

# Bidirectional interactions between neuronal and hemodynamic responses to transcranial direct current stimulation (tDCS): challenges for brain-state dependent tDCS

Anirban Dutta

# ▶ To cite this version:

Anirban Dutta. Bidirectional interactions between neuronal and hemodynamic responses to transcranial direct current stimulation (tDCS): challenges for brain-state dependent tDCS. Frontiers in Systems Neuroscience, 2015, 9, pp.107-122. 10.3389/fnsys.2015.00107. hal-01203367

# HAL Id: hal-01203367 https://inria.hal.science/hal-01203367

Submitted on 16 Apr 2019  $\,$ 

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

# Bidirectional interactions between neuronal and hemodynamic responses to transcranial direct current stimulation (tDCS): challenges for brain-state dependent tDCS

# 4 Anirban Dutta<sup>1,2\*</sup>

<sup>5</sup> <sup>1</sup>INRIA (Sophia Antipolis) – CNRS : UMR5506 – Université Montpellier, Montpellier, France

6 <sup>2</sup>Laboratoire d'Informatique de Robotique et de Microélectronique de Montpellier (LIRMM), CNRS :

7 UMR5506 – Université Montpellier, Montpellier, France

8

9 \*Correspondence: Anirban Dutta, INRIA - CNRS : UMR5506 - Université Montpellier,

- 10 Montpellier, France
- 11 Email: adutta@ieee.org

# 12 Abstract

Transcranial direct current stimulation (tDCS) has been shown to modulate cortical neural activity. 13 14 During neural activity, the electric currents from excitable membranes of brain tissue superimpose in the extracellular medium and generate a potential at scalp, which is referred as the 15 electroencephalogram (EEG). Respective neural activity (energy demand) has been shown to be 16 closely related, spatially and temporally, to cerebral blood flow (CBF) that supplies glucose (energy 17 supply) via neurovascular coupling. The hemodynamic response can be captured by near-infrared 18 spectroscopy (NIRS), which enables continuous monitoring of cerebral oxygenation and blood 19 volume. This neurovascular coupling phenomenon led to the concept of neurovascular unit (NVU) 20 that consists of the endothelium, glia, neurons, pericytes, and the basal lamina. Here, recent works 21 suggest NVU as an integrated system working in concert using feedback mechanisms to enable 22 23 proper brain homeostasis and function where the challenge remains in capturing these mostly nonlinear spatiotemporal interactions within NVU for brain-state dependent tDCS. In principal 24 accordance, we propose EEG-NIRS-based whole-head monitoring of tDCS-induced neuronal and 25 hemodynamic alterations during tDCS. 26

# 27 Challenges in clinical translation of transcranial brain stimulation - an introduction

Transcranial direct current stimulation (tDCS) - an electrically based intervention directed at the 28 central nervous system level - is a promising tool to alter cortical excitability and facilitate 29 neuroplasticity (Nitsche and Paulus, 2011). However, inter-subject variability and intra-subject 30 reliability currently limits clinical translation (Horvath et al., 2014). Indeed, a recent meta-analysis 31 showed that the treatment effects of transcranial brain stimulation in patients with stroke are rather 32 inconsistent across studies and the evidence for therapeutic efficacy is still uncertain (Raffin and 33 Siebner, 2014). Here, it may be possible to reduce inter-subject variability and improve intra-subject 34 reliability using simultaneous neuroimaging that can objectively quantify the individual brain-state 35 before and during tDCS. Non-invasive neuroimaging techniques that have previously been combined 36 with tDCS include electrophysiological, e.g., EEG (Schestatsky et al., 2013) and haemodynamic, 37 e.g., functional magnetic resonance imaging (fMRI) (Meinzer et al., 2014) and NIRS (McKendrick et 38 al., 2015) approaches. Here, NIRS presents several advantages relative to fMRI, such as 39 measurement of concentration changes in both oxygenated (HbO2) and deoxygenated (HHb) 40 hemoglobin, finer temporal resolution, ease of administration and relative insensitivity to movement 41

42 artefacts. Although fMRI has become the benchmark for in-vivo imaging of the human brain, in practice, NIRS and EEG are more convenient and less expensive technology than fMRI for 43 simultaneous neuroimaging for brain-state dependent tDCS. However, the challenge remains in 44 modeling whole-head spatiotemporal coupling of neuronal and hemodynamic alterations induced by 45 tDCS where such brain-state dependent tDCS needs not only to consider the brain as a dynamical 46 system but also need to consider that its parameters will be inter-individually heterogeneous, 47 dependent on brain injury (and maladaptive plasticity, e.g., reactive gliosis (Buffo et al., 2008)), task 48 49 characteristics (e.g., attention issues) and other factors (Raffin and Siebner, 2014).

# 50 Biophysical models for capturing hemodynamic alterations induced by tDCS

Neural activity has been shown to be closely related, spatially and temporally, to cerebral blood flow 51 (CBF) that supplies glucose via neurovascular coupling (Girouard and Iadecola, 2006). The 52 hemodynamic response to neural activity can be captured by near-infrared spectroscopy (NIRS), 53 which enables continuous monitoring of cerebral oxygenation and blood volume (Siesler et al., 54 2008). The regulation of CBF and its spatiotemporal dynamics may be probed with short-duration 55 anodal tDCS which challenges the system with a vasoactive stimulus in order to observe the system 56 response. Based on prior works (Nitsche and Paulus, 2000)(Dutta et al., 2015), such short-duration 57 (<1min) anodal tDCS is postulated to cause no aftereffects and may be used to probe neurovascular 58 coupling (and NVU) (Jindal et al., 2015b). Here, CBF is increased in brain regions with enhanced 59 neural activity via metabolic coupling mechanisms (Attwell et al., 2010) while cerebral 60 autoregulation mechanisms ensure that the blood flow is maintained during changes of perfusion 61 pressure (Lucas et al., 2010). During such a short-duration anodal tDCS experiment, cerebrovascular 62 reactivity (CVR) can be measured as the change in CBF per unit change in relation to anodal tDCS 63 64 intensity. Moreover, the rate of change of hemodynamic responses to same tDCS intensity may explain inter-individual differences in tDCS after-effects (Han et al., 2014). Also, phenomological 65 model for metabolic coupling mechanisms (Attwell et al., 2010) can be used to capture CVR that 66 represents the capacity of blood vessels to dilate during anodal tDCS due to neuronal activity-related 67 increased demands of oxygen (Dutta et al., 2013). Here, CVR reflects the capacity of blood vessels to 68 69 dilate, and is an important marker for brain vascular reserve (Markus and Cullinane, 2001). Indeed 70 pressure-perfusion-cognition relationships may be monitored with the brain vascular reserve (Novak, 2012) where the CVR distributes CBF toward the brain areas in need of increased perfusion 71 due to enhanced neural activity. 72

73

74 Prior work has shown a significant correlation between tDCS current strength and increase in regional CBF in the on-period relative to the pre-stimulation baseline (Zheng et al., 2011). We 75 investigated regional CVR during anodal tDCS by adapting an arteriolar compliance model of the 76 cerebral blood flow response to a neural stimulus (Behzadi and Liu, 2005). Regional CVR was 77 defined as the coupling between changes in CBF and cerebral metabolic rate of oxygen (CMRO2) 78 during anodal tDCS-induced local brain activation (Leontiev and Buxton, 2007). The complex path 79 from the tDCS-induced change of the synaptic transmembrane current, u(t) (only excitatory effects 80 considered) (Molaee-Ardekani et al., 2013) to a change in the concentration of multiple vasoactive 81 agents (such as NO, potassium ions, adenosine), represented by a single vascular flow-inducing 82 vasoactive signal, <sup>s</sup>, was captured by a first-order Friston's model (Friston et al., 2000). Chander and 83 Chakravarthy (Chander and Chakravarthy, 2012) presented a computational model that studied the 84

85 effect of metabolic feedback on neuronal activity to bridge the gap between measured hemodynamic

response and ongoing neural activity. Here, the NVU (see Figure 1) consists of the endothelium, glia,

87 neurons, pericytes, and the basal lamina that has been proposed to maintain the homeostasis of the

88 brain microenvironment (Iadecola, 2004). In this connection, the role of lactate as a signaling

89 molecule was described recently (Yang et al., 2014), which supports a (delayed) "reverse" influence in the NVU from the vessel back to neuron via lactate (Chander and Chakravarthy, 2012). Recently, a 90 detailed biophysical model of the brain's metabolic interactions was presented by Jolivet et al. 91 (Jolivet et al., 2015). This not only supported the astrocyte-neuron lactate shuttle (ANLS) hypothesis 92 that the lactate produced in astrocytes (a type of glial cell) can also fuel neuronal activity but it also 93 provided a quantitative mathematical description of the metabolic activation in neurons and glial 94 cells, as well as of the macroscopic measurements obtained during brain imaging. Indeed, this model 95 96 captured the pattern of neurovascular responses observed in rodents in response to sustained sensory stimulation where CBF only starts to increase above its baseline ~0.5-1 sec after the onset of 97 stimulation (Jolivet et al., 2015). We also found such onset effects (called "initial dip") of anodal 98 tDCS in stroke patients (Dutta et al., 2015). Moreover, Jolivet et al. (Jolivet et al., 2015) highlighted 99 the neuron-astrocyte cross-talk during oscillations linked to blood oxygenation levels (DiNuzzo et al., 100 2011) where such oscillations also occurred after anodal tDCS-based perturbation of the neuroglial 101 networks in our EEG-NIRS stroke study (Dutta et al., 2015). We therefore postulate that short-102 duration anodal tDCS can be used to perturb neuroglial networks in health and disease to probe the 103 spatiotemporal dynamics of the NVU based on simultaneous EEG-NIRS neuroimaging (Dutta, 2014) 104 (Dutta et al., 2015) and biophysical model (Jolivet et al., 2015) based analysis. 105



**Figure 1:** Illustration of the effects of transcranial direct current stimulation (tDCS) from simultaneous recording of electroencephalogram (EEG) and near infra red spectroscopy (NIRS). The colors on the plot corresponds to the sensors. Here, only one anode and one cathode is highlighted for tDCS, however, local current steering based on NIRS-EEG feedback is possible to optimally orient the electric field with multiple return electrodes (Khadka et al., 2015). Here, NVU consists of the endothelium, glia, neurons, pericytes, and the basal lamina in which neurons, astrocytes, and vessels are semi-independent networks operating in tandem. Neuronal network activity drives functional hyperemia via direct effects on the blood vessels network as well as indirect effects via the astrocytic network. Also, the hemodynamics changes can impact neuronal network activity via direct (diffusible messengers, electromechanical and thermal interactions) and indirect (via astrocytic network) pathways.

106

# 107 Neural mass or field models for capturing neuronal alterations induced by tDCS

During neural activity, the electric currents from excitable membranes of brain tissue superimpose at 108 a given location in the extracellular medium and generate a potential, which is referred to as the EEG 109 (Nunez and Srinivasan, 2006). Here, neural mass models (NMM) can provide insights into the 110 neuromodulatory mechanisms underlying alterations of cortical activity induced via tDCS (Molaee-111 Ardekani et al., 2013). Specifically, the origin of tDCS-induced alterations in the EEG power 112 spectrum was captured using a thalamocortical NMM (Dutta and Nitsche, 2013). The NMM for a 113 single cortical source comprises of 4 neuronal subpopulations, excitatory pyramidal neurons (ePN), 114 excitatory interneurons (eIN), slow inhibitory interneurons (siIN), and fast inhibitory interneurons 115

(fiIN) (Zavaglia et al., 2006). The NMM for the cortical source was coupled with another 116 representing the thalamus (Sotero et al., 2007), which comprises of 2 neuronal subpopulations - an 117 excitatory thalamocortical (eTCN) and an inhibitory reticular-thalamic (iRT). The basis of our 118 cortical NMM is the Friston model (Moran et al., 2007) that emulates the activity of a cortical area 119 using three neuronal subpopulations, ePN, eIN, and siIN. A population of ePN (output) cells receives 120 inputs from inhibitory and excitatory populations of interneurons via intrinsic connections (intrinsic 121 connections are confined to the cortical sheet). An extrinsic thalamo-cortico-thalamic loop consists 122 123 of eTCN and iRT in the thalamic NMM (Ursino et al., 2010). Our lumped thalamo-cortico-thalamic network model can be used to simulate the subject-specific EEG power spectral density changes 124 during/following tDCS (Dutta and Nitsche, 2013) by modifying the model parameters (e.g., average 125 gain of synapses, their time constants) (Zavaglia et al., 2006). We found that anodal tDCS enhances 126 127 activity and excitability of the excitatory pyramidal neuron at a population level in a non-specific 128 manner and mu-rhythm desynchronization is generated (Dutta and Nitsche, 2013). The tDCS effects on the population kinetics depend on the direction of cortical current flow determining the relative 129 influence of acute tDCS on the cellular targets responsible for modulation of synaptic efficacy, which 130 are primarily somata and axon terminals (Rahman et al., 2013). Basal and apical dendrites can be 131 concomitantly polarized in opposite directions, and Laver V pyramidal neurons exhibit the highest 132 measured somatic sensitivities to subthreshold fields (Rahman et al., 2013). Therefore, not all neural 133 tissue will be equally affected by a given stimulation protocol which may distinctly affect neuronal 134 populations/neuronal compartments. Indeed, a recent computational modeling study suggested that 135 tDCS may induce opposing effects on different types of interneurons (Molaee-Ardekani et al., 2013). 136 Here, the excitation versus inhibition effects (Krause et al., 2013) of tDCS on the population kinetics 137 can produce a whole spectrum of EEG signals within the oscillatory regime of a neural mass model 138 (David and Friston, 2003). 139

There are several prior works that have shown both "online" effects of tDCS on EEG with EEG 140 performed during tDCS as well as "offline" effects with EEG performed after tDCS. Here, it is 141 142 important to separate studies where tDCS is applied during a rest state (Ardolino et al., 2005) (Zaehle et al., 2011) (Spitoni et al., 2013) or an active task state (Matsumoto et al., 2010) (Mangia et al., 143 2014). We computationally found (Dutta and Nitsche, 2013) in concordance with the experimental 144 results of Matsumoto et al. (Matsumoto et al., 2010) that tDCS effects on mu-rhythm 145 desynchronization depend on the direction of cortical current flow determining the relative influence 146 of acute tDCS on the cellular targets. Matsumoto et al. (Matsumoto et al., 2010) found that tDCS 147 applied over the left primary motor area for 10min at 1mA with a 35cm<sup>2</sup> electrode influenced event-148 related desynchronization (ERD) during right hand grasping where the mu ERD increased after 149 anodal tDCS and decreased after cathodal tDCS. Here, not only the "local" effects but the "distant" 150 151 effects of tDCS are also relevant where Polanía et al. (Polanía et al., 2012) reported that the functional connectivity patterns significantly increased after anodal tDCS (i.e., "offline" effects) over 152 the primary motor cortex where tDCS modulated functional connectivity of cortico-striatal and 153 thalamo-cortical circuits. Notturno et al. (Notturno et al., 2014) showed spatial diffusion of anodal 154 tDCS (during a motor task) effects where an increment of low alpha band power over the course of 155 pre- and post-stimulation recording sessions was found during motor task that was localized in the 156 sensorimotor and parieto-occipital regions. Indeed, not only the "offline" effects, but changes in 157 functional connectivity patterns may start evolving during tDCS (i.e., "online" effects) as shown by 158 our modeling study (Dutta and Nitsche, 2013). tDCS/EEG co-registration studies have shown that 159 anodal tDCS mostly modulate spontaneous cortical activity in the alpha band where alpha-rhythm 160 states have a significant effect on perceptual learning (Sigala et al., 2014). In fact, more than 60% of 161 the observed inter-subject variability in perceptual learning can be ascribed to ongoing alpha activity 162 where Sigala et al. (Sigala et al., 2014) highlighted the need for multidisciplinary approaches 163

combining assessment of behavior and multi-scale neuronal activity, active modulation of ongoing 164 brain states and computational modeling to reveal the mathematical principles of the complex 165 neuronal interactions. We therefore postulate that concurrent EEG-NIRS-based neuroimaging of the 166 short-duration tDCS-induced modulation can be analyzed by combining a biophysical model (Jolivet 167 et al., 2015) of the NVU with the computational model (neural mass or field model) of multi-scale 168 neuronal activity of the whole brain (Sigala et al., 2014) to capture the spatiotemporal dynamics of 169 the interactions between the neuronal and hemodynamic responses in health and disease. Here, the 170 171 challenges remain in ensuring the observability of the NVU with intelligent placement of EEG-NIRS sensors since presence of symmetry in the nonlinear network of NVU (see Figure 1) may decrease 172 observability (although networks containing only rotational symmetries remain observable) (Whalen 173 et al., 2015). 174

# 175 Bidirectional interactions between neuronal and hemodynamic responses to tDCS - a discussion

In our prior work (Dutta et al., 2015), we found an initial dip in the oxy-haemoglobin concentration 176 and concomitant increase in the mean power spectral density within lower (<12Hz) EEG frequency 177 178 band. It was postulated that the immediate need to fuel neuronal energy recovery was via the lactate shuttle (Pellerin and Magistretti, 1994) where blood glucose supply has a longer delay (Gruetter et 179 al., 1996). A detailed biophysical model of the brain's metabolic interactions by Jolivet et al. (Jolivet 180 et al., 2015) also supported the ANLS hypothesis. Moreover, recent works showed that lactate can 181 modulate the activity of primary cortical neurons through a receptor-mediated pathway (Bozzo et al., 182 183 2013) and vasomotion rhythms can influence neural firing patterns (Nikulin et al., 2014). Also, lactate promotes plasticity gene expression by potentiating NMDA signaling in neurons, and the 184 action of lactate is mediated by the modulation of NMDA receptor activity (Yang et al., 2014). These 185 dynamic ANLS interactions leave us to question its role in tDCS facilitated neuroplasticity and 186 learning (Suzuki et al., 2011). Also, the spatiotemporal dynamics of the millisecond-to-second-range 187 direct (diffusible messengers, electromechanical and thermal interactions) and seconds-to-tens-of-188 seconds-range indirect interaction in the NVU following tDCS, i.e. the hemo-neural hypothesis 189 (Moore and Cao, 2008), may at least partially explain the time course of the induction of homeostatic 190 191 plasticity generated by repeated tDCS of the human motor cortex (Fricke et al., 2011). Fricke and 192 coworkers (Fricke et al., 2011) hypothesized a role of L-type voltage-gated Ca2+ channels (L-VGCC) in short-term homeostatic plasticity, since tDCS has been shown to induce a long-lasting 193 disturbance of Ca2+ homeostasis (Islam et al., 1995) and induce calcium-dependent plasticity 194 (Nitsche et al., 2003). Here, the glial network may have an important role (i.e., spatial buffering) in 195 regulating neural activity by distributing ions (Halnes et al., 2013) in seconds-to-tens-of-seconds-196 range where an influence of long-lasting disturbance of Ca2+ homeostasis via tDCS on the myogenic 197 and the metabolic control of cerebral circulation cannot be excluded. In fact, astrocytes, a sub-type of 198 glia in the central nervous system, can integrate a large number of synapses and can respond to 199 neuronal activity via neurotransmitter-evoked activation of astrocytic receptors (Araque et al., 2001). 200 Indeed, neuronal activity can mobilize internal calcium in astrocytes and the calcium wave in 201 different spatial-temporal dimensions can result in a higher level of brain integration (Volterra et al., 202 2014) where the evidence for tDCS-induced large scale changes in brain synchronization and 203 topological functional organization has been shown after acute stimulation (Polanía et al., 2011). 204

205

Based on these prior works, we recently proposed EEG-NIRS-based monitoring of neurovascular coupling functionality under perturbation with tDCS (Jindal et al., 2015b). Here, neuronal and hemodynamic responses measured with EEG-NIRS neuroimaging can be represented abstractly as the system response of the NVU to tDCS perturbation (see Figure 1) where presence of symmetry in

the nonlinear network of NVU (see Figure 1) may decrease observability (Whalen et al., 2015). Since 210 no real-world network has exact symmetries so with intelligent placement of EEG-NIRS sensors (e.g. 211 to avoid systemic interference (Sood et al., 2015)) along with system identification and parameter 212 estimation techniques, it may be possible to track the spatiotemporal change of the states of the NVU. 213 This observer model can then be used to drive multi-electrode tDCS (Dmochowski et al., 2011) for 214 active spatiotemporal modulation of the brain states (e.g., posterior alpha-rhythm). Here, we base our 215 discussions on the recent advances in Kalman filtering approaches to spatiotemporal nonlinear 216 systems (Schiff and Sauer, 2008) and an understanding from group representation theory in controller 217 or observer design by obtaining a modal decomposition into decoupled controllable and 218 219 uncontrollable (observable and unobservable) subspaces (Whalen et al., 2015). Specifically, Schiff and Sauer (Schiff and Sauer, 2008) showed the feasibility of unscented Kalman filter (UKF) for 220 221 recursive estimation of system state for nonlinear systems, including unobserved variables and parameter tracking, in a spatiotemporal model of cortex where such a nonlinear system is controllable 222 using an adaptive feedback electrical field. Here, discretization of the whole-brain detailed 223 biophysical model of NVU (Jolivet et al., 2015), for example with Galerkin methods that are used 224 quite robustly in fluid dynamics, will be necessary where each discrete element corresponds to a 225 volume of tissue imaged as well as stimulated with the EEG-NIRS/tDCS unit (see Figure 1). As an 226 227 alternative to a fundamental NVU model (Jolivet et al., 2015) for the volume of tissue imaged and stimulated with EEG-NIRS/tDCS unit, we tried (Dutta et al., 2015) to find an empirical model where 228 we performed empirical mode decomposition (EMD) and the Hilbert spectrum (Huang et al., 1998) 229 to model the system dynamics and found a negative cross-correlation between one of the intrinsic 230 mode function (IMF) of the HbO2 time-series and log-transformed mean-power time-course of EEG 231 primarily within 0.5Hz-11.25Hz frequency band (i.e., one of the EEG IMFs). In principal accordance, 232 for whole-head monitoring, we propose independent component analysis (ICA) to transform multi-233 channel EEG-NIRS/tDCS unit imaging data to a spatially transformed "virtual channel" (i.e., a 234 spatial filter) (Jung et al., 2001). Then, the "virtual channel" activity (e.g., posterior alpha band 235 activity) can be subjected to EMD (i.e., a temporal filter) to reduce the dimension of the observable 236 dynamics (i.e., further observer model reduction) before developing the Kalman filter observer 237 (Schiff and Sauer, 2008) using the IMFs. For EEG-NIRS-based monitoring of NVU under 238 perturbation with tDCS, as shown in the Figure 2, the NVU dynamics is captured by the function F 239 and the IMF observations by the function W. The UKF approach should match the nonlinear IMF 240 dynamics up to the second order statistics where the feasibility remains to be tested experimentally in 241 242 future studies. Furthermore, it may be possible to use the Kalman observer to calculate proportional 243 control (see Figure 2) of the brain-state (e.g., cortical excitability (Jindal et al., 2015a)) with tDCS. Here, intelligent placement of EEG-NIRS sensors and tDCS effectors is necessary to ensure 244 245 observability and controllability (Whalen et al., 2015) where Whalen et al. (Whalen et al., 2015) suggested in general that more direct incoming connections into an observed node lead to higher 246 observability and more direct outgoing connections from a controlled node lead to higher 247 controllability. Furthermore, controllability may be enhanced with multi-modal non-invasive brain 248 stimulation (NIBS), e.g. with direct electrical stimulation (Pulgar, 2015) and photobiostimulation 249 (Gonzalez-Lima and Barrett, 2014), which needs to be investigated. 250

Towards such brain-state dependent tDCS, the challenges include the nature of observability and controllability in whole-brain complex NVU networks as well as the subtleties of the tDCS interaction with the whole-brain NVU (e.g., based on heterogeneous geometrical characteristics (Molaee-Ardekani et al., 2013)) that can also have multi-timescale cross-talk and resulting complex non-linear dynamics (Jolivet et al., 2015) where the spatiotemporal observability and controllability remains to be verified in future studies.

257



**Figure 2:** State modulation of the neurovascular unit (NVU) with transcranial direct current stimulation (tDCS) to facilitate a brain state, e.g., spatiotemporal alpha-rhythm state. F is the function to capture NVU system, W is the function to capture observations, ICA is independent component (IC) analysis is a linear decomposition method to transform electroencephalogram (EEG) - near infrared spectroscopy (NIRS) data collected at single scalp channels to a spatially transformed "virtual channel" (i.e., a spatial filter on multi-channel EEG-NIRS data), EMD is empirical model decomposition of the "virtual channel" observations that provide intrinsic mode functions (IMF) for proportional control (gain is G) of individual "virtual channel" activity or IC (e.g., posterior alpha band activity) with tDCS.

258

# 259 Acknowledgments

260 Research was conducted within the context of the regional NUMEV funding, Franco-German PHC-

261 PROCOPE 2014 funding, and Franco-Indian INRIA-DST funding. The help and advice received

262 from the German collaborator (Dr. med. Michael A. Nitsche), Indian collaborators (Dr. med. Abhijit

Das, Dr. Shubhajit Roy Chowdhury, and Dr. Dipanjan Roy), and the French collaborators (Dr.
Mitsuhiro Hayashibe, Dr. Stephane Perrey and Dr. Mark Muthalib) are gratefully acknowledged.

# 265 **References**

- Araque, A., Carmignoto, G., and Haydon, P. G. (2001). Dynamic signaling between astrocytes and
   neurons. *Annu. Rev. Physiol.* 63, 795–813. doi:10.1146/annurev.physiol.63.1.795.
- Ardolino, G., Bossi, B., Barbieri, S., and Priori, A. (2005). Non-synaptic mechanisms underlie the
   after-effects of cathodal transcutaneous direct current stimulation of the human brain. J.
   *Physiol.* 568, 653–663. doi:10.1113/jphysiol.2005.088310.
- Attwell, D., Buchan, A. M., Charpak, S., Lauritzen, M., Macvicar, B. A., and Newman, E. A. (2010).
  Glial and neuronal control of brain blood flow. *Nature* 468, 232–243.
  doi:10.1038/nature09613.
- Behzadi, Y., and Liu, T. T. (2005). An arteriolar compliance model of the cerebral blood flow
  response to neural stimulus. *NeuroImage* 25, 1100–1111.
  doi:10.1016/j.neuroimage.2004.12.057.
- Bozzo, L., Puyal, J., and Chatton, J.-Y. (2013). Lactate Modulates the Activity of Primary Cortical
   Neurons through a Receptor-Mediated Pathway. *PLoS ONE* 8, e71721.
   doi:10.1371/journal.pone.0071721.
- Buffo, A., Rite, I., Tripathi, P., Lepier, A., Colak, D., Horn, A.-P., Mori, T., and Götz, M. (2008).
   Origin and progeny of reactive gliosis: A source of multipotent cells in the injured brain.
   *Proc. Natl. Acad. Sci.* 105, 3581–3586. doi:10.1073/pnas.0709002105.
- Chander, B. S., and Chakravarthy, V. S. (2012). A Computational Model of Neuro-Glio-Vascular
   Loop Interactions. *PLoS One* 7, e48802. doi:10.1371/journal.pone.0048802.
- David, O., and Friston, K. J. (2003). A neural mass model for MEG/EEG: coupling and neuronal
   dynamics. *NeuroImage* 20, 1743–1755.
- DiNuzzo, M., Gili, T., Maraviglia, B., and Giove, F. (2011). Modeling the contribution of neuron astrocyte cross talk to slow blood oxygenation level-dependent signal oscillations. *J. Neurophysiol.* 106, 3010–3018. doi:10.1152/jn.00416.2011.
- Dmochowski, J. P., Datta, A., Bikson, M., Su, Y., and Parra, L. C. (2011). Optimized multi-electrode
   stimulation increases focality and intensity at target. *J. Neural Eng.* 8, 046011.
   doi:10.1088/1741-2560/8/4/046011.
- Dutta, A. (2014). EEG-NIRS based low-cost screening and monitoring of cerebral microvessels
   functionality. *International Stroke Conference 2014, At San Diego, Volume: Junior Investigator Session II: Invited Symposium.*
- Dutta, A., Chowdhury, S. R., Dutta, A., Sylaja, P. N., Guiraud, D., and Nitsche, M. (2013). A
   phenomological model for capturing cerebrovascular reactivity to anodal transcranial direct
   current stimulation. in *2013 6th International IEEE/EMBS Conference on Neural Engineering* (*NER*), 827–830. doi:10.1109/NER.2013.6696062.

300	Dutta, A., Jacob, A., Chowdhury, S. R., Das, A., and Nitsche, M. A. (2015). EEG-NIRS Based
301	Assessment of Neurovascular Coupling During Anodal Transcranial Direct Current
302	Stimulation - a Stroke Case Series. J. Med. Syst. 39, 205. doi:10.1007/s10916-015-0205-7.
303	Dutta, A., and Nitsche, M. (2013). Neural mass model analysis of online modulation of
304	electroencephalogram with transcranial direct current stimulation. in 2013 6th International
305	IEEE/EMBS Conference on Neural Engineering (NER), 206–210.
306	doi:10.1109/NER.2013.6695908.
307	Fricke, K., Seeber, A. A., Thirugnanasambandam, N., Paulus, W., Nitsche, M. A., and Rothwell, J. C.
308	(2011). Time course of the induction of homeostatic plasticity generated by repeated
309	transcranial direct current stimulation of the human motor cortex. <i>J. Neurophysiol.</i> 105, 1141–
310	1149. doi:10.1152/jn.00608.2009.
311	Friston, K. J., Mechelli, A., Turner, R., and Price, C. J. (2000). Nonlinear responses in fMRI: the
312	Balloon model, Volterra kernels, and other hemodynamics. <i>NeuroImage</i> 12, 466–477.
313	doi:10.1006/nimg.2000.0630.
314	Girouard, H., and Iadecola, C. (2006). Neurovascular coupling in the normal brain and in
315	hypertension, stroke, and Alzheimer disease. J. Appl. Physiol. Bethesda Md 1985 100, 328–
316	335. doi:10.1152/japplphysiol.00966.2005.
317 318	Gonzalez-Lima, F., and Barrett, D. W. (2014). Augmentation of cognitive brain functions with transcranial lasers. <i>Front. Syst. Neurosci.</i> 8. doi:10.3389/fnsys.2014.00036.
319	Gruetter, R., Novotny, E. J., Boulware, S. D., Rothman, D. L., and Shulman, R. G. (1996). 1H NMR
320	Studies of Glucose Transport in the Human Brain. J. Cereb. Blood Flow Metab. 16, 427–438.
321	doi:10.1097/00004647-199605000-00009.
322	Halnes, G., ?stby, I., Pettersen, K. H., Omholt, S. W., and Einevoll, G. T. (2013). Electrodiffusive
323	Model for Astrocytic and Neuronal Ion Concentration Dynamics. <i>PLoS Comput. Biol.</i> 9.
324	doi:10.1371/journal.pcbi.1003386.
325	Han, CH., Song, H., Kang, YG., Kim, BM., and Im, CH. (2014). Hemodynamic responses in rat
326	brain during transcranial direct current stimulation: a functional near-infrared spectroscopy
327	study. <i>Biomed. Opt. Express</i> 5, 1812–1821. doi:10.1364/BOE.5.001812.
328	Horvath, J. C., Carter, O., and Forte, J. D. (2014). Transcranial direct current stimulation: five
329	important issues we aren't discussing (but probably should be). <i>Front. Syst. Neurosci.</i> 8.
330	doi:10.3389/fnsys.2014.00002.
331	Huang, N. E., Shen, Z., Long, S. R., Wu, M. C., Shih, H. H., Zheng, Q., Yen, NC., Tung, C. C., and
332	Liu, H. H. (1998). The empirical mode decomposition and the Hilbert spectrum for nonlinear
333	and non-stationary time series analysis. <i>Proc. R. Soc. Lond. Ser. Math. Phys. Eng. Sci.</i> 454,
334	903–995. doi:10.1098/rspa.1998.0193.
335 336	Iadecola, C. (2004). Neurovascular regulation in the normal brain and in Alzheimer's disease. <i>Nat. Rev. Neurosci.</i> 5, 347–360. doi:10.1038/nrn1387.

- Islam, N., Aftabuddin, M., Moriwaki, A., Hattori, Y., and Hori, Y. (1995). Increase in the calcium
  level following anodal polarization in the rat brain. *Brain Res.* 684, 206–208.
- Jindal, U., Sood, M., Chowdhury, S. R., Das, A., Kondziella, D., and Dutta, A. (2015a). Corticospinal
   excitability changes to anodal tDCS elucidated with NIRS-EEG joint-imaging: an ischemic
   stroke study. in *ResearchGate* Available at:
- http://www.researchgate.net/publication/277800701\_Corticospinal\_excitability\_changes\_to\_a
   nodal tDCS elucidated with NIRS-EEG joint-imaging an ischemic stroke study
- 344 [Accessed July 8, 2015].
- Jindal, U., Sood, M., Dutta, A., and Chowdhury, S. R. (2015b). Development of Point of Care Testing
  Device for Neurovascular Coupling From Simultaneous Recording of EEG and NIRS During
  Anodal Transcranial Direct Current Stimulation. *IEEE J. Transl. Eng. Health Med.* 3, 1–12.
  doi:10.1109/JTEHM.2015.2389230.
- Jolivet, R., Coggan, J. S., Allaman, I., and Magistretti, P. J. (2015). Multi-timescale Modeling of
   Activity-Dependent Metabolic Coupling in the Neuron-Glia-Vasculature Ensemble. *PLoS Comput Biol* 11, e1004036. doi:10.1371/journal.pcbi.1004036.
- Jung, T.-P., Makeig, S., McKeown, M. J., Bell, A. J., Lee, T.-W., and Sejnowski, T. J. (2001).
   Imaging brain dynamics using independent component analysis. *Proc. IEEE* 89, 1107–1122.
   doi:10.1109/5.939827.
- Khadka, N., Truong, D. Q., and Bikson, M. (2015). Principles of Within Electrode Current Steering1.
   *J. Med. Devices* 9, 020947–020947. doi:10.1115/1.4030126.
- Krause, B., Márquez-Ruiz, J., and Cohen Kadosh, R. (2013). The effect of transcranial direct current
   stimulation: a role for cortical excitation/inhibition balance? *Front. Hum. Neurosci.* 7, 602.
   doi:10.3389/fnhum.2013.00602.
- Leontiev, O., and Buxton, R. B. (2007). Reproducibility of BOLD, perfusion, and CMRO2
   measurements with calibrated-BOLD fMRI. *NeuroImage* 35, 175–184.
   doi:10.1016/j.neuroimage.2006.10.044.
- Lucas, S. J. E., Tzeng, Y. C., Galvin, S. D., Thomas, K. N., Ogoh, S., and Ainslie, P. N. (2010).
   Influence of Changes in Blood Pressure on Cerebral Perfusion and Oxygenation.
   *Hypertension* 55, 698–705. doi:10.1161/HYPERTENSIONAHA.109.146290.
- Mangia, A. L., Pirini, M., and Cappello, A. (2014). Transcranial direct current stimulation and power
   spectral parameters: a tDCS/EEG co-registration study. *Front. Hum. Neurosci.* 8.
   doi:10.3389/fnhum.2014.00601.
- Markus, H., and Cullinane, M. (2001). Severely impaired cerebrovascular reactivity predicts stroke
   and TIA risk in patients with carotid artery stenosis and occlusion. *Brain* 124, 457–467.
   doi:10.1093/brain/124.3.457.
- Matsumoto, J., Fujiwara, T., Takahashi, O., Liu, M., Kimura, A., and Ushiba, J. (2010). Modulation
  of mu rhythm desynchronization during motor imagery by transcranial direct current
  stimulation. *J. Neuroengineering Rehabil.* 7, 27. doi:10.1186/1743-0003-7-27.

McKendrick, R., Parasuraman, R., and Ayaz, H. (2015). Wearable functional near infrared 375 spectroscopy (fNIRS) and transcranial direct current stimulation (tDCS): expanding vistas for 376 neurocognitive augmentation. Front. Syst. Neurosci., 27. doi:10.3389/fnsys.2015.00027. 377 Meinzer, M., Lindenberg, R., Darkow, R., Ulm, L., Copland, D., and Flöel, A. (2014). Transcranial 378 direct current stimulation and simultaneous functional magnetic resonance imaging. J. Vis. 379 *Exp. JoVE.* doi:10.3791/51730. 380 Molaee-Ardekani, B., Márquez-Ruiz, J., Merlet, I., Leal-Campanario, R., Gruart, A., Sánchez-381 Campusano, R., Birot, G., Ruffini, G., Delgado-García, J.-M., and Wendling, F. (2013). 382 Effects of transcranial Direct Current Stimulation (tDCS) on cortical activity: 383 A computational modeling study. Brain Stimulat. 6, 25–39. doi:10.1016/j.brs.2011.12.006. 384 385 Moore, C. I., and Cao, R. (2008). The hemo-neural hypothesis: on the role of blood flow in information processing. J. Neurophysiol. 99, 2035-2047. doi:10.1152/jn.01366.2006. 386 Moran, R. J., Kiebel, S. J., Stephan, K. E., Reilly, R. B., Daunizeau, J., and Friston, K. J. (2007). A 387 neural mass model of spectral responses in electrophysiology. NeuroImage 37, 706-720. 388 389 doi:10.1016/j.neuroimage.2007.05.032. Nikulin, V. V., Fedele, T., Mehnert, J., Lipp, A., Noack, C., Steinbrink, J., and Curio, G. (2014). 390 391 Monochromatic Ultra-Slow (~0.1Hz) Oscillations in the human electroencephalogram and their relation to hemodynamics. *NeuroImage*. doi:10.1016/j.neuroimage.2014.04.008. 392 393 Nitsche, M. A., Fricke, K., Henschke, U., Schlitterlau, A., Liebetanz, D., Lang, N., Henning, S., Tergau, F., and Paulus, W. (2003). Pharmacological modulation of cortical excitability shifts 394 induced by transcranial direct current stimulation in humans. J. Physiol. 553, 293-301. 395 396 doi:10.1113/jphysiol.2003.049916. 397 Nitsche, M. A., and Paulus, W. (2000). Excitability changes induced in the human motor cortex by weak transcranial direct current stimulation. J. Physiol. 527 Pt 3, 633-639. 398 399 Nitsche, M. A., and Paulus, W. (2011). Transcranial direct current stimulation--update 2011. Restor. Neurol. Neurosci. 29, 463–492. doi:10.3233/RNN-2011-0618. 400 Notturno, F., Marzetti, L., Pizzella, V., Uncini, A., and Zappasodi, F. (2014). Local and remote effects 401 of transcranial direct current stimulation on the electrical activity of the motor cortical 402 network. Hum. Brain Mapp. 35, 2220-2232. doi:10.1002/hbm.22322. 403 Novak, V. (2012). Cognition and Hemodynamics. Curr. Cardiovasc. Risk Rep. 6, 380-396. 404 doi:10.1007/s12170-012-0260-2. 405 Nunez, P. L., and Srinivasan, R. (2006). Electric Fields of the Brain: The Neurophysics of EEG. 406 407 Oxford University Press. Pellerin, L., and Magistretti, P. J. (1994). Glutamate uptake into astrocytes stimulates aerobic 408 glycolysis: a mechanism coupling neuronal activity to glucose utilization. Proc. Natl. Acad. 409 Sci. U. S. A. 91, 10625–10629. 410

- Polanía, R., Nitsche, M. A., and Paulus, W. (2011). Modulating functional connectivity patterns and
  topological functional organization of the human brain with transcranial direct current
  stimulation. *Hum. Brain Mapp.* 32, 1236–1249. doi:10.1002/hbm.21104.
- Polanía, R., Paulus, W., and Nitsche, M. A. (2012). Modulating cortico-striatal and thalamo-cortical
  functional connectivity with transcranial direct current stimulation. *Hum. Brain Mapp.* 33,
  2499–2508. doi:10.1002/hbm.21380.
- Pulgar, V. M. (2015). Direct electric stimulation to increase cerebrovascular function. *Front. Syst. Neurosci.* 9, 54. doi:10.3389/fnsys.2015.00054.
- Raffin, E., and Siebner, H. R. (2014). Transcranial brain stimulation to promote functional recovery
   after stroke. *Curr. Opin. Neurol.* 27, 54–60. doi:10.1097/WCO.0000000000059.
- Rahman, A., Reato, D., Arlotti, M., Gasca, F., Datta, A., Parra, L. C., and Bikson, M. (2013). Cellular
  effects of acute direct current stimulation: somatic and synaptic terminal effects. *J. Physiol.*591, 2563–2578. doi:10.1113/jphysiol.2012.247171.
- Schestatsky, P., Morales-Quezada, L., and Fregni, F. (2013). Simultaneous EEG Monitoring During
   Transcranial Direct Current Stimulation. J. Vis. Exp. JoVE. doi:10.3791/50426.
- Schiff, S. J., and Sauer, T. (2008). Kalman filter control of a model of spatiotemporal cortical dynamics. *J. Neural Eng.* 5, 1–8. doi:10.1088/1741-2560/5/1/001.
- Siesler, H. W., Ozaki, Y., Kawata, S., and Heise, H. M. (2008). *Near-Infrared Spectroscopy: Principles, Instruments, Applications.* John Wiley & Sons.
- Sigala, R., Haufe, S., Roy, D., Dinse, H. R., and Ritter, P. (2014). The role of alpha-rhythm states in perceptual learning: insights from experiments and computational models. *Front. Comput. Neurosci.* 8, 36. doi:10.3389/fncom.2014.00036.
- Sood, M., Jindal, U., Chowdhury, S. R., Das, A., Kondziella, D., and Dutta, A. (2015). Anterior
  temporal artery tap to identify systemic interference using short-separation NIRS
  measurements: a NIRS/EEG-tDCS study. in *ResearchGate* Available at:
  http://www.researchgate.net/publication/277710873\_Anterior\_temporal\_artery\_tap\_to\_identif
  y\_systemic\_interference\_using\_short-separation\_NIRS\_measurements\_a\_NIRSEEGtDCS\_study [Accessed July 8, 2015].
- Sotero, R. C., Trujillo-Barreto, N. J., Iturria-Medina, Y., Carbonell, F., and Jimenez, J. C. (2007).
  Realistically coupled neural mass models can generate EEG rhythms. *Neural Comput.* 19, 478–512. doi:10.1162/neco.2007.19.2.478.
- Spitoni, G. F., Cimmino, R. L., Bozzacchi, C., Pizzamiglio, L., and Di Russo, F. (2013). Modulation
  of spontaneous alpha brain rhythms using low-intensity transcranial direct-current
  stimulation. *Front. Hum. Neurosci.* 7, 529. doi:10.3389/fnhum.2013.00529.
- Suzuki, A., Stern, S. A., Bozdagi, O., Huntley, G. W., Walker, R. H., Magistretti, P. J., and Alberini, C.
   M. (2011). Astrocyte-neuron lactate transport is required for long-term memory formation.
   *Cell* 144, 810–823. doi:10.1016/j.cell.2011.02.018.

- Ursino, M., Cona, F., and Zavaglia, M. (2010). The generation of rhythms within a cortical region:
  analysis of a neural mass model. *NeuroImage* 52, 1080–1094.
  doi:10.1016/j.neuroimage.2009.12.084.
- Volterra, A., Liaudet, N., and Savtchouk, I. (2014). Astrocyte Ca<sup>2+</sup> signalling: an unexpected
   complexity. *Nat. Rev. Neurosci.* 15, 327–335. doi:10.1038/nrn3725.
- Whalen, A. J., Brennan, S. N., Sauer, T. D., and Schiff, S. J. (2015). Observability and Controllability
  of Nonlinear Networks: The Role of Symmetry. *Phys. Rev. X* 5, 011005.
  doi:10.1103/PhysRevX.5.011005.
- Yang, J., Ruchti, E., Petit, J.-M., Jourdain, P., Grenningloh, G., Allaman, I., and Magistretti, P. J.
  (2014). Lactate promotes plasticity gene expression by potentiating NMDA signaling in neurons. *Proc. Natl. Acad. Sci.* 111, 12228–12233. doi:10.1073/pnas.1322912111.
- Zaehle, T., Sandmann, P., Thorne, J. D., Jäncke, L., and Herrmann, C. S. (2011). Transcranial direct current stimulation of the prefrontal cortex modulates working memory performance:
  combined behavioural and electrophysiological evidence. *BMC Neurosci.* 12, 2.
  doi:10.1186/1471-2202-12-2.
- Zavaglia, M., Astolfi, L., Babiloni, F., and Ursino, M. (2006). A neural mass model for the simulation
   of cortical activity estimated from high resolution EEG during cognitive or motor tasks. *J. Neurosci. Methods* 157, 317–329. doi:10.1016/j.jneumeth.2006.04.022.
- Zheng, X., Alsop, D. C., and Schlaug, G. (2011). Effects of transcranial direct current stimulation
  (tDCS) on human regional cerebral blood flow. *NeuroImage* 58, 26–33.
  doi:10.1016/j.neuroimage.2011.06.018.

469

# 470 Figure legends

471 Figure 1: Illustration of the effects of transcranial direct current stimulation (tDCS) from simultaneous recording of electroencephalogram (EEG) and near infra red spectroscopy (NIRS). The 472 colors on the plot corresponds to the sensors. Here, only one anode and one cathode is highlighted for 473 474 tDCS, however, local current steering based on NIRS-EEG feedback is possible to optimally orient the electric field with multiple return electrodes (Khadka et al., 2015). NVU consists of the 475 476 endothelium, glia, neurons, pericytes, and the basal lamina in which neurons, astrocytes, and vessels are semi-independent networks operating in tandem. Neuronal network activity drives functional 477 hyperemia via direct effects on the blood vessels network as well as indirect effects via the astrocytic 478 479 network. Also, the hemodynamics changes can impact neuronal network activity via direct (diffusible messengers, electromechanical and thermal interactions) and indirect (via astrocytic network) 480 481 pathways.

482

Figure 2: State modulation of the neurovascular unit (NVU) with transcranial direct current 483 stimulation (tDCS) to facilitate a brain state, e.g., spatiotemporal alpha-rhythm state. F is the function 484 to capture NVU system, W is the function to capture observations, ICA is independent component 485 486 (IC) analysis is a linear decomposition method to transform electroencephalogram (EEG) - near infrared spectroscopy (NIRS) data collected at single scalp channels to a spatially transformed 487 "virtual channel" (i.e., a spatial filter on multi-channel EEG-NIRS data), EMD is empirical model 488 decomposition of the "virtual channel" observations that provide intrinsic mode functions (IMF) for 489 490 proportional control (gain is G) of individual "virtual channel" activity or IC (e.g., posterior alpha band activity) with tDCS. 491