectories (H_1), half from an Ornstein-

FIG 3. *Monte Carlo estimate of the power of the test at level $\alpha = 0.05$ according to the trajectory length n and the parameter associated to the following parametric alternatives : (a) Brownian motion with drift (parameter $v = (v_1, v_2)$ such that $v_1 = v_2$); (b) the Ornstein-Uhlenbeck process (parameter λ) and (c) fractional Brownian motion (parameter $\mathfrak{h}$). We use $10\,001$ Monte Carlo replications for computing each point of the power curves.*

Uhlenbeck process with parameter $\lambda > 0$, half from a fractional Brownian motion with Hurst index $0 < \mathfrak{h} < 1/2$;

• $(m - m_0)/2$ superdiffusive trajectories (H_2), half from a Brownian motion with drift $v \in \mathbb{R}^2$, half from a fractional Brownian motion with Hurst index $1/2 < \mathfrak{h} < 1$.

The parameters to simulate these trajectories are given in Table 4. We take the parameters corresponding to a power of the single test procedure of 80%. Such parameters are used to produce Figure 1 (a). This choice seems coherent in regards to trajectories from real data, see Figure 1 (b). For a given m, the proportion of true null hypotheses H_0 varies : $m_0/m \in$

Parameters used for simulating the alternative hypotheses in the simulation for multiple testing. These choices of parameters correspond to a power of 80% on the power curve of the single test procedure. For simplicity we took $\sigma = 1$ for all processes (including Brownian motion). We choose $\Delta = 1$.

Hypothesis	Process	Parameter	Value
H_1	Ornstein-Uhlenbeck	λ	0.53
H_1	Fractional Brownian	$\mathfrak{h}$	0.13
H_2	Brownian motion with drift	$\|v\|$	0.66
H_2	Fractional Brownian	$\mathfrak{h}$	0.85

TABLE 5
Monte Carlo estimate of the FDR and mdFDR for both standard and adaptive Procedure 1 at level $\alpha = 0.05$. The number of replications is $10\,001$. The error rate estimations are expressed in percentages.

m	m_0/m	Standard		Adaptive	
		FDR	mdFDR	FDR	mdFDR
100	0	0	0	0	0.2
	0.2	1	1	3.7	3.7
	0.4	2.1	2.1	4.2	4.2
	0.6	3.2	3.2	4.7	4.7
	0.8	4.1	4.1	4.8	4.8
200	0	0	0	0	0.4
	0.2	1	1	3.4	3.4
	0.4	2.1	2.1	4	4
	0.6	3.2	3.2	4.6	4.6
	0.8	4	4	4.7	4.7

$\{0, 0.2, 0.4, 0.6, 0.8\}$.

The mdFDR is a rate which controls the error of type I and type III. It is defined as $\mathbb{E}((V + S)/\max(R, 1))$ (see Table 3). Table 5 shows that the Procedure 1 also controls the mdFDR. The mdFDR and FDR appear to be very close meaning that the number of type III errors is extremely low. Furthermore, the adaptive Procedure 1 (where m_0 is estimated) is less conservative than the standard Procedure 1. As expected, the FDR and mdFDR increase as the proportion of true null hypotheses increases.

To assess the performance of our multiple test procedure, we use the average power (Grandhi, 2015) :

$$\mathbb{E}\left(\frac{S_i}{m_i}\right), \quad i = 1, 2 \tag{5.1}$$

where m_i is the number of true alternatives H_i and S_i $(i = 1, 2)$ is defined in Table 3. In our simulation scheme, we set $m_i = (m - m_0)/2$. The average power is the expected proportion of hypotheses accepted as H_i among all

true alternatives H_i. Average powers of the different simulations corresponding to different values of m_0/m and m are shown on Figure 4.

First, we can see that the powers of H_1 and H_2 are not very sensitive to the number of hypotheses m for both the standard Procedure 1 and the adaptive Procedure 1. Secondly, the adaptive Procedure 1 is more powerful than the standard Procedure 1 (red and blue dashed lines respectively above red and blue solid lines in Figure 4). The benefit of the adaptive Procedure 1 over the standard Procedure 1 decreases as the proportion of true null hypotheses m_0/m increases (solid and dashed line of same color getting closer as m_0/m increases in Figure 4). This is due to the fact that, as m_0/m tends to 1, m_0 and then $\hat{m}_0$ tend to m. As a result, the adaptive and standard Procedure 1 become similar.

REMARK 5.1. *We observe that, given a certain procedure (standard or adaptive Procedure 1), the average power of H_1 is lower than the average power of H_2 (for our particular choice of parametric models for the alternatives H_1 and H_2), see Figure 4. It is not due to the choice of parameters as both alternatives H_1 and H_2 are simulated to share the same power (80%) with the single test procedure. Actually, it comes from the fact that the p-values under H_2 are stochastically smaller than the p-values under H_1 (see the supplementary material (Briane, Kervrann and Vimond, 2017, Figure 2)). Then, the true superdiffusive trajectories are more easily detected as non Brownian in the first step of the (adaptive) Procedure 1 than the true subdiffusive trajectories. We note that, if we use other parametric models for modelling subdiffusion (H_1) and superdiffusion (H_2), we can have the opposite situation. The main idea is that, as long as the distribution of p-values are not similar under the alternatives H_1 and H_2, one of the two alternative H_1 or H_2 will always be favoured by the (adaptive) Procedure 1.*

Finally, we compare the adaptive Procedure 1 to the MSD classification of Feder et al. (1996), based on a fit of the MSD curve to $t \to t^\beta$, see Section 2.1. We assess the two methods on a single collection of trajectories $\mathcal{X}_m$ with $m = 200$ and $m_0/m = 0.4$, composed of a mixture of Brownian motion, subdiffusion and superdiffusion as described at the beginning of this section. We get the confusion matrices Table 7 and 6 for respectively the adaptive Procedure 1 and the MSD method. As suggested by the limiting curves used by Feder et al. (1996) (see Figure 2), the MSD method mixes up the Brownian trajectories with both subdiffusion and superdiffusion (see line 1 of Table 6). Another big issue is that 40% of the particles undergoing subdiffusion are considered as immobile by the MSD method. On the other hand, the adaptive procedure 1 detects well subdiffusion and superdiffusion
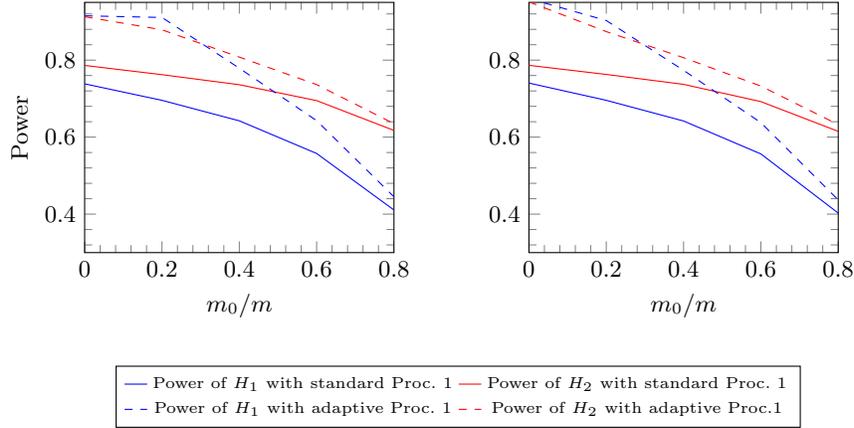
FIG 4. *Monte Carlo estimate of the average power against the proportion of true null hypothesis m_0/m in the collection of hypotheses. On the left we test $m = 100$ hypotheses, on the right $m = 200$.*

TABLE 6
Confusion matrix for the MSD method

Ground truth/Test label	Brownian	Subdiffusion	Superdiffusion	Not moving
Brownian	19	45	36	0
Subdiffusion	0	60	0	40
Superdiffusion	3	0	97	0
Not moving	0	0	0	0

TABLE 7
Confusion matrix for the adaptive Proc.1

Ground truth/Test label	Brownian	Subdiffusion	Superdiffusion
Brownian	96	0	4
Subdiffusion	23	77	0
Superdiffusion	10	0	90

in the setting of this simulation (line 2 and 3 of Table 7). More importantly, it controls the number of false discoveries through the FDR (line 1 of Table 7).

5.3. *Real data : the Rab11a protein sequence.*   Fluorescence imaging and microscopy has a prominent role in life science and medical research. It consists of detecting specific cellular and intracellular objects of interest at the diffraction limit (200 nm). These objects are first tagged with genetically engineered proteins that emit fluorescence. Then, they can be observed using wide field or confocal microscopy. Several image analysis methods have been developed to quantify intracellular trafficking, including object detection and

tracking of fluorescent tags in cells (Chenouard et al. (2014); Kervrann et al. (2016)).

Here, we are particularly interested in studying the exocytosis process, that is the mechanism of active transport of proteins out of the cell. Small structures, called the vesicles, travel from organelles to the cell membrane, propelled by motor activity. The vesicle fuses with the plasma membrane and delivers the transported protein in the extra-cellular medium. Given computed trajectories, we investigate here the quantification of vesicles dynamics and trafficking. As explained earlier in the paper, the trajectories can be generally classified into three categories : Brownian motion, subdiffusion and superdiffusion.

As a model of exocytosis/recycling, we focus on the Rab11a protein. This protein is a member of the dynamic architecture of the complex molecular assembly which regulates recycling organelles trafficking. It plays an essential role in the regulation of late steps of vesicle recycling to the plasma membrane, namely the tethering-docking process (Schafer et al. (2014)). During exocytosis, Rab11a is attached to the vesicle membrane. Then, tracking Rab11a amounts to tracking the vesicle during the exocytosis phase. After the fusion of the vesicle to the cell membrane, Rab11a is recycled in the cytosol. During the recycling step, the tracking of Rab11a is not accurate as the proteins are detached from the vesicle and scatter around the cytosol. It is currently under investigation. For that reason, we focus on the exocytosis process until the fusion time with the cell membrane.

An illustration of the Rab11a sequence is shown in Figure 5 where the dark spots correspond to Rab11a vesicles in a "crossbow" micro-patterned shape cell. A typical image extracted from an image sequence is shown Figure 5. The image sequence is composed of 600 images of size 256×240 (1 pixel=160nm) acquired at 10 frames/s ($\Delta = 0.1s$) [1]. We tracked 1 561 trajectories with the multiple hypothesis tracking method with default parameters (Chenouard et al. (2013)), available on the Icy software (http: www.icy.org). However, we discarded too small and to long trajectories

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[1]Live-cell imaging is performed using simultaneous dual color Total Internal Reflection Fluorescence (TIRF) microscopy. All imaging acquisition was carried out in full conditioned medium at 37°C and 5% CO2 unless otherwise indicated. Simultaneous dual color TIRF microscopy sequences were acquired on a Nikon TE2000 inverted microscope equipped with a x100 TIRF objective (NA=1.49), an Azymuthal TIRF module (Ilas2, Roper Scientifc), an image splitter (DV, Roper Scientific) installed in front of an EMCCD camera (Evolve, Photometrics) that can be bypassed or not, depending on the experimental conditions and a temperature controller. GFP and m-Cherry were excited with a 488 nm and a 561 nm laser, respectively (100mW). The system was driven by the Metamorph software (Molecular Devices).

corresponding to tracking errors in most cases. We used a second molecular marker (Transferin Receptor (TfR)) to select trajectories related to the transport of vesicles until the fusion time. The transmembrane TfR protein is fluorescently labeled with a pH-sensitive probe, the pHluorin. Before the fusion time, the pH inside the vesicle is acidic, leading to a very low pHluorin photon emission. When the vesicle fuses to the plasma membrane, the pHluorin gets exposed to the neutral extracellular medium and the fluorescence suddenly increases in the TIRF image plane. This feature allows us to detect the fusion time and the end of the exocytosis process (Basset et al. (2015)). Now the steps to select the trajectories of Rab11a undergoing exocytosis are the following ones :

1. We simultaneously observe two sets of trajectories : TfR and Rab11a trajectories.
2. We match each trajectory of Rab11a with the corresponding trajectory of TfR.
3. We cut the trajectory of Rab11a at the time when the matched trajectory of TfR starts (fusion time).

There is an additional step of selection of trajectories based on mathematical considerations. As we model particles motions with the diffusion processes 2.1, the particles are expected to move over time. Then, we have to get rid of the particles that do not move enough and consequently, can not be modelled by diffusion processes. In practice, we analyse only the trajectories with at least 20 distinct positions and the vesicles that stop at the same position less than $K = \lfloor n/10 \rfloor$ times (with n the length of the trajectory). In the case of the aforementioned image sequence, we end up with 166 trajectories whose median length is $n = 83$.

In Figure 5, our results show that the four procedures – adaptive Procedure 1, standard Procedure 1, single test and MSD method – do not produce similar classification results visually. From the simulations, we found that the MSD method tends to wrongly over-detect subdiffusion and superdiffusion (see Tables 6 and 7). This is probably true also in the case of real Rab11a sequence. In Table 8, we give the proportion of each type of diffusion for the different methods aforementioned. The adaptive Procedure 1 tends to decrease the number of Brownian trajectories compared to the standard Procedure 1. It is not surprising as the adaptive Procedure 1 is defined to be more powerful than the standard Procedure 1 : it rejects more easily the null hypothesis. This gain in power benefits to the alternative H_1 (subdiffusion). In fact we detect 23% of subdiffusion for the adaptive Procedure 1 against 16% for the standard Procedure 1 while both detect 4% of superdiffusion

TABLE 8

Percentages of Brownian, superdiffusive and subdiffusive trajectories in the Rab11a sequence according to the different methods of classification.

Method	Brownian	Subdiffusion	Superdiffusion
Standard Proc. 1	80	16	4
adaptive Proc. 1	73	23	4
Single test	66	28	6
MSD	16	63	21

(see Table 8). The single test procedure detects even less Brownian motion but we know that it can not control the FDR. In Figure 5, the subdiffusion trajectories labelled with the test approach are more located in the center of the cell in a region corresponding to the Endosomal Recycling Compartment which is known to organize Rab11a carrier vesicles (Schafer et al. (2014)). It is also true for the subdiffusion trajectories labelled with the MSD analysis but we have just said that there is probably an over-detection of the subdiffusion with this method. We note that we carry the classification of trajectories with our different test procedures and the MSD method on multiple sequences of Rab11a protein, see the supplementary material (Briane, Kervrann and Vimond, 2017, Figure 1).

6. Discussion. In this paper, we proposed a method for classifying the particle trajectories observed in living cells into three types of diffusion : Brownian motion, subdiffusion and superdiffusion. We used a test approach with the Brownian motion as the null hypothesis. More specifically, we developed a non-parametric three-decision test whose alternatives are subdiffusion and superdiffusion. On the one hand we built a single test procedure for testing a single trajectory, on the other hand we proposed a multiple test procedure for testing a collection of trajectories. These procedures control respectively the type I error and the false discovery rate at level α. It is worth noting that the length of the trajectory n is taken into account in our classification rule. Our approach can be considered as an alternative to the MSD method. It gives more reliable results as confirmed by our Monte Carlo simulations and evaluations on real sequences of images depicting protein dynamics acquired with TIRF or SPT-PALM microscopy.
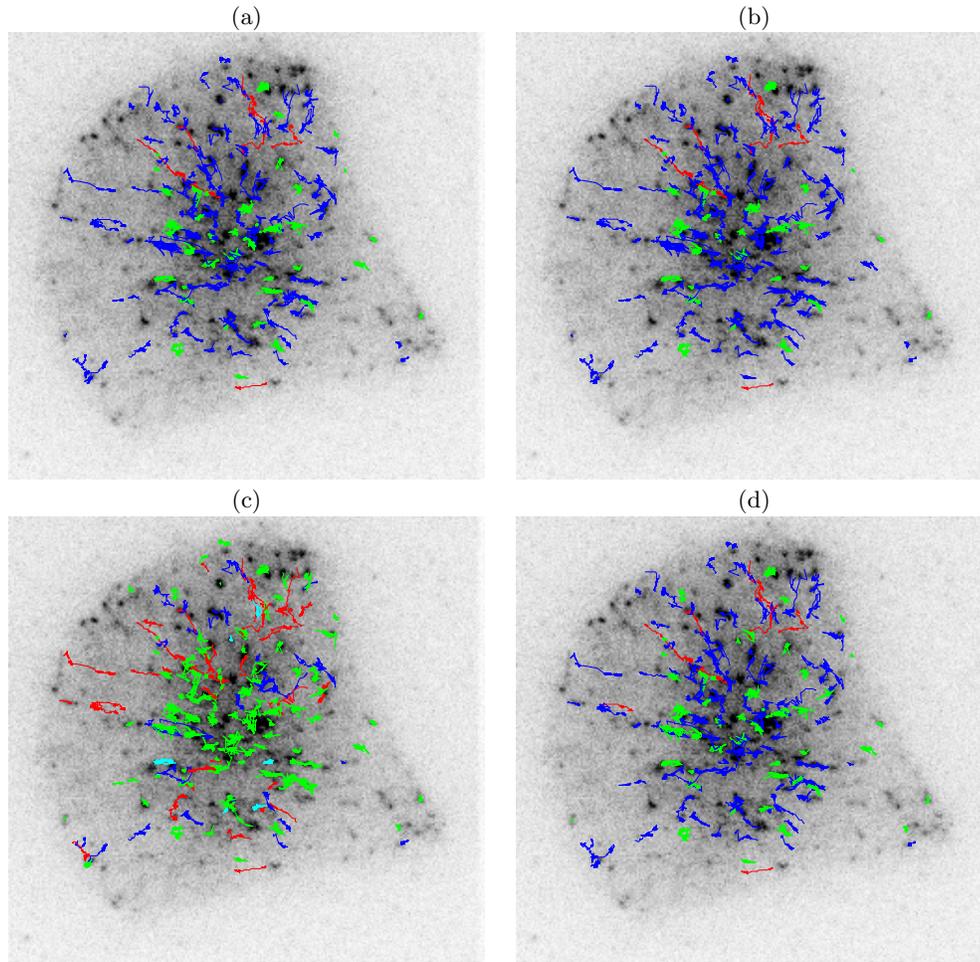
FIG 5. *Map of the classification of the trajectories of the Rab11a sequence with (a) standard multiple test procedure 1, (b) its adaptative version, (c) MSD, (d) single test procedure. The following colour code is used : blue (Brownian motion), red (superdiffusion) and green (subdiffusion).*

SUPPLEMENTARY MATERIAL

Supplement A: Source code
(https://serpico.rennes.inria.fr/doku.php?id=software:thot:index). matlab code source

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