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## Detection of volume loss in hippocampal layers in Alzheimer's disease using 7 T MRI: A feasibility study



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### ABSTRACT

In Alzheimer's disease (AD), the hippocampus is an early site of tau pathology and neurodegeneration. Histological studies have shown that lesions are not uniformly distributed within the hippocampus. Moreover, alterations of different hippocampal layers may reflect distinct pathological processes. 7 T MRI dramatically improves the visualization of hippocampal subregions and layers. In this study, we aimed to assess whether 7 T MRI can detect volumetric changes in hippocampal layers *in vivo* in patients with AD. We studied four AD patients and seven control subjects. MR images were acquired using a whole-body 7 T scanner with an eight channel transmit-receive coil. Hippocampal subregions were manually segmented from coronal T2\*-weighted gradient echo images with  $0.3 \times 0.3 \times 1.2 \text{ mm}^3$  resolution using a protocol that distinguishes between layers richer or poorer in neuronal bodies. Five subregions were segmented in the region of the hippocampal body: alveus, strata radiatum, lacunosum and moleculare (SRLM) of the cornu Ammonis (CA), hilum, stratum pyramidale of CA and stratum pyramidale of the subiculum. We found strong bilateral reductions in the SRLM of the cornu Ammonis and in the stratum pyramidale of the subiculum ( $p < 0.05$ ), with average cross-sectional area reductions ranging from  $-29\%$  to  $-49\%$ . These results show that it is possible to detect volume loss in distinct hippocampal layers using segmentation of 7 T MRI. 7 T MRI-based segmentation is a promising tool for AD research.

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### 1. Introduction

The hippocampal formation is an early site of Alzheimer's disease (AD) pathology (Braak and Braak, 1991). Hippocampal atrophy is a well-established imaging marker of Alzheimer's disease and has been included in recently proposed research diagnostic criteria (Albert et al., 2011; Dubois et al., 2007; Jack et al., 2011). Hippocampal atrophy can be measured from T1-weighted magnetic resonance imaging (MRI), using manual or automated segmentation (Colliot et al., 2008; Jack et al., 1992; Lehéricy et al., 1994) which provides an estimate of the

**Abbreviations:** AD, Alzheimer's disease; 7 T, 7 Tesla; MRI, magnetic resonance imaging; SRLM, strata radiatum, lacunosum and moleculare; CA, cornu Ammonis; DG, dentate gyrus; SP, stratum pyramidale; CN, cognitively normal.

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hippocampal volume. Hippocampal volume can discriminate AD patients from elderly controls with high sensitivity and specificity as demonstrated by numerous studies (Jack et al., 1992; Jack et al., 1997; Killiany et al., 1993; Laakso et al., 1996; Xu et al., 2000). Traditionally, hippocampal volumetry considers the hippocampus as a single anatomical entity and thus only provides a global volumetric measure. However, the hippocampal formation is an anatomically complex structure which is composed of different subregions (Duvernoy, 2005). It is made of two convoluted sheets of gray matter that are folded one onto another: the cornu Ammonis (CA) and the dentate gyrus (DG). The cornu Ammonis is further composed of four sectors: CA1–4. CA1 is the largest sector and is continuous to the subiculum. Furthermore, subregions are composed of different layers that vary in terms of cellular composition. The stratum pyramidale (SP) is richer in neuronal bodies while the strata radiatum, lacunosum and moleculare (SRLM) are poorer in neuronal bodies.

In Alzheimer's disease, histological studies have shown that lesions are not uniformly distributed within hippocampal subregions. Neurofibrillary tangles, neuronal loss and decrease in synaptic density are dominant in the first sector of the cornu Ammonis (CA1) and the subiculum, the dentate gyrus being relatively spared (Fukutani et al., 1995; Hyman et al., 1984; Rössler et al., 2002; Scheff et al., 2007; Simić et al., 1997; West et al., 1994). Neuronal loss results in a reduction of the thickness of the layers richer in neuronal bodies, while the loss of synapses results in the reduction of the layers poorer in neuronal bodies (Braak and Braak, 1997; Lace et al., 2009; Thal et al., 2000). Histological studies have suggested that synaptic loss in SRLM could occur earlier than neuronal loss in SP (Braak and Braak, 1997; Fukutani et al., 1995; Hyman et al., 1984; Rössler et al., 2002; Thal et al., 2000). *In vivo* measurement of hippocampal layers would thus be of major interest for AD research.

In the past years, several studies have analyzed volumes of hippocampal subregions in patients with AD and mild cognitive impairment (MCI), based on advances of high-field MRI (La Joie et al., 2013; Mueller et al., 2010; Pluta et al., 2012). Most studies were based on T2-weighted or proton-density weighted sequences with about 0.4 mm in plane coronal resolution and 2 mm slice thickness, performed at 3 T or 4.7 T. They have reported volume reductions in different subregions, including the subiculum (La Joie et al., 2013; Mueller et al., 2010), CA1 (La Joie et al., 2013; Mueller et al., 2010; Pluta et al., 2012) and the dentate gyrus (Pluta et al., 2012).

7 T MRI provides new contrasts, increased signal-to-noise ratio and higher spatial resolution which dramatically improve the visualization of hippocampal subregions (Theysohn et al., 2009). Using 7 T MRI, Kerchner et al. (2010) found atrophy of the strata radiatum, lacunosum and moleculare (the layers poorer in neuronal bodies) in AD patients. However, they used linear measurements of thickness at selected points and did not perform subregional segmentation. Wisse et al. (2012) have proposed a new manual segmentation protocol for hippocampal subregions at 7 T and subsequently applied this protocol to AD patients (Wisse et al., 2014). However, the protocol did not distinguish between layers richer and poorer in neuronal bodies. Chupin et al. (2009) and Henry et al. (2011) have performed segmentation of subregions at 7 T, using a protocol distinguishing between layers poorer and richer in neuronal bodies, in healthy subjects and patients with temporal lobe epilepsy.

The purpose of our study was to assess if 7 T MRI-based segmentation can detect volume loss of hippocampal layers in patients with AD compared to control subjects.

## 2. Materials and methods

### 2.1. Participants

The regional ethics committee approved this prospective study and written informed consent was obtained from all participants. We studied four AD patients (3 men, 1 woman; mean age  $\pm$  standard deviation [SD]:  $65.8 \pm 7.0$  years, range: 60–76 years, Mini Mental State Examination [MMSE] score:  $19.0 \pm 5.1$ , range: 13–25) and seven cognitively

normal (CN) participants (3 men and 4 women; age:  $63.1 \pm 5.6$  years, range: 55–72 years; MMSE score:  $29.1 \pm 1.1$ , range: 27–30). The two groups did not differ for age (Mann–Whitney U test,  $p = 0.5$ ) and gender (Chi-square test,  $\chi^2 = 0.25$ , 1df,  $p = 0.62$ ). AD patients were required to meet the criteria of the National Institute of Neurological Disorders and Stroke and Alzheimer's Disease and Related Disorders Association for mild to moderate AD (McKhann et al., 1984) with a CDR (Clinical Dementia Rating Scale) of 1.0 or more and an IADL score (Instrumental Activities of Daily Living) of more than 1 for each item or more than 2 for one item. The criteria for the inclusion of cognitively normal subjects were: the absence of memory complaint, normal scores for MMSE, normal scores for tests of executive functions and normal memory performance defined as a free recall score of 25/48 or more and a total recall score of more than 44/48 in the Free and Cued Selective Reminding Test (FCSRT). Subjects with recent cardiovascular disease (less than 1 year), neurological or psychiatric disease (except for AD) or focal lesion on brain MRI were excluded. Demographic and clinical characteristics of the participants are given in Table 1.

### 2.2. MRI acquisition

Imaging was performed on a whole-body 7 T scanner (Magnetom 7 T, Siemens Healthcare, Erlangen, Germany) with an eight channel transmit–receive coil. The acquisition protocol included a 3D T1-weighted magnetization prepared rapid gradient echo (MP-RAGE) acquisition covering the whole brain and a T2\*-weighted 2D gradient echo (GRE) coronal acquisition covering the hippocampal formation. The T2\*-weighted 2D gradient echo was composed of three acquisition slabs (Fig. 1). It was angulated perpendicular to the longitudinal axis of the hippocampus, as visualized on a parasagittal view of the 3D T1 sequence. Acquisition parameters of each sequence are presented in Table 2. Prior to the study, we performed several pilot acquisitions with different resolution and sequence parameters, with a specific study of the influence of GRE echo time (from about 10–45 ms) on contrast and visibility of the different layers and on the presence of artifacts in the structure of interest. We chose the two echo times that maximized contrast while keeping a reasonable acquisition time.

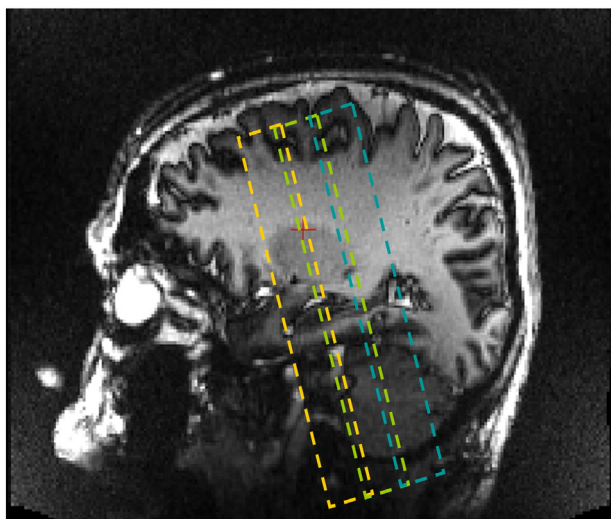
### 2.3. Segmentation of hippocampal subregions

Manual segmentation of subregions within the hippocampal body was performed by a trained rater (CB), blinded to clinical data and diagnosis. The rater (C.B.) is a neuroradiologist with 3 years of experience in neuroradiology at the time when the segmentations were carried out. She had previous experience of hippocampal segmentation and was specifically trained to the neuroanatomy of hippocampal subregions and their segmentation on MRI data. Segmentation was performed on the longer echo T2\*-weighted images of the second acquisition slab and external borders of hippocampal subregions were refined on the first echo. Each slice was visually inspected and the visibility of the

**Table 1**  
Demographic and clinical characteristics of the population.

| Subject no. | Group | Gender | Age | MMSE | CDR |
|-------------|-------|--------|-----|------|-----|
| 1           | CN    | F      | 68  | 30   | 0   |
| 2           | CN    | F      | 55  | 30   | 0   |
| 3           | CN    | F      | 72  | 29   | 0   |
| 4           | CN    | F      | 62  | 27   | 0   |
| 5           | CN    | M      | 61  | 29   | 0   |
| 6           | CN    | M      | 61  | 30   | 0   |
| 7           | CN    | M      | 63  | 29   | 0   |
| 8           | AD    | M      | 60  | 25   | 1   |
| 9           | AD    | M      | 76  | 21   | 2   |
| 10          | AD    | M      | 63  | 17   | 1   |
| 11          | AD    | M      | 64  | 13   | 2   |

CN = Cognitively normal subjects, AD = Alzheimer's disease patients. F = Female, M = male. Age is in years.



**Fig. 1.** Illustration of the positioning of the three acquisition slabs of the T2\*-weighted GRE sequence, based on a T1-weighted parasagittal view. The last posterior slice of the first acquisition slab was positioned on the last slice of the head of the hippocampus. The first acquisition slab is represented in orange, the second in green and the third in blue.

internal structure was judged sufficient to allow manual segmentation, not masked by the movement and artifacts nor by the loss of signal. Fig. 2 presents, in a single representative AD subject, the complete set of slices of the second slab used to define hippocampal layers. Segmentation was done using the object-based ROI module of the Anatomist software (<http://www.brainvisa.info/index.html>). The hippocampal head and tail were not segmented because the more complex shape of these structures posed difficulties in labeling procedures that were difficult to solve with the GRE images, susceptibility artifacts yielding more blurred borders in the head of the hippocampus.

Manual segmentation of hippocampal body subregions was performed using the following protocol, extending the one proposed by Henry et al. (2011). The hippocampal body was subdivided in five subregions based on the anatomy described by Duvernoy (2005). The following subregions were sequentially segmented. (1) Alveus. (2) SRLM corresponding to the strata radiatum, lacunosum and moleculare of CA1–3, the strata lacunosum and moleculare of the subiculum and the stratum moleculare of the dentate gyrus (DG). This corresponds to the layers poorer in neuronal bodies. (3) Hilum corresponding to the stratum pyramidale of CA4 and the stratum granulosum and polymorphic layer of DG, which are the layers richer in neuronal bodies. (4) Cornu Ammonis Stratum Pyramidale (CA\_SP), assumed to correspond to the stratum pyramidale of CA1–3, layers richer in neuronal bodies of CA1–3. (5) Subiculum\_SP corresponding to the stratum pyramidale of the subiculum, layers richer in neuronal bodies of the subiculum. The fimbria was not considered here, due to the difficulties in discriminating it from blood vessels on T2\* weighted images.

Anterior and posterior limits of the segmented region in the hippocampal body were defined on coronal sections, with reference mainly to sagittal sections to ensure 3D consistency. The first slice considered for the segmentation was chosen as the first slice posterior to the

uncus. The posterior slice of the segmentation was the last slice of the middle slab. The five subregions were then segmented as follows.

1. Alveus. The superior medial border of the alveus was set at the point where CA merges within the hilum; its inferior lateral border was at the junction with the collateral eminence of the lateral ventricle.
2. SRLM. The superior medial border of SRLM was straightforward to define as the point where CA revolved around it to enter the hilum; its inferior medial border corresponded to the point where the parahippocampal gyrus bulged upwards and changed slope.
3. Hilum. The superior medial border of the hilum was the fimbriodentate sulcus; its superior border was defined by drawing an imaginary line between the superior medial borders of the alveus and SRLM, and could be described as the end of the ribbon-like aspect of CA; its inferior lateral border was formed by SRLM; its medial border was defined by the cerebro-spinal fluid in the cisterna ambiens.
4. CA\_SP. CA\_SP was delineated between the alveus and SRLM. Its superior border was determined by the hilum; its inferior medial border was defined as follows: first by tracing the largest diameter of the hilum, then by tracing a line perpendicular to the CA axis passing by the medium of the hilum diameter. This perpendicular line was the infero-medial border of CA\_SP.
5. Subiculum\_SP. The medial limit corresponded to that of SRLM; the superior border was SRLM; its inferior border was the temporal white matter.

#### 2.4. Comparison to linear measurement of hippocampal subregions

For comparison with volumetric measures, we also performed manual linear measurements of thickness, as in Kerchner et al. (2010) for SRLM, CA\_SP and Subiculum\_SP. While Kerchner et al. (2010) measured SRLM and CA\_SP, we chose to also measure thickness of Subiculum\_SP for comparison with our volumetric measurements of this specific structure. Linear measurements were made by the same rater (CB), blind to manual segmentation, using Dicom Viewer 2.0 software (Rubo Medical Imaging, the Netherlands) on the longer echo T2\*-weighted images of the second acquisition slab. Linear measures were performed using the following procedure that was chosen as close as possible to that of Kerchner et al. (2010). Thickness was measured by drawing an orthogonal line between two tissue plane interfaces. Six linear measurements were made for each side and for each subregion, on two different slices at the level of red nucleus (three measurements on each slice for each subregion). We chose two slices separated from each other by 2.4 mm and not adjacent slices since the slice thickness (1.2 mm) of our acquisitions was different from that of Kerchner et al. (2010) (2 mm). This allows to have a distance between the two slices which is as close as possible to that of Kerchner et al. (2010). For each subregion and each side, we computed the average of the 6 linear measurements.

#### 2.5. Statistical analysis

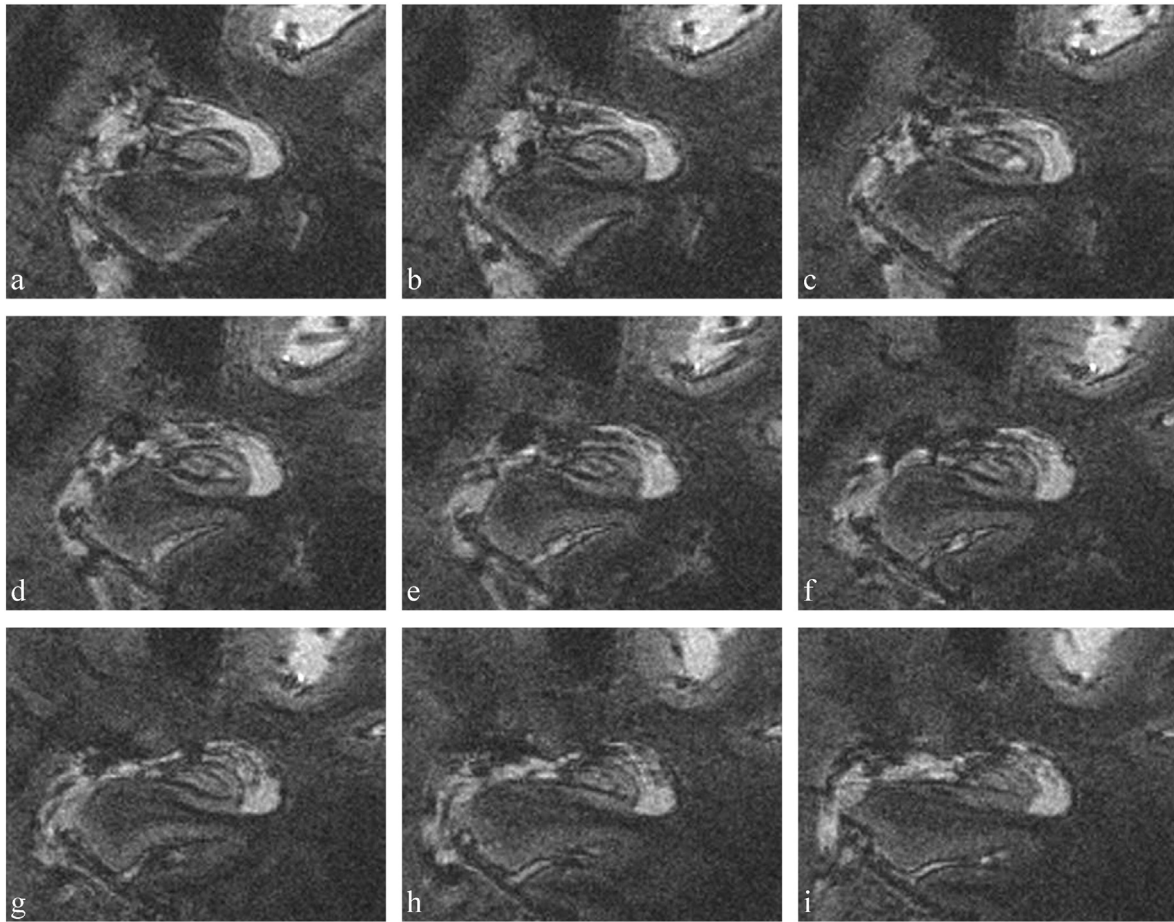
We normalized the volume of each subregion to the ipsilateral body length, resulting in measures of average cross-sectional area. Group differences in subregional areas between AD and CN subjects were assessed using Mann–Whitney U tests. Group differences in linear thickness measures were also assessed using Mann–Whitney U tests.

**Table 2**  
MRI acquisition protocol.

| Sequence   | Acquisition plane and coverage  | Number of slices | Field of view | Matrix    | Acquisition time | Flip angle | TR/TE/TI (ms)                | Resolution (mm) |
|------------|---------------------------------|------------------|---------------|-----------|------------------|------------|------------------------------|-----------------|
| MP-RAGE    | Sagittal - Head                 | 176              | 230 × 230     | 256 × 256 | 11 min           | 6°         | 2640/3.28/1100               | 0.9 × 0.9 × 0.9 |
| 2D T2* GRE | Coronal - Hippocampal formation | 3 × 15           | 173 × 100     | 576 × 576 | 3 × 7 min        | 65°        | 742/16.41 (TE1); 33.22 (TE2) | 0.3 × 0.3 × 1.2 |

MP-RAGE = Magnetization prepared rapid gradient echo; GRE = gradient recalled echo; TR/TE/TI = repetition time/echo time/inversion time.





**Fig. 2.** Coronal slices of a TE2 T2\*-weighted acquisition in a single AD patient: complete set of slices of the second slab used to define hippocampal layers, in an anterior-to-posterior direction from a to i.

Correlation between average cross-sectional area and linear measurements was assessed using Spearman's rank correlation coefficient. Significance threshold was set at  $p < 0.05$ . Statistical analyses were performed by C.B. using MedCalc for Windows, version 12.2.1 (MedCalc Software, Mariakerke, Belgium).

### 3. Results

Segmentation of hippocampal subregions was feasible in all participants. Examples of T2\*-weighted acquisitions and segmented hippocampal subregions are presented on Figs. 3 and 4.

#### 3.1. Volumetric measurement of hippocampal subregions

Average subregional cross-sectional areas are presented in Table 3. Individual measures are presented on Fig. 5. We found strong and significant ( $p < 0.05$ ) bilateral reductions of SRLM and Subiculum\_SP in AD patients compared to CN subjects. For left Subiculum\_SP, there was no overlap between patients and controls. The alveus area was significantly smaller on the left in AD patients. There was a trend towards reduction of left CA\_SP area ( $p = 0.058$ ).

#### 3.2. Comparison to linear measurement of hippocampal subregions

In CN subjects, left and right average linear measurements were respectively mean  $\pm$  standard deviation =  $0.69 \pm 0.08$  mm and  $0.64 \pm 0.09$  mm for SRLM,  $1.61 \pm 0.39$  mm and  $1.69 \pm 0.19$  mm for CA\_SP and  $1.76 \pm 0.5$  mm and  $1.74 \pm 0.23$  for Subiculum\_SP. In AD subjects, left and right average linear measurements were respectively  $0.45 \pm 0.04$  mm and  $0.49 \pm 0.08$  mm for SRLM,  $1.17 \pm$

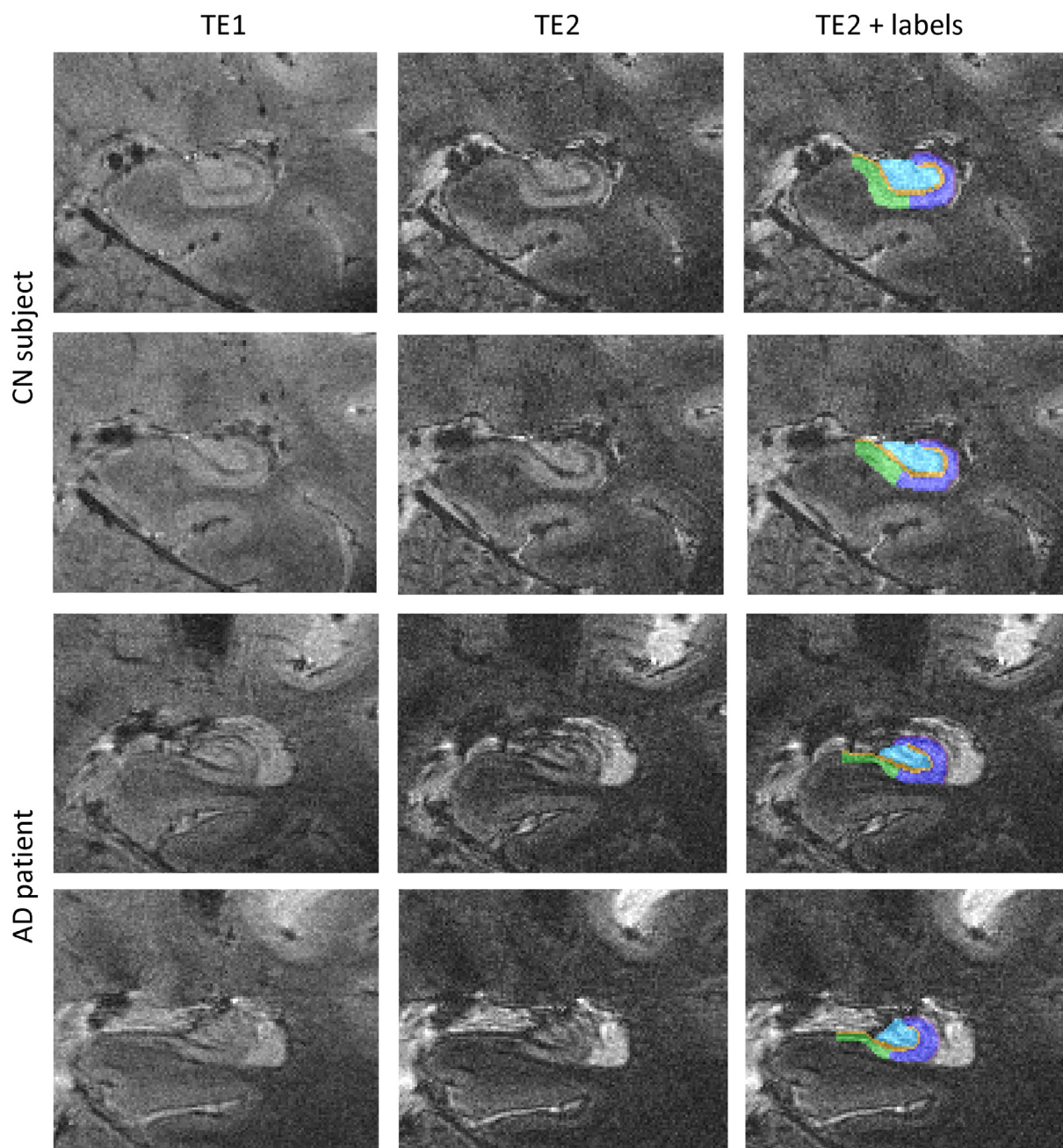
$0.12$  mm and  $1.29 \pm 0.29$  mm for CA\_SP and  $1.09 \pm 0.16$  mm and  $1.18 \pm 0.27$  for Subiculum\_SP. Bilaterally, SRLM and Subiculum\_SP were significantly thinner in AD patients compared to CN subjects ( $p < 0.05$ ). Left CA\_SP was also significantly thinner ( $p < 0.05$ ). Average cross-sectional areas of SRLM, CA\_SP and Subiculum\_SP correlated significantly ( $p < 0.05$ ) with linear measurements except for the right SRLM ( $p = 0.0710$ ).

### 4. Discussion

In this study, we demonstrated the feasibility of measuring volumetric alterations of the hippocampal inner structure in AD patients using ultra-high resolution MRI at 7 T. In particular, we were able to distinguish *in vivo* layers which are richer in neuronal bodies (stratum pyramidale – SP) from those which are poorer (strata radiatum, lacunosum and moleculare – SRLM).

We found strong significant bilateral atrophy of the SP of the subiculum and of SRLM. The largest reduction was found in the left subiculum, reaching an average of 49%. Subiculum atrophy has been previously reported in subregional segmentation studies at 4.7 T (Mueller et al., 2010) and 3 T (La Joie et al., 2013; Wisse et al., 2014). Very recently, a 7 T MRI study reported subiculum and CA atrophy in AD patients (Wisse et al., 2014). Consistently, histological studies report large numbers of neurofibrillary tangles and important neuronal loss in the subicular area (Hyman et al., 1984; Rössler et al., 2002; Simić et al., 1997).

We found a strong bilateral atrophy of SRLM. Dominant atrophy of SRLM is consistent with histological knowledge which described a primary degeneration of synapse-rich CA1 SRLM which precedes neuronal

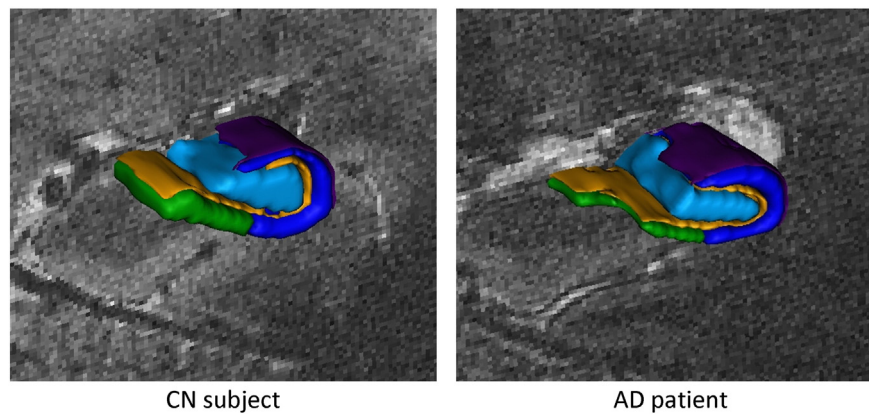


**Fig. 3.** Coronal slices of a T2-weighted acquisition (first echo TE1 and second echo TE2) in a CN subject and an AD patient at the level of the hippocampal body and corresponding segmentations. Segmentations were performed on the second echo image. Purple, alveus; dark blue, CA\_SP; yellow, SRLM; cyan, hilum and green, subiculum\_SP. Abbreviations: CA\_SP = stratum pyramidale of CA1–3; SRLM = strata radiatum, lacunosum and moleculare of CA1–3, strata lacunosum and moleculare of the subiculum and stratum moleculare of gyrus dentatus; hilum = stratum pyramidale of CA4 and stratum granulosum and polymorphic layer of gyrus dentatus; subiculum\_SP = stratum pyramidale of the subiculum. CN = cognitively normal, AD = Alzheimer's disease.

loss of CA1 SP (Braak and Braak, 1997; Fukutani et al., 1995; Hyman et al., 1984; Rössler et al., 2002; Thal et al., 2000). Synaptic loss in CA1 SRLM would be due to aggregates of microtubule-associated hyperphosphorylated tau-protein, and would appear earlier than pyramidal neuron loss, due to neurofibrillary tangles formation, localized in CA1 SP (Braak and Braak, 1997; Fukutani et al., 1995; Hyman et al., 1984; Rössler et al., 2002; Thal et al., 2000). To our knowledge, only Kerchner et al. (Kerchner et al., 2014; Kerchner et al., 2013; Kerchner et al., 2010) have previously measured SRLM *in vivo* in AD patients, initially on T2\*-weighted gradient echo acquisitions with line drawings (Kerchner et al., 2010), then on T2 Fast Spin Echo using a semi-automated edge-detection algorithm to integrate SRLM

width (Kerchner et al., 2014; Kerchner et al., 2013; Kerchner et al., 2012). They also found a significant atrophy of SRLM (Kerchner et al., 2010). Furthermore, SRLM atrophy was stronger in APOE  $\epsilon$ 4 carriers (Kerchner et al., 2014) and shared vulnerability with the entorhinal cortex (Kerchner et al., 2013). The in-plane resolution in our study is less high (0.3 mm) than that of Kerchner et al. (2010) (0.195 mm) and our AD patients are at a later disease stage (mean MMSE: 19 vs 23). However, our slice thickness is smaller (1.2 mm) than that of Kerchner et al. (2010) (2 mm) resulting in less partial volume effect and allowing a better delineation of the limits during the segmentation. To further assess the agreement between our results and those of Kerchner et al. (2010), we compared our





**Fig. 4.** Three-dimensional renderings of the segmentations. Purple, alveus; dark blue, CA\_SP; yellow, SRLM; cyan, hilum and green, subiculum\_SP. Abbreviations: CA\_SP = stratum pyramidale of CA1–3; SRLM = strata radiatum, lacunosum and moleculare of CA1–3, strata lacunosum and moleculare of the subiculum and stratum moleculare of gyrus dentatus; hilum = stratum pyramidale of CA4 and stratum granulosum and polymorphic layer of gyrus dentatus; subiculum\_SP = stratum pyramidale of the subiculum. CN = cognitively normal, AD = Alzheimer's disease.

volumetric measures to manual linear measurements. We found that volumetric and linear measures were in strong agreement.

We did not find a significant atrophy of the stratum pyramidale of CA, even though there was a trend towards reduction on the left. This result is also in line with Kerchner et al. (2010) which did not find atrophy of CA SP. On the other hand, several studies at lower field strength (3 T/4.7 T) have found atrophy of CA1 and/or CA1–2 transition in AD patients (La Joie et al., 2013; Mueller et al., 2010; Pluta et al., 2012). Several factors may explain this difference with our results: smaller number of patients in our study, the fact that studies at 3 T/4.7 T did not distinguish between SP and SRLM and the fact that we did not separate CA1 from other CA sectors. Separation of CA1 from other sectors is often based on geometrical criteria (e.g. Mueller et al., 2007; La Joie et al., 2010). We did not use this strategy, in order to minimize the number of arbitrary geometrical limits. Another option is to define the limit of CA1 based on the thinning of the stratum pyramidale, as in Kerchner et al. (2012). Indeed, according to Duvernoy (2005), the limit between CA1 and CA2 corresponds to a thinning of the stratum pyramidale and a thickening of the stratum moleculare. We assessed the possibility of identifying this limit (thinning of SP) in our data but this identification did not prove to be reliable, in particular in AD patients. For these reasons, we decided not to subsegment CA.

In the past years, substantial progresses have been made for *in vivo* measurement of hippocampal subregions in AD. The first studies of sub-regional hippocampal atrophy in AD have relied on local shape analysis of the external hippocampal boundary, based on various techniques

including deformation-based (Csernansky et al., 2000; Wang et al., 2006), radial mapping (Frisoni et al., 2008; Thompson et al., 2004), voxel-based (Chételat et al., 2008) and spherical harmonics (Gerardin et al., 2009) approaches. However, these approaches do not segment hippocampal subregions and the localization of the detected local atrophy with respect to different subfields relies on approximate mapping to a template. More recently, using advances of high-field MRI, several studies have shown that it is possible to detect atrophy of hippocampal subregions in AD and MCI patients based on T2-weighted or proton-density-weighted sequences at 3 T or 4.7 T (La Joie et al., 2013; Mueller et al., 2010; Pluta et al., 2012). 7 T MRI pushes even further the ability to measure subtle anatomical alterations of hippocampal subregions (Chupin et al., 2009; Henry et al., 2011; Kerchner et al., 2014; Kerchner et al., 2010; Thomas et al., 2008; Wisse et al., 2014; Wisse et al., 2012). In particular, it allows distinguishing between layers depending on their richness in neuronal bodies. In this work, we show that 7 T MRI reveals atrophy patterns in different subregions in AD patients.

Our study has the following limitations. First, only few patients were included, resulting in limited statistical power. It is thus possible that volume reductions in other subregions such as the hilum, alveus or CA\_SP would have been significant with a larger patient group. A related limitation is that we did not correct for multiple comparisons corresponding to the different segmented regions. However, one can note that most other studies (Wisse et al., 2014; Kerchner et al., 2010; Pluta et al., 2012; Mueller et al., 2010; La Joie et al., 2013) of hippocampal subfield volumetry did not correct for multiple comparisons either. Furthermore, a larger patient group is required to establish the sensitivity and specificity of these new measurements at the individual level. Another limitation is that we did not assess reproducibility of volumetric measures because the manual segmentation procedure is highly time consuming. These limitations underline the preliminary nature of our findings.

In conclusion, we were able to detect atrophy in distinct hippocampal layers in AD patients. 7 T MRI-based segmentation of hippocampal layers is a promising new technique for AD research and future investigations in that direction are worthwhile.

## Disclosure statement

Competing financial interests related to the present article: none to disclose for all authors. Competing financial interests unrelated to the present article. OC reports having received lecture fees from Lundbeck and consulting fees from Guerbet.

**Table 3**

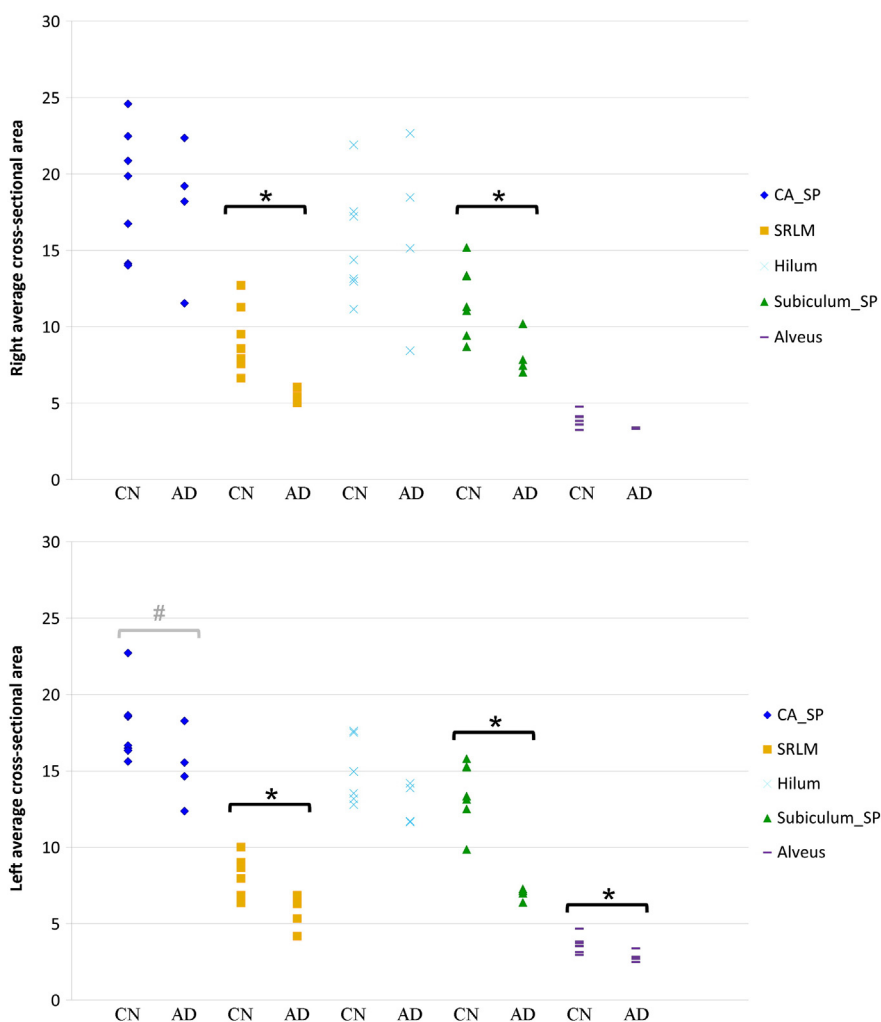
Mean average cross-sectional area of hippocampal body subregions (volume normalized to the ipsilateral hippocampal body length) (data are means  $\pm$  standard deviations, in mm<sup>2</sup>).

|              | AD (n = 4)            |                         | CN (n = 7)     |                |
|--------------|-----------------------|-------------------------|----------------|----------------|
|              | Right                 | Left                    | Right          | Left           |
| CA_SP        | 17.8 $\pm$ 4.6 (–6%)  | 15.2 $\pm$ 2.4** (–15%) | 18.9 $\pm$ 4.1 | 17.9 $\pm$ 2.4 |
| SRLM         | 5.6 $\pm$ 0.5* (–39%) | 5.7 $\pm$ 1.2* (–29%)   | 9.2 $\pm$ 2.2  | 7.9 $\pm$ 1.3  |
| Hilum        | 16.2 $\pm$ 6.0 (–4%)  | 12.9 $\pm$ 1.4 (–14%)   | 15.5 $\pm$ 3.7 | 14.9 $\pm$ 2.0 |
| Subiculum_SP | 8.1 $\pm$ 1.4* (–31%) | 6.9 $\pm$ 0.4* (–49%)   | 11.8 $\pm$ 2.3 | 13.6 $\pm$ 2.1 |
| Alveus       | 3.4 $\pm$ 0.04 (–13%) | 2.8 $\pm$ 0.4* (–21%)   | 3.8 $\pm$ 0.5  | 3.6 $\pm$ 0.6  |

CA\_SP = stratum pyramidale of CA1–3; SRLM = strata radiatum, lacunosum and moleculare of CA1–3, strata lacunosum and moleculare of the subiculum and stratum moleculare of gyrus dentatus; Hilum = stratum pyramidale of CA4 and stratum granulosum and polymorphic layer of gyrus dentatus; Subiculum\_SP = stratum pyramidale of the subiculum.

\* Indicates a significant difference between AD and CN (Mann–Whitney U test,  $p < 0.05$ ).

\*\* Indicates a trend which did not reach the significance threshold (Mann–Whitney U test,  $p = 0.058$ ).



**Fig. 5.** Right and left hippocampal subregional average cross-sectional area in CN and AD subjects (in mm<sup>2</sup>). Abbreviations: CA\_SP = stratum pyramidale of CA1–3; SRLM = strata radiatum, lacunosum and moleculare of CA1–3; strata lacunosum and moleculare of the subiculum and stratum moleculare of gyrus dentatus; hilum = stratum pyramidale of CA4 and stratum granulosum and polymorphic layer of gyrus dentatus; subiculum\_SP = stratum pyramidale of the subiculum. CN = cognitively Normal subjects, AD = Alzheimer's disease patients. \*Indicates a significant difference between AD and CN (Mann–Whitney U test,  $p < 0.05$ ). #Indicates a trend which did not reach the significance threshold (Mann–Whitney U test,  $p = 0.058$ ).

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